

SEARCHING AND PARASITISM OF *Diatraea saccharalis* (LEPIDOPTERA: CRAMBIDAE) BY *Trichospilus diatraeae* (HYMENOPTERA: EULOPHIDAE)

Búsqueda y parasitismo de *Diatraea saccharalis* (Lepidoptera: Crambidae) por *Trichospilus diatraeae* (Hymenoptera: Eulophidae)

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ABSTRACT

The ability of *Trichospilus diatraeae* Cherian and Margabandhu, 1942 (Hymenoptera: Eulophidae) to search and parasitize *Diatraea saccharalis* (Fabricius, 1794) (Lepidoptera: Crambidae) pupae in sugarcane stalks was evaluated. To analyze the ability for search and parasitism were used stalks of sugarcane (20 cm) where it was introduced a pupa of *D. saccharalis* (T1); a pupa and a caterpillar (T2) or a pupa and fecal matter (T3). Each stalk was placed in a transparent plastic bottle with 21 females of *T. diatraeae*. These pupae were isolated, after 72 h, in glass tubes at 25 ± 2 °C, 70 ± 10 % relative humidity, 14:10 light/dark. The experiment was developed in an entirely casualized design with three treatments and 12 repetitions. Percentage of *D. saccharalis* pupa parasitized by *T. diatraeae* was 50 %, 83.33 % and 16.66 % in the T1, T2 and T3, respectively ($\chi^2 = 3.896$, $p = 0.04$). The presence of *D. saccharalis* caterpillars favored searching and parasitism of this host.

Keywords: biological control, host location, parasitoid.

RESUMEN

La capacidad de *Trichospilus diatraeae* Cherian y Margabandhu, 1942 (Hymenoptera: Eulophidae) para buscar y parasitar las pupas de *Diatraea saccharalis* (Fabricius, 1794) (Lepidoptera: Crambidae) en los tallos de la caña de azúcar fue estudiada. Para analizar la habilidad de búsqueda y parasitismo fueron utilizados tallos de la caña de azúcar (20 cm) donde se introdujo una pupa de *D. saccharalis* (T1); pupas y orugas (T2) o pupa y residuos fecales (T3). Cada tallo fue colocado en una botella plástica transparente con 21 hembras de *T. diatraeae*. Esas pupas fueron individualizadas, luego de 72 h, en tubos de vidrio a 25 ± 1 °C, 70 ± 10 % UR y 14 h de foto período. El experimento se desarrolló en un diseño completamente al azar, con tres tratamientos y 12 repeticiones. Los porcentajes de pupas de *D. saccharalis* parasitadas por *T. diatraeae* fueron del 50 %, 83,33 % y 16,66 % en el T1, T2 y T3, respectivamente ($\chi^2 = 3.896$, $p = 0,04$). La presencia de *D. saccharalis* en los tallos de caña de azúcar favorecieron la búsqueda y parasitismo de su hospedero.

Palabras clave: control biológico, localización hospedero, parasitoide.

RESUMO

A capacidade de *Trichospilus diatraeae* Cherian e Margabandhu, 1942 (Hymenoptera: Eulophidae) para procurar e parasitar pupas de *Diatraea saccharalis* (Fabricius, 1794) (Lepidoptera: Crambidae) em colmos de cana-de-açúcar foi avaliada. Para analisar a capacidade de busca e parasitismo foram usados colmos de cana-de-açúcar (20 cm), onde foi introduzida uma pupa de *D. saccharalis* (T1); uma pupa e uma lagarta (T2) or uma pupa e resíduo fecal (T3). Cada colmo foi colocado em uma garrafa plástica transparente com 21 fêmeas de *T. diatraeae*. Essas pupas foram retiradas dos colmos após 72 h, e colocadas em tubos de vidro a 25 ± 2 ° C, 70 ± 10 % de umidade relativa e fotofase de 14 h. O experimento foi desenvolvido em delineamento inteiramente casualizado, com três tratamentos e 12 repetições. A porcentagem de pupas de *D. saccharalis* parasitadas por *T. diatraeae* foi de 50 %, 83,33 % e 16,66 % em T1, T2 e T3, respectivamente ($\chi^2 = 3,896$, $p = 0,04$). A presença de lagartas de *D. saccharalis* favoreceu a busca e parasitismo deste hospedeiro.

Palavras-chave: controle biológico, localização de hospedeiro, parasitoide.

INTRODUCTION

Adult female parasitoids respond to semiochemicals in the micro-habitat to find host pupae (Pinto *et al.*, 2007; Fontana *et al.*, 2011). These compounds are released from plants infested by herbivores and/or from fecal matter of these insects which are detected by parasitoid receptors (Fatouros *et al.*, 2007; Dicke *et al.*, 2009; Girling *et al.*, 2011; Hegde *et al.*, 2011). The efficiency of parasitoids depends on its ability to detect hosts in the field natural conditions (Silva-Torres *et al.*, 2009), although it may be reduced in absence of hosts at right developmental stages for parasitism (Hausmann *et al.*, 2005). The capacity parasitoids to find endophytic pupa is important because they usually remain hidden which reduces emission of detectable volatiles (Bruinsma *et al.*, 2009). Caterpillars may build shelters as protection that reduce its parasitism (Rodvalho *et al.*, 2007).

Females of the parasitoid *Brachymeria intermedia* (Nees, 1834) (Hymenoptera: Chalcididae) can identify kairomones of *Lymantria dispar* (Linnaeus, 1758) (Lepidoptera: Lymantriidae) after being exposed to pupa of this host (Cardé and Lee, 1989). This indicates that searching-behaviors may depend on acquired experience of parasitoids (Hoballah and Turlings, 2005; Peñaflor *et al.*, 2011). Contact with host before releasing may improve searching efficiency of laboratory reared parasitoids released in the field (González *et al.*, 2011). *Trichospilus diatraeae* Cherian and Margabandhu, 1942 (Hymenoptera: Eulophidae) parasitizes insect pupa, mainly those of lepidopterans, and present potentially for biological control of agricultural and forest pests (Boucek, 1976; Zaché *et al.*, 2010; Melo *et al.*, 2011). The ability of this insect to find and parasitize *D. saccharalis* pupae inside sugarcane stalks

are related with semiochemicals (Krugner *et al.*, 2008), vibrations (Fischer *et al.*, 2003), and/or chromatic and achromatic cues (Fischer *et al.*, 2004).

Diatraea saccharalis is an important pest of sugarcane, because the intensity of the attack and cause losses in sugar and alcohol productivity (Segato *et al.*, 2006; Dinardo-Miranda *et al.*, 2012). Controlling the sugarcane borer with chemicals is difficult, because this insect develops in a protected location, inside the stem of the plant (Vacari *et al.*, 2012). Therefore, biological control with parasitoids has been the most used way to combat this insect, so it is important to simulate the natural conditions before performing releases of *T. diatraeae* in the plantations of sugarcane, to check efficiency of parasitism in the field.

The present work evaluated capacity of *T. diatraeae* to search and parasitize *D. saccharalis* pupae in the presence of final instar caterpillars or fecal matter of this host.

MATERIALS AND METHODS

Rearing *D. saccharalis*

Diatraea saccharalis pupae were supplied by Empresa Agentes Biológicos BUG. This insect was reared with the following methodology: recently hatched caterpillars were maintained in glass tubes (8.5 x 2.5 cm) sealed with cotton and fed an artificial diet until pupa stage. Pupae were collected, sexed, and 20 males and 30 females were placed together for oviposition in PVC cages (22 x 10 cm) lined with sheets of sulfite paper humidified with distilled water. This PVC cages were sealed with a voile-type fabric and elastic (Parra, 2007).

Rearing *T. diatraeae*

Trichospilus diatraeae adults were maintained in glass tubes (14 x 2.2 cm) sealed with cotton and fed with drops of pure honey. *D. saccharalis* pupae with 24 to 48 hours old were subsequently introduced into these tubes during 24 hours. Pupae were removed and individualized in glass tubes (14 x 2.2 cm) in a chamber with controlled conditions chamber (25 ± 2 °C, 70 ± 10 % relative humidity, under a 14:10 light/dark) until emergence (Pereira *et al.*, 2008).

Experimental Design

Sugarcane stalks were cut in 20 cm long segments. An orifice was made in each segment and a 24 hour-old *D. saccharalis* pupa (184.00 ± 0.01 mg) was placed in each cavity. After the fixed inside each cavity and the stalk segments placed individually in transparent plastic tubes (25 x 9 cm) with 21 *T. diatraeae* females for 72 hours (Chichera *et al.*, 2012). Pupae were then removed and individualized in glass tubes (14 x 2.2 cm) in a chamber with controlled conditions (25 ± 2 °C, 70 ± 10 % relative humidity, 14:10 light/dark) to evaluate parasitism and to observe the emergence of *T. diatraeae*. Controls consisted of *D. saccharalis* pupae fixed into sugarcane stalk segments under identical conditions, but without *T. diatraeae* females.

Treatments consisted of exposing the parasitoid to: a pupa

(T1); a pupa and a caterpillar (T2) or a pupa and the fecal matter (T3) of *D. saccharalis*. Treatment T2 had two holes in the stalks (one at each internode) with one pupa in one hole and a fourth or fifth instar caterpillar in the other one. Treatment T3 had one pupa in an orifice into the stalk with fecal matter of *D. saccharalis* around its external area.

Each parcel was exposed to 21 *T. diatraeae* females, with 12 replications in an entirely randomized design. It was evaluated the percentage of parasitism [(number of pupae of *D. saccharalis* with emergence of parasitoid + pupae without adult emergence of *D. saccharalis*) / (total number of pupae) × 100]; the emergence percentage [(number of pupae of *D. saccharalis* to adult emergence of parasitoids) / (number of parasitized pupae) × 100] and progeny per pupa (number of parasitoid emerged per pupa of *D. saccharalis*). The successful of search of the parasitoid was measured by the percentage of parasitism of pupae.

Percentages of *T. diatraeae* parasitism were analyzed with a general linear model with binomial distribution ($p \leq 0.05$) using the R Statistical System software package (Ihaka and Gentleman, 1996). This analysis was performed using the original non-parametric data in percentages to facilitate interpretation. Data of emergence of *T. diatraeae* progeny were submitted to an analysis of variance with the test F at 5 % significance.

RESULTS

The present of final instar caterpillars or fecal matter of *D. saccharalis* affected the search and percentage of parasitism of females *T. diatraeae*. Percentage of *D. saccharalis* pupa parasitized by *T. diatraeae* was 50 %, 83.33 % and 16.66 % in the T1, T2 and T3, respectively ($\chi^2 = 3.896$, $p = 0.04$) (Fig. 1). The

presence of *D. saccharalis* caterpillars increased its searching and parasitism of *T. diatraeae*.

Emergence of *T. diatraeae* adults from *D. saccharalis* pupa was similar between treatments with pupa (66 %) or pupa and caterpillars (70 %) ($p > 0.05$). This parasitoid did not emerged in the treatment with pupa and fecal matter of *D. saccharalis*.

Progeny of *T. diatraeae* from *D. saccharalis* pupae was similar between treatments with pupae and caterpillars or only pupae (Table 1) of this host.

DISCUSSION

The ability of *T. diatraeae* to find *D. saccharalis* pupae in the orifices in sugarcane stalks this species is important for biological control (Chichera *et al.*, 2012). High parasitism levels of *D. saccharalis* pupae indicated potential of *T. diatraeae* as a biological control agent of this lepidopteran.

The parasitism of *T. diatraeae* in pupae of *D. saccharalis* introduced in the stalks of sugarcane was 50 %. Chichera *et al.* (2012) also noted 56 % parasitism in the experiment where the pupae of *D. saccharalis* were introduced in stems and exposed to *T. diatraeae*. A total of 83.33 % of *D. saccharalis* pupae was parasitized in the presence of caterpillars that host, this indicates that *T. diatraeae* was stimulated in the presence of larvae.

Higher ability of *T. diatraeae* to search *D. saccharalis* pupae with host caterpillars present suggests that it can recognize substances released by them and that they were close to pupation. As *Cotesia kariyai* (Watanabe, 1937) (Hymenoptera: Braconidae) females can distinguish chemical from damage by *Pseudaletia separata* (Walker, 1865) (Lepidoptera: Noctuidae) caterpillars up to fourth instar, but not between

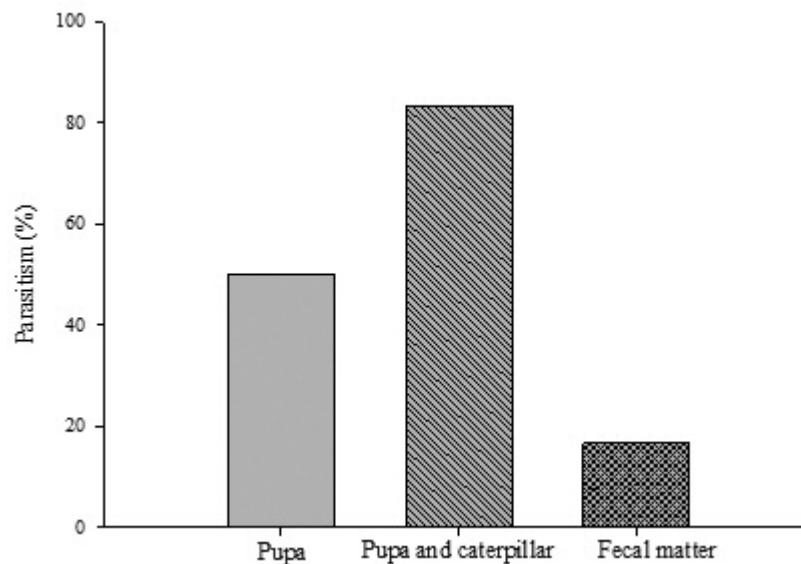


Figure 1. Percentage of parasitized pupae of *Diatraea saccharalis* (Lepidoptera: Crambidae) in sugarcane stalks by *Trichospilus diatraeae* (Hymenoptera: Eulophidae). Treatments represented by: pupa, pupa and caterpillar or pupa and fecal matter. 25 ± 2 °C, RH: 70 ± 10 % and 14 hours ($\chi^2 = 3.896$, $p = 0.04$).

Table 1. Progeny (mean \pm standard error) of *Trichospilus diatraeae* (Hymenoptera: Eulophidae) on pupae of *Diatraea saccharalis* (Lepidoptera: Crambidae) in sugarcane stalks. Treatments: pupa, pupa and a caterpillar or pupa and fecal matter 25 ± 2 ° C, RH: 70 ± 10 % and 14 hours.

Treatments	Progeny
Pupa	188.00 \pm 55.50 a
Pupa and caterpillar	231.57 \pm 45.64 a
Pupa fecal matter	-

Means followed by same letter do not differ by the F test ($p > 0.05$).

(-) No progeny was obtained.

later ones (Takabayashi *et al.*, 1995) – when they are not appropriate for parasitism.

The emergence of the progeny of *T. diatraeae* indicates that this parasitoid can search, parasitized and develop within *D. saccharalis* pupae in the orifices in sugarcane stalks – again demonstrating its potential for biological control (Keasar and Steinberg, 2008; Chichera *et al.*, 2012). The emergence of progeny favors establishment of this parasitoid in plantations (Bellows *et al.*, 2006) and may reduce the numbers of re-introductions and costs of producing and releasing this wasp (Gichini *et al.*, 2008).

Lower parasitism of *D. saccharalis* pupae by *T. diatraeae* with host fecal matter may be due to odors released these residues. Additionally, this fecal matter used in the experiment came from *D. saccharalis* caterpillars reared on artificial diet, and may have different composition in fecal matter of caterpillars fed on sugarcane plants in the natural environment. The searching behavior of the parasitoid *Cotesia flavipes* (Cameron, 1891) (Hymenoptera: Braconidae) is mediated by a water-soluble substance from the fecal matter of *D. saccharalis* caterpillars. The contact with this substance induces searching-behavior characterized by reduced locomotion and tapping the feces with its antenna (Van Leerdam *et al.*, 1985). On the other hand, semiochemicals may not be important for *Spathius agrili* Yang, 2005 (Hymenoptera: Braconidae) to find *Agrius planipennis* Fairmaire, 1888 (Coleoptera: Buprestidae) by this parasitoid females rely on host-generated vibrations to find suitable hosts (Wang *et al.*, 2010).

Pupae and caterpillars of *D. saccharalis* together in sugarcane stalks favored parasitism of this pest by *T. diatraeae* by simulating naturally infested plants. Volatile compounds released from the sugarcane stalks due to *D. saccharalis* caterpillar-feeding may facilitated searching of pupae of this host by this natural enemy. Cabbage plants infested by *Plutella xylostella* (Linnaeus, 1758) (Lepidoptera: Plutellidae) (Girling *et al.*, 2011) stimulated searching behavior of females of the larval parasitoid *Cotesia vestalis* (Haliday, 1834) (Hymenoptera: Braconidae). The parasitoid *Trichogramma pretiosum* Riley, 1879 (Hymenoptera: Trichogrammatidae) was attracted by volatile compounds released by corn plants damaged by caterpillars of *Elasmopalpus lignosellus* (Zeller, 1848) (Lepidoptera: Pyralidae) (Xavier *et al.*, 2011).

The presence of *D. saccharalis* caterpillars in sugarcane stalks increased its searching and parasitism by *T. diatraeae* – which could be due to the perception and identification of substances released by the caterpillars or to odors from damaged sugarcane stalks. On the other hand, fecal matter from *D. saccharalis* reduced parasitism, which could be attributed to chemicals such as not identified by *T. diatraeae*. The origin and compositions of chemical substances that help this parasitoid to search host pupae need to be investigated to understand the efficiency of *T. diatraeae* to control *D. saccharalis* in the field.

In summary, *T. diatraeae* females searched and parasitized *D. saccharalis* pupae in sugarcane stalks in the laboratory. The presence of *D. saccharalis* caterpillars favored searching and increased parasitism of pupae of this pest in sugarcane stalks. Fecal matter from *D. saccharalis* caterpillars reduced parasitism by *T. diatraeae*.

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BIBLIOGRAPHY

- Bellows JTS, Paine TD, Bezark LG, Ball J. Optimizing natural enemy release rates, and associated pest population decline rates, *Encarsia inaron* Walker (Hymenoptera: Aphelinidae) and *Siphoninus phillyreae* (Haliday) (Homoptera: Aleyrodidae). *Biol Control*. 2006; 37(1):25-31.
- Bruinsma M, Posthumus MA, Mumm R, Mueller MJ, Van Loon JJA, Dicke M. Jasmonic acid-induced volatiles of *Brassica oleracea* attract parasitoids: effects of time and dose, and comparison with induction by herbivores. *J Exp Bot*. 2009;60(9):2575-2587.
- Boucek Z. The African and Asiatic species of *Trichospilus* and *Cotterellia* (Hymenoptera, Eulophidae). *B Entomol Res*. 1976;65(4):669-681.
- Cardé RT, Lee H. Effect of experience on the responses of the parasitoid *Brachymeria intermedia* (Hymenoptera: Chalcididae) to its host, *Lymantria dispar* (Lepidoptera: Lymantriidae), and kairomone. *Ann Entomol Soc Am*.

- 1989;82(5):653-657.
- Chichera RA, Pereira FF, Kassab O, Barbosa RH, Pastori PL, Rossoni C. Capacidad de búsqueda y reproducción de *Trichospilus diatraeae* y *Palmistichus elaeisis* (Hymenoptera: Eulophidae) en pupas de *Diatraea saccharalis* (Lepidoptera: Crambidae). *Interciencia*. 2012;37(11):852-856.
- Dicke M, Van Lon JJA, Soler R. Chemical complexity of volatiles from plants induced by multiple attacks. *Nat Chem Biol*. 2009;5(5):317-324.
- Dinardo Miranda LL, Anjos IA, Costa VP, Fracasso JV. Resistance of sugarcane cultivars to *Diatraea saccharalis*. *Pesq Agropec Bras*. 2012;47(1):1-7.
- Fatouros NE, Kiss BG, Dicke M, Hilker M. The response specificity of *Trichogramma* egg parasitoids towards infochemicals during host location. *J Insect Behav*. 2007;20(1):53-65.
- Fischer S, Samietz EJ, Dorn ES. Efficiency of vibrational sounding in parasitoid host location depends on substrate density. *J Comp Physiol*. 2003;189(10):723-730.
- Fischer S, Samietz J, Wackers FL, Dorn S. Perception of chromatic cues during host location by the pupal parasitoid *Pimpla turionellae* (L.) (Hymenoptera: Ichneumonidae). *Environ Entomol*. 2004;33(1):81-87.
- Fontana A, Held M, Fantaye CA. Attractiveness of constitutive and herbivore-induced sesquiterpene blends of maize to the parasitic wasp *Cotesia marginiventris* (Cresson). *J Chem Ecol*. 2011;37(6):582-591.
- Gichini G, Löhr B, Rossbash A, Nyambo B, Gathu R. Can low releases numbers lead to establishment and spread of an exotic parasitoid, *Diadegma semiclausum* (Hellén), in East Africa. *Crop Prot*. 2008;27(6):906-914.
- Girling RB, Stewart-Jones A, Dherbecourt JT, Wright DJ, Poppy GM. Parasitoids select plants more heavily infested with their caterpillar hosts: a new approach to aid interpretation of plant headspace volatiles. *Proc R Soc*. 2011;278(1718):2646-2653.
- González JM, Cusumano A, Williams HJ, Colazza S, Vinson SB. Behavioral and chemical investigations of contact kairomones released by the mud Dauber Wasp *Trypoxylon politum*, a host of the parasitoid *Melittobia digitata*. *J Chem Ecol*. 2011;37(6):629-639.
- Hausmann C, Mattiacci L, Dorn S. Role of host feeding niches and host refuges in habitat-related behaviour of *Hyssopus pallidus* (Hymenoptera: Eulophidae), a larval parasitoid of the codling moth. *B Entomol Res*. 2005;95(5):429-436.
- Hegde M, Oliveira JN, Costa JG, Bleicher E, Santana AEG, Bruce TJA, et al. Identification of semiochemicals released by cotton, *Gossypium hirsutum*, upon infestation by the cotton aphid, *Aphis gossypii*. *J Chem Ecol*. 2011;37(7):741-750.
- Hoballah ME, Turlings TCJ. The role of fresh versus old leaf damage in the attraction of parasitic wasps to herbivore-induced maize volatiles. *J Chem Ecol*. 2005;31(9):2003-2018.
- Ihaka R, Gentleman R. R: a language for data analysis and graphics. *J Comput Graph Stat*. 1996;5(3):299-314.
- Keasar T, Steinberg S. Evaluation of the parasitoid *Copidosoma koehleri* for biological control of the potato tuber moth, *Phthorimaea operculella*, in Israeli potato fields. *Biocontrol Sci Technol*. 2008;18(4):325-336.
- Krugner R, Johnson MW, Daane KM, Morse JG. Olfactory responses of the egg parasitoid, *Gonatocerus ashmeadi* Girault (Hymenoptera: Mymaridae), to host plants infested by *Homalodisca vitripennis* (Germar) (Hemiptera: Cicadellidae). *Biol Control*. 2008;47(1):8-15.
- Melo RL, Pratisoli D, Polanczyk RA, Tavares M, Milanez AM, Melo DF. Ocorrência de *Trichospilus diatraeae* (Hym.: Eulophidae) em broca-das-cucurbitáceas, no Brasil. *Hortic Bras*. 2011;29(2):228-230.
- Parra JRP. Técnicas de Criação de Insetos para Programa de Controle Biológico, sexta ed. Piracicaba: ESALQ/FEALQ; 2007.
- Peñaflor MFGV, Erb M, Miranda, LA, Werneburg AG, Bent JMS. Herbivore-induced plant volatiles can serve as host location cues for a generalist and a specialist egg parasitoid. *J Chem Ecol*. 2011;37(12):1304-1313.
- Pereira FF, Zanoncio JC, Tavares MT, Pastori PL, Jacques GC, Vilela EF. New record of *Trichospilus diatraeae* as a parasitoid of the eucalypt defoliator *Thyriniteina arnobia* in Brazil. *Phytoparasitica*. 2008;36(3):304-306.
- Pinto DM, Nerg AM, Holopainen JK. The role of ozone-reactive compounds, terpenes, and green leaf volatiles (GLVs), in the orientation of *Cotesia plutellae*. *J Chem Ecol*. 2007;33(12):2218-2228.
- Rodvalho SR, Laumann RA, Diniz IV. Ecological aspects of lepidopteran caterpillar parasitoids from *Caryocar brasiliense* Camb. (Caryocaraceae) in a cerrado sensu stricto of Central Brazil. *Biota Neotrop*. 2007;7(3):239-243.
- Segato SV, Pinto AS, Jediroba E, Nóbrega JCM, organizadores. Atualização em produção de cana-de-açúcar. Piracicaba: CP2; 2006.
- Silva-Torres CSA, Barros R, Torres JB. Efeito da idade, foto-período e disponibilidade de hospedeiro no comportamento de *Oomyzus sokolowskii* Kurdjumov (Hymenoptera: Eulophidae). *Neotrop Entomol*. 2009;38(4):512-519.
- Takabayashi J, Takahashi S, Dicke M, Posthumus MA. Developmental stage of herbivore *Pseudaletia separata* affects production of herbivore-induced synomone by corn plants. *J Chem Ecol*. 1995;21(3):273-287.
- Vacari AM, De Bortoli SA, Borba DF, Martins MIEG. Quality of *Cotesia flavipes* (Hymenoptera: Braconidae) reared at different host densities and the estimated cost of its commercial production. *Biol Control*. 2012;63(2):102-106.
- Van Leerdam MBJ, Smith JW, Fuchs TW. Frass-mediated host-finding behavior of *Cotesia flavipes*, a braconid parasite of *Diatraea saccharalis* (Lepidoptera: Pyralidae).

- Ann Entomol Soc Am. 1985;78(5):646-650.
- Wang XY, Yang ZQ, Gould JR, Wu H, Ma JH. Host-seeking behavior and parasitism by *Spathius agrili* Yang (Hymenoptera: Braconidae), a parasitoid of the emerald ash borer. Biol Control. 2010;52(1):24-29.
- Xavier LMS, Laumann RA, Borges M, Magalhães DM, Vilela EF, Blassioli-Moraes MC. *Trichogramma pretiosum* attraction due to the *Elasmopalpus lignosellus* damage in maize. Pesq Agropec Bras. 2011;46(6):578-585.
- Zaché B, Wilcken CF, Dacosta RR, Soliman EP. *Trichospilus diatraeae* Cherian & Margabandhu, 1942 (Hymenoptera: Eulophidae), a new parasitoid of *Melanolophia consimilaria* (Lepidoptera: Geometridae). Phytoparasitica. 2010;38(4): 355-357.