

GENETIC DIVERSITY OF *Lutzomyia (Nyssomyia) intermedia* IN AN ENDEMIC AREA OF AMERICAN CUTANEOUS LEISHMANIASIS, STATE OF PARANÁ, BRAZIL

Diversidad genética de *Lutzomyia (Nyssomyia) intermedia* en un área endémica de leishmaniasis cutánea americana del estado de Paraná, Brasil

André Luiz GONÇALVES^{1,2}, Edilene Alcântara de CASTRO², Ennio LUZ^{2,†}, Ricardo Cancio FENDRICH¹, Nataly Araújo de SOUZA³, Vanete THOMAZ-SOCCOL^{1,*}

¹. Departamento de Engenharia de Bioprocessos e Biotecnologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil

². Programa de Pós-graduação em Microbiologia, Parasitologia e Patologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil

³. Laboratório Interdisciplinar de Vigilância Entomológica em Díptera e Hemiptera, Fundação Oswaldo Cruz, Rio de Janeiro, Rio de Janeiro, Brazil

† In memoriam

* For correspondence: vanetesoccol@gmail.com

Received: 18th January 2020. Returned for revision: 5th August 2020. Accepted: 2nd November 2020.

Associate Editor: Rafael Gutiérrez López

Citation/ citar este artículo como: Gonçalves AL, Castro EA, Luz E, Fendrich RC, Souza NA, Thomaz-Soccol V. Genetic diversity of *Lutzomyia (Nyssomyia) intermedia* in an endemic area of american cutaneous leishmaniasis, state of Paraná, Brasil. Acta Biol Colomb. 2021;26(3):365-373. Doi: <https://doi.org/10.15446/abc.v26n3.84619>

ABSTRACT

Lutzomyia intermedia (Diptera: Psychodidae) features as one of the main vectors that are involved in the transmission of American cutaneous leishmaniasis (ACL) in the Neotropical region. However, genetic studies involving this taxon are still incipient and important for understanding the level of variability of different populations, their role, and implications as vectors. The aim of this study was to determine the level of genetic diversity of *L. intermedia* present in the Ribeira River Valley, an area of ACL transmission in the state of Paraná, Brazil, through the Random amplified polymorphic DNA (RAPD). Two municipalities were chosen to collect sand flies: Cerro Azul (new transmission area of the ACL) and Adrianópolis (endemic area of the ACL). The insects were captured in the house, in the peridomicile and in the wild (forest). Two of the used markers made it possible to estimate the polymorphism of the studied populations, resulting in 40 genotypes, most of them from peridomicile. The dendrogram generated by the analysis with the primer A10 showed different degrees of similarity, suggesting that there may be gene flow in the studied populations. The Principal Coordinate Analysis (PCO) with the A2 primer, was useful in grouping *L. intermedia* according to its ecological and geographical origin. There was no distinction between the lineages composing the *L. intermedia* complex. The results of this study, with the record of great genotypic diversity in *L. intermedia*, may contribute to explain the maintenance of the life cycle of *Leishmania braziliensis* (Kinetoplastida: Trypanosomatidae) in the region.

Keywords: intraspecific variation, molecular epidemiology, phenetic analysis, Phlebotominae.

RESUMEN

Lutzomyia intermedia (Diptera: Psychodidae) es uno de los principales vectores que participan en la transmisión de leishmaniasis cutánea americana (LCA) en la región Neotropical. A pesar de que aún los estudios genéticos que involucran a este taxón son incipientes, tienen una gran importancia para comprender el nivel de variabilidad de las diferentes poblaciones y sus implicaciones en su papel vectorial. El objetivo de este estudio fue determinar el nivel de diversidad genética de *L. intermedia* presente en el Valle del Río Ribeira, área de transmisión de LCA en el estado de Paraná, Brasil, mediante RAPD (ADN polimórfico amplificado aleatoriamente). Los flebotomos fueron recolectados en los municipios Cerro Azul (nueva área de transmisión de LCA) y Adrianópolis (área endémica de LCA), donde fueron capturados en ambientes residenciales, en el peridomicilio y en el bosque. Dos de los marcadores utilizados permitieron estimar el polimorfismo en las poblaciones estudiadas con la obtención de 40 genotipos, la mayoría de ellos en el peridomicilio. El dendrograma generado por el análisis con el cebador A10 mostró diferentes grados de similitud, lo que sugiere

que puede haber flujo génico en las poblaciones. El Análisis de Coordenadas Principales (PCO) con el cebador A2 fue útil para agrupar *L. intermedia* según su origen ecológico y geográfico. No hubo distinción entre los linajes que componen el complejo *L. intermedia*. Los resultados de este estudio, con el registro de gran diversidad genotípica en *L. intermedia*, pueden contribuir a explicar el mantenimiento del ciclo biológico de *Leishmania braziliensis* (Kinetoplastida: Trypanosomatidae) en la región.

Palabras Clave: análisis fenético, epidemiología molecular, Phlebotominae, variación intraespecífica.

INTRODUCTION

Considered one of the biggest public health problems in the world, the leishmaniasis is caused by protozoa of the genus *Leishmania* Ross, 1903. One of the clinical spectra present in the Americas is the cutaneous form (ACL), endemic in Brazil (Savoia, 2015). In the South Region of Brazil, most human cases occurred in the state of Paraná, with 4582 reported cases between 2007 and 2018, according to data from the Ministry of Health of Brazil. One of the surroundings in which autochthonous transmission of ACL occurs in the state of Paraná corresponds to the Ribeira de Iguape River Valley, where for over a century the disease has been registered along with report of the species *Leishmania (Viannia) braziliensis* Vianna, 1911 that was isolated from patients (Castro *et al.*, 2005).

Ribeira River Valley is a region heavily influenced by anthropogenic changes. The Phlebotominae sand fly populations have changed their profile in relation to previous research over recent decades, and *Lutzomyia (Nyssomyia) intermedia* (Lutz and Neiva, 1912) have been currently found in abundance and considered the vector of *Leishmania* in domestic, peridomestic and sylvatic environments (Forattini *et al.*, 1976; Castro *et al.*, 2005; Gonçalves *et al.*, 2019).

Lutzomyia intermedia is widely distributed in Brazil, from the North and Northeast to the South. It is also present in northern Argentina, Paraguay and southern part of Bolivia (Andrade Filho *et al.*, 2007; Salomón *et al.*, 2016). Adopting morphological criteria, *Lutzomyia (Nyssomyia) neivai* (Pinto, 1926) was redescribed by Marcondes (1996) as a different species of *L. intermedia*. The author adopted the relationship between genital and extragenital structures to distinguish males of *L. intermedia* and *L. neivai*, such as ejaculatory ducts in males and the spermathacae and cibarial teeth of females.

Besides morphometric data, Marcondes (1997) has also studied mitochondrial DNA sequences and observed two phylogenetic lineages of *L. intermedia*: *L. intermedia s.s.* and *L. neivai*. Studies evaluating cryptic speciation, and the level of genetic structuring of populations of Phlebotominae in endemic areas, are epidemiologically relevant since they show how these species or their lineages differ in significance as vectors of *Leishmania* (Azevedo *et al.*, 2000; Bejarano *et al.*, 2009; Freitas *et al.*, 2018). Molecular data may contain enough variations to assist in these matters, revealing variations between populations from different geographic areas that are not distinguishable by morphology and those that constitute species complexes (Chan-Chable *et al.*, 2019; Lozano-Sardaneta *et al.*, 2020).

The RAPD technique can be used to determine the level of genetic variability in sand flies and clarify doubts regarding the current species complexes, which are difficult to distinguish solely by morphological features. This technique has been used to investigate the genetic characteristics of insect vectors including the taxonomic relationships between cryptic species (Adamson *et al.*, 1993; Fraga *et al.*, 2011), intra- and interpopulational variability (Meneses *et al.*, 2005; Rocha *et al.*, 2007; Seblova *et al.*, 2013) and the identification of populations resistant or susceptible to insecticides (Hiragi *et al.*, 2009) or phytochemicals (Sharma *et al.*, 2017).

Considering the possibilities of *Leishmania* transmission in different ecotypes in the Ribeira Valley and considering the high-density of *L. intermedia* in these environments, we raised the hypothesis that the vector role in the region is influenced by the degree of genetic structure of the populations of this sand fly. Therefore, may *L. intermedia* show enough variation to maintain different parasite transmission cycles? If these variations exist, may support occurrence of gene flow between populations of different ecotypes, areas or lineages? And this aforementioned variation, may also support the differentiation between *L. intermedia s.s.* and *L. neivai*, or among populations of different ecotypes or areas?

In this study, using RAPD markers, we analyzed the genetic variability of *L. intermedia* in three environments (house, peridomestic and wild), in two areas of Ribeira River Valley in the state of Paraná: the municipalities of Cerro Azul (a new transmission area) and Adrianópolis (an endemic area), with 164 and 118 cases of ACL reported between 2007 and 2018, respectively. We seek to assess the relationship among the domestic, peridomestic and sylvatic parasite transmission cycles. These markers were used to assess the level of divergence between the populations that comprise the taxon *L. intermedia*: *L. intermedia s.s.* and *L. neivai*, that are present in the region.

Thus, this study contributes to a better understanding about genetic variations of sand flies, detected by the technique of RAPD, to disclose the possible roles of these vectors in the epidemiology of ACL in the Ribeira de Iguape River Valley.

MATERIALS AND METHODS

Sampling area and sand fly captures and identification

Sand flies were captured in Cerro Azul (24°49'08"S, 49°15'37"W and 324 m.a.s.l.) and Adrianópolis (24°39'42"S,

48°59'30"W and 181 m. a. s. l.). These municipalities belong to the state of Paraná, southern Brazil, in the Ribeira de Iguape River basin (Fig. 1). Originally formed by covers of dense ombrophilous and mixed ombrophilous forest, this region is currently occupied by secondary forest, planted forest and by agricultural and extractive activities, as well as by the growth of extractive and transformation industries (IPARDES, 2021). Another anthropogenic influence in the region is the passage of the Bolivia-Brazil gas pipeline (1998-2009).

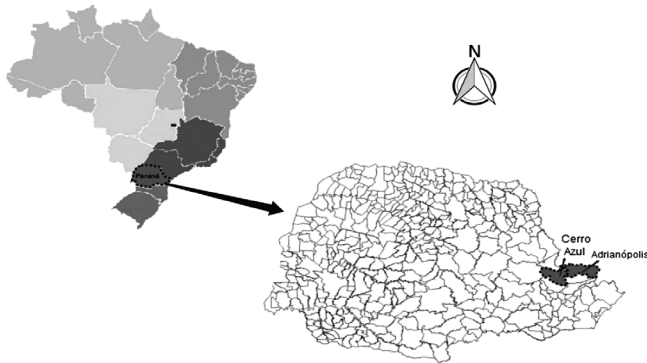


Figure 1. Geographical localization of the sand fly captures: municipalities of Cerro Azul and Adrianópolis, State of Paraná, Brazil

For the capture of sand flies, CDC-type light traps (Sudia and Chamberlain, 1962) were installed in areas with the occurrence of ACL transmission, in different ecotypes: inside the house, in the peridomicile and in the wild (forest). The samples were collected during the months of December 2008 and February, May and November 2009, between 7 p.m. and 12 a.m. (midnight).

For identification, the last abdominal segments of sand flies were clarified in 20 % potassium hydroxide solution. After 24 h, the sand flies were observed through an optical microscope and identified according to characteristics following the key proposed by Young and Duncan (1994). Marcondes' (1996) criterion was adopted to distinguish between the two lineages of *L. intermedia*; females with more than 11 spermathecae rings were identified as belonging to *L. intermedia* s.s., while the remainder were identified as *L. intermedia* s.l., together with all the males.

Molecular/RAPD analysis

Samples were subjected to molecular analysis by RAPD-PCR (Polymerase chain reaction), selected from 52 individuals (45 males and five females of *L. intermedia* s.l. and two females of *L. intermedia* s.s.) out of a total of 403 previously collected specimens (Gonçalves *et al.*, 2019).

The DNA of the samples was extracted as proposed by Loxdale and Lushai (1998). Briefly, samples were triturated in 300 μ L of 5 % Chelex100 within sterilized 1.5 mL microtubes with plastic pistils. Then, they were stirred strongly (for 15 s) and spun (19 500 \times g) for 20 s. After that,

they were incubated in a water bath (80 °C) for 30 min and then shaken again and centrifuged (19 500 \times g/20 s). The supernatant containing the DNA was transferred to the new tubes and stored at -20 °C.

Then, the DNA were amplified by RAPD-PCR, using the following primers: A10 (5'-GTGATCGCAG-3'), A2 (5'-TGCCGAGCTG-3'), A3 (5'-AGTCAGCCAC-3') and A9 (5'-GGGTAACGCC-3') (Tibayrenc *et al.*, 1993). The reaction was performed in a final volume of 25 μ L containing 1.5 μ L of DNA (1.64 η g), 16.8 μ L of ultra-pure sterilized water, 2.5 μ L of buffer (10X), 2.5 μ L of primer (5 μ M), 0.75 μ L of MgCl₂ (50 mM), 0.5 μ L of dNTP (10 mM) and 0.45 μ L of *Taq* DNA-polymerase (Invitrogen, Carlsbad, CA, USA) (5 U/ μ L).

Amplification was performed using a MJ Research® thermocycler (Watertown, MA, USA), with an initial denaturation at 94 °C for five min, followed by amplification with 45 cycles of denaturation (94 °C, one minute), annealing (36 °C, one minute) and polymerization (72 °C, two minutes), and a final extension at 72 °C for seven minutes. After amplifying the fragments by RAPD-PCR, the products obtained were submitted to separation according to molecular weight by horizontal electrophoresis on 1.5 % agarose gel in 1X TBE buffer (Tris-base, boric acid and EDTA pH 8.3) at 60 Volts for three hours. For size standards, we used the one kb and 100 bp Plus DNA Ladder Gibco markers.

The gels were stained with ethidium bromide (0.5 mg/mL) and visualized under ultraviolet light (TFX-35M Gibco BRL UV Transilluminator). In addition, we photographed and then analyzed the gels.

Statistical/Data analysis

The amplification profiles were analyzed in groups of ten samples of the same type of ecotype for each marker (A10 and A2). Thus, for the phenetic analysis, representative samples of each observed polymorphism were chosen. Based on the patterns obtained following RAPD, the Jaccard distance (D_{ij}) was used to create data matrices, considering the presence or absence of bands (one or zero, respectively) (Jaccard, 1908). Each band of the RAPD gel was encoded with a number starting with one for the slowest band (which contained a larger DNA fragment); thus, each genotype is represented by a set of numbers for each primer. This coefficient is represented by $D_{ij} = 1 - [C/(2N - C)]$, where C is the number of common bands between genotypes *i* and *j*, and N is the total number of bands in both genotypes. The units were grouped by the UPGMA (Unweighted pair-group method with arithmetical average) method, which is a hierarchical clustering model that allows the construction of dendrograms (Sneath and Sokal, 1973). The Mantel test of correlation of matrices (Mantel, 1967) was performed to observe the significance of the correlation between the genetic similarity matrix and the cophenetic matrix obtained from the data presented by the dendrograms. PCO

was performed to test for possible clustering in the genetic similarity matrix. The dendrograms, Mantel's test and PCO were obtained using the program NTSYS pc 2.1 (Rohlf, 2000). The strength of the nodes in the dendrogram was evaluated by bootstrap analysis using the Bood 3.03 program (Coelho, 2005) with 10 000 permutations.

RESULTS

Of the four primers selected for this study, two (A10 and A2) generated genetic profiles that allowed us to identify the level of polymorphism in populations of *L. intermedia s.l.* In this study, the amplification profiles obtained using primers A3 and A9 have shown none of the polymorphism visualized in products amplified with the use of primers A10 and A2. Therefore, the samples of sand fly DNA amplified with these primers have not been submitted to phenetic analysis. However, the use of primers A3 and A9 generated characteristic fragments with molecular weights between 400 and 500 bp. Overall, 40 genotypes were identified within the population of the lineage *L. intermedia s.l.* studied in 50 sand flies, collected from three ecotypes in two municipalities (Table 1). The peridomestic ecotype presented the highest genotypic diversity.

Table 1. Total number of genotypes obtained by RAPD of *Lutzomyia intermedia* from the three different ecotypes in the municipalities of Cerro Azul and Adrianópolis.

	Ecotypes		
	House	Peridomestic	Wild
Number of genotypes	8	20	12
Number of samples analyzed	10	20	20

The five *L. intermedia s.l.* females that were subjected to the RAPD technique, did not present relationship determined between the genotypes and the number of rings in the spermatheca of the females. Two females captured from the wild area in Cerro Azul had the same genotype, but one female had eight rings in the spermatheca, while the other presented nine rings. By contrast, three other females, also from the wild area of the same municipality, presented ten, nine, and seven rings respectively, in each spermatheca, and each showed different genetic profiles. In this way, the primers used in this research did not generate genotypes that maintained a relationship with morphological markers in *L. intermedia s.l.*

The genetic diversity of the population of *L. intermedia s.l.* studied was revealed not only by the number of genotypes identified, but also by analyzing the dendrogram generated by the matrix constructed from the profiles of DNA bands amplified with the A10 primer. The Mantel test showed r values (correlation between matrices) > 0.7 and p (significance test of r) < 0.01 .

The dendrogram showed different degrees of similarity (30 to 80 %), according to the Jaccard coefficient for the same

ecotype (Fig. 2). This suggests intraspecific polymorphism and the possibility of genetic flow among the studied populations.

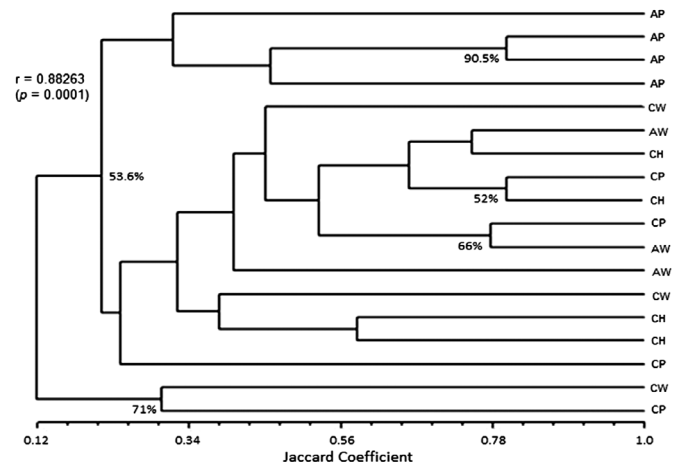


Figure 2. Dendrogram construction from data obtained by electrophoresis on 1.5% agarose gel, performed with primer A10 RAPD-PCR products. The specimens of *Lutzomyia intermedia* were captured in the municipalities of Cerro Azul and Adrianópolis. The dendrogram was generated by the program NTSYS-pc 2.1 with clustering by the UPGMA method and the Jaccard coefficient distance. Legend: house environment (CH), peridomestic (CP) and wild area (CW) of Cerro Azul and peridomestic (AP) and wild area (AW) of Adrianópolis. Bootstrap values (%) of each group are shown.

The PCO from the RAPD with the primer A2 clustered the sand flies in relation to their environment and municipality of collection (Fig. 3). Analyzing the graph, the samples from Cerro Azul were grouped together, and the samples from Adrianópolis formed another grouping. One specimen female classed as *L. intermedia s.s.* lineage, collected in the wild in Adrianópolis, was grouped with the other sand flies from this municipality (*L. intermedia s.l.*).

DISCUSSION

Molecular markers, such as RAPD, have been used in several studies of sand flies in the New and Old World, including taxonomic identification and population genetics (Golczer and Arrivillaga, 2015).

Mukhopadhyay *et al.* (2000) have used RAPD to find species-specific DNA profiles of closely related and morphologically similar sand flies, *Phlebotomus (Phlebotomus) papatasi* (Scopoli, 1786) and *Phlebotomus (Phlebotomus) duboscqi* Neveu-Lemaire, 1906. One of the tested primers has generated amplifications with differential profiles between the two species, with a characteristic band of 700 bp present in *P. papatasi*, and one of 490 bp present in *P. duboscqi*. Regarding the present study, the fragments generated by primers A3 and A9 may constitute a species-specific, genus-specific or even ecotype-specific marker. Likewise, Adamson *et al.* (1993) have also identified a 320 bp characteristic band present in *Lutzomyia youngi* Feliciangeli and Murillo,

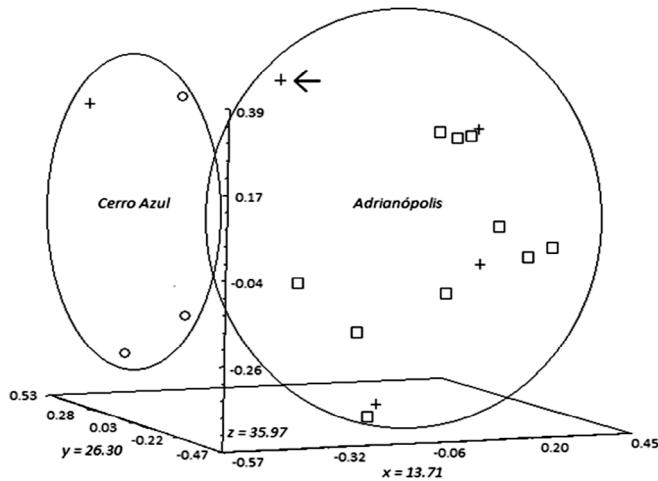


Figure 3. Principal Coordinate Analysis (PCO) of specimens of *Lutzomyia intermedia* from Cerro Azul and Adrianópolis. The graph was generated by the program NTSYS pc 2.1, from electrophoresis data performed with primer A2 RAPD-PCR products. Symbols: (°) house, (□) peridomicile, (+) wild. Specimen indicated by the arrow belongs to the *Lutzomyia intermedia* s.s. lineage.

1987, which has allowed to differentiate from the nearby species, *Lutzomyia spinicrassa* Morales, Osorno-Mesa, Osorno and Hoyos, 1969.

In our study, RAPD markers did not generate genetic profiles that allowed us to identify the sand flies according to the morphological criteria used to distinguish the females of the taxon *L. intermedia* proposed by Marcondes (1996). This is contrasting to the study by Silva *et al.* (2011), who analyzed populations of *Lutzomyia* (*Lutzomyia*) *longipalpis* (Lutz and Neiva, 1912) in the state of Maranhão, northeastern Brazil, using RAPD, and divided the population according to the morphological markers found in males that they used to distinguish the existing varieties within this species. In the case of *L. longipalpis*, the possible existence of a species complex based on the morphological markers present in males has been justified by several genetic and behavioral studies (Souza *et al.*, 2004b; Lima Costa *et al.*, 2015; Vigoder *et al.*, 2015; Souza *et al.*, 2017).

The results obtained in this study, using primer A10, denoted the existence of gene flow within the study population (old and new transmission areas of ACL). Specimens collected in different ecotypes and different municipalities were grouped in the same phenetic subgroup with varying degrees of similarity, as the results obtained by similar studies involving *Anopheles* Meigen, 1818 species (Posso *et al.*, 2003; González *et al.*, 2007). The evidence of gene flow is denoted not only by analysis of the dendrogram, but also considering the observation that sand flies from different municipalities showed the same genetic profile. In this study, the occurrence of gene flow can be explained by the lack of substantial physical barriers between the two selected areas that are about 30 km distant.

Analogous studies have reported similar trends, e.g., the work proposed by Pinedo-Cancino *et al.* (2006). The authors used RAPD to verify microgeographic genetic variation in nine geographically separated populations (20-60 km) of *Anopheles* (*Nyssorhynchus*) *darlingi* Root, 1926 in the municipality of Iquitos (Peru). The average genetic distance obtained demonstrated the genetic homogeneity between these populations and, thus, the presence of a high level of gene flow. As a counterpoint, the genetic homogeneity verified in populations of *L. longipalpis* in the northeast region of Brazil, is justified by Balbino *et al.* (2006) as the result of geographic isolation and restricted gene flow. This is compatible with the hypothesis that there is a single species of the sand fly in this area.

The municipalities of Cerro Azul and Adrianópolis belong to the same ecosystem, the Ribeira River Valley, and they border each other. The absence of geographic isolation of these populations allows sand flies to migrate and exchange genetic material among themselves in different areas. Furthermore, this process may have been facilitated by the impact of the construction of the Bolivia-Brazil gas pipeline, which runs through the municipality of Cerro Azul. In fact, large-scale anthropogenic interventions can influence the levels of genetic structuring in vectors populations and, as a consequence, have an impact on the transmission potential of pathogens (Burkett-Cadena and Vittor, 2018; Suesdek, 2019).

Considering the lack of barriers to flight, *L. intermedia* can move, specially helped by the environmental changes of the gas pipeline construction and might ensure the gene flow in the Ribeira Valley, Paraná. This hypothesis is supported by the dispersal potential that these insects possess. The *L. intermedia* dispersion and survival were studied by Galati *et al.* (2009) in Iporanga, endemic area of ACL in the Ribeira Valley of the state of São Paulo. They reported that the dispersal distances were 109 m for *L. intermedia* s.s. and 100 m for *L. neivai*. The maximum dispersal distance was 180 m for *L. intermedia* s.s., while they recovered *L. neivai* in a pasture 250 m away and another in a pigsty 520 m away, showing a trend to disperse to more open areas.

The RAPD analysis can also be a useful tool to distinguish insects depending on their breeding sites/locations. This is demonstrated by the graph generated by PCO using data generated by the A2 primer. It reveals two major subgroups: the first formed by sand fly samples from Adrianópolis, and the second consisting of samples captured in Cerro Azul. A study proposed by Dvorak *et al.* (2006) sought to clarify the taxonomic nature of *Phlebotomus* (*Paraphlebotomus*) *sergenti* Parrot, 1917 by developing colonies with populations of this sand fly from Turkey and Israel. They performed RAPD with the offspring, which indicated the formation of distinct subgroups related to the origin of each colony. However, there was a high level of variability within each subgroup.

In this study, the group that involved sand flies from Adrianópolis included samples from the forest together

with peridomestic areas. In the Cerro Azul group, a sand fly from the forest belonged to the same group as three other samples from the inside of the house. These arrangements of specimens from ecotypes that differ from those in which they were prevalent can also be justified by the existence of migration and gene flow. After evaluating the genetic variability of four populations of *Lutzomyia* (*Nyssomyia*) *whitmani* (Antunes and Coutinho, 1939) from different areas of ACL transmission in Brazil using RAPD, Souza *et al.* (2004a) reported two main spatial groupings with a high level of intra-population variability: Corte de Pedra, Ilhéus (Bahia) and Serra de Baturité (Ceará) in the first group, and Martinho Campos (Minas Gerais) in the second. The vast majority of individuals of these populations were grouped according to their region of origin. However, unlike females, 13 % of males from Martinho Campos moved to Serra de Baturité, instead of Ilhéus. The non-uniformity in the genetic lineage of Martinho Campos is justified by the existence of sympatric populations in these regions, because individuals with intermediate profiles can be explained by the gene flow among these populations.

Here, the peridomicile was the ecotype with the highest genotypic diversity. This diversity can be explained by geographical conditions present at the collection sites. Meneses *et al.* (2005) compared three populations of *L. intermedia* from the same endemic area of ACL in the state of Rio de Janeiro, Brazil. Using RAPD, they were able to distinguish between two major subgroups, one related to the house or peridomicile and one related to the wild (woods). The latter was the most polymorphic, with the largest number of genotypes and a low degree of similarity. According to the authors, due to the high level of gene flow between different habitats, five individuals from the wild were grouped with the subgroup formed by sand flies from the house and peridomicile, and one individual from the house area was grouped with the subgroup of samples collected in the wild.

To explain the genetic diversity within and between populations of *L. intermedia* from the municipalities of Afonso Cláudio and Viana (state of Espírito Santo, Brazil), Rocha *et al.* (2007) mention the years of human colonization of the two sites (200 and 80 years, respectively), which may reflect the differences in the behavior of these populations. The authors point out that although there was a trend to group individuals collected in the same ecotype into the same phenetic group, this fact does not necessarily indicate a genetic separation between the populations. Moreover, according to Rocha *et al.* (2007), the high level of genetic organization of populations of *L. intermedia* in different ecotypes of the Viana may reflect the independence between the domestic and peridomestic transmission cycles of *Leishmania*, such that the genetic divergence of *L. intermedia* may be occurring because of changes in their original forest habitat due to the ability to adapt to a new environment.

The Ribeira Valley is a region heavily influenced by anthropogenic changes, with varying degrees of degradation in the original landscape. *L. intermedia* is a species that has great adaptability in modified environments, showing synanthropic behavior and attraction to domestic animals. These aspects favor the transmission of the parasite in the house and peridomestic area (Gomes *et al.*, 1986; Saraiva *et al.*, 2012).

The high genetic diversity verified herein cannot suggest an independence between the cycles of wild, peridomicile, and household transmission of ACL, because these ecotypes are very close to each other and lack the geographical barriers that could separate the populations according to ecotypes or strains (Saraiva *et al.*, 2012; Gonçalves *et al.*, 2019). Therefore, the high genetic variability shown for *L. intermedia*, combined with its high prevalence, supports *L. intermedia* as a vector of *L. braziliensis* in the Ribeira Valley.

Since it has shown to be the most prevalent sand fly species in the Ribeira River Valley region and has been considered one of the primary vectors of *L. braziliensis* in South America, *L. intermedia* has become an undisputed target for further studies.

The present investigation has met a part of these demands, by contributing to the genetic characterization of this species. Further investigation is important not only for new knowledge about its taxonomy and genetic diversity, but for further clarification on the participation in the *Leishmania* transmission cycle. Thus, underlying research involving molecular biology using techniques and resembling this study becomes compelling. Among these, we can mention the characterization of ribosomal DNA sequences (Almazán *et al.*, 2020); that have used other markers recognized as useful in specific differentiation, such as the *period* gene (Freitas *et al.*, 2018); and protein analyzes, such as those performed by mass spectrometry (Mathis *et al.*, 2015).

CONCLUSIONS

In this study, a high genotypic diversity and a detectable gene flow have been found among populations of *L. intermedia* that have been collected in the Ribeira River Valley Region, southern Brazil, through the application of RAPD. It was verified, in the studied area, that the highest level of *L. intermedia* polymorphism was found among the insects captured in the peridomicile. Furthermore, the data suggest that the occurrence of sand fly migrations among this ecotype, the wild and the house, allow the transmission of *L. braziliensis* between animals and humans in the Ribeira River Valley in both ecotypes, as well as along the investigated municipalities. This condition of vector species is corroborated by the high genetic variability found.

RAPD analysis has been frequently used to determine the level of genetic variability in various insect species. In public health, in addition to population genetics, the RAPD and other molecular biology techniques offer special interest regarding pathogen vectors, both to elucidate whether there is a relationship

between the vector genotype and its vector competence, as well as to rule out doubts about the species complexes that present difficulties for identification by morphological characters. In this study, it has not been possible to obtain profiles that were related to morphological markers used to distinguish between the lineages that set *L. intermedia*. However, the analysis with different markers has enabled to group populations according to their bio-geographical origin.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ACKNOWLEDGMENTS

The authors are grateful to the CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) and to the CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico) for providing financial support. We also thank the residents of Cerro Azul and Adrianópolis for authorizing us to collect sand flies on their properties and Daniel Ernesto Rodríguez Fernández, PhD, from the Universidade Federal do Rio de Janeiro, for reviewing the *resumen*. The authors gratefully acknowledge the valuable suggestions from two referees.

REFERENCES

- Adamson RE, Ward RD, Feliciangeli MD, Maingon R. The application of random amplified polymorphic DNA for sandfly species identification. *Med Vet Entomol.* 1993;7(3):203-207. Doi: <https://doi.org/10.1111/j.1365-2915.1993.tb00677.x>
- Almazán MC, Copa GN, Lauthier JJ, Gil JF, Quiroga IL, Hoyos CL, *et al.* Sand fly typing: a simple and morphologically-supported method based on polymorphism of 18S rRNA gene in a Leishmaniasis endemic area of Argentina. *Acta Trop.* 2020;211:105609. Doi: <https://doi.org/10.1016/j.actatropica.2020.105609>
- Andrade Filho JD, Galati EAB, Falcão AL. *Nyssomyia intermedia* (Lutz & Neiva, 1912) and *Nyssomyia neivai* (Pinto, 1926) (Diptera: Psychodidae: Phlebotominae) geographical distribution and epidemiological importance. *Mem Inst Oswaldo Cruz.* 2007;102(4):481-487. Doi: <https://doi.org/10.1590/S0074-02762007005000035>
- Azevedo ACR, Monteiro FA, Cabello PH, Souza NA, Rosa-Freitas MG, Rangel EF. Studies on Populations of *Lutzomyia longipalpis* (Lutz & Neiva, 1912) (Diptera: Psychodidae: Phlebotominae) in Brazil. *Mem Inst Oswaldo Cruz.* 2000;95(3):305-322. Doi: <https://doi.org/10.1590/S0074-02762000000300005>
- Balbino VQ, Coutinho-Abreu IV, Sonoda IV, Melo MA, de Andrade PP, de Castro JA, *et al.* Genetic structure of natural populations of the sand fly *Lutzomyia longipalpis* (Diptera: Psychodidae) from the Brazilian northeastern region. *Acta Trop.* 2006;98(1):15-24. Doi: <https://doi.org/10.1016/j.actatropica.2006.01.007>
- Bejarano EE, Rojas W, Uribe S, Vélez ID, Porter, CH. Genetic analysis of a recently detected urban population of *Lutzomyia evansi* (Diptera: Psychodidae) in Colombia. *Rev Soc Entomol Argent.* 2009;68(1-2):135-141.
- Burkett-Cadena ND, Vittor AY. Deforestation and vector-borne disease: Forest conversion favors important mosquito vectors of human pathogens. *Basic Appl Ecol.* 2018;26:101-110. Doi: <https://doi.org/10.1016/j.baae.2017.09.012>
- Castro EA, Luz E, Telles FQ, Pandey A, Biseto A, Dinaiski M, *et al.* Eco-epidemiological survey of *Leishmania (Viannia) braziliensis* American cutaneous and mucocutaneous leishmaniasis in Ribeira Valley River, Paraná State, Brazil. *Acta Trop.* 2005;93(2):141-149. Doi: <https://doi.org/10.1016/j.actatropica.2004.10.004>
- Chan-Chable RJ, Martínez-Arce A, Mis-Avila PC, Ortega-Morales AI. DNA barcodes and evidence of cryptic diversity of anthropophilic mosquitoes in Quintana Roo, Mexico. *Ecol Evol.* 2019;9(8):4692-4705. Doi: <https://doi.org/10.1002/ece3.5073>
- Coelho ASG. Bood: Avaliação de dendrogramas baseados em estimativas de distâncias/similaridades genéticas através do procedimento de bootstrap [computer program]. Version 3.03. Goiânia: Universidade Federal de Goiás; 2005.
- Dvorak V, Aytekin AM, Alten B, Skarupova S, Votypka J, Volf P. A comparison of the intraspecific variability of *Phlebotomus sergenti* Parrot, 1917 (Diptera: Psychodidae). *J Vector Ecol.* 2006;31(2):229-238. Doi: [https://doi.org/10.3376/1081-1710\(2006\)31\[229:ACOTIV\]2.0.CO;2](https://doi.org/10.3376/1081-1710(2006)31[229:ACOTIV]2.0.CO;2)
- Forattini OP, Rabello EX, Serra OP, Cotrim MD, Galati EAB, Barata JM. Observações sobre a transmissão da Leishmaniose Tegumentar no Estado de São Paulo, Brasil. *Rev Saude Publica.* 1976;10(1):31-43. Doi: <https://doi.org/10.1590/S0034-89101976000100003>
- Fraga J, Rodriguez J, Fuentes O, Hernández Y, Castex M, Gonzalez R, *et al.* Genetic variability of *Triatoma flavida* and *Triatoma bruneri* (Hemiptera: Reduviidae) by RAPD-PCR technique. *Rev Inst Med Trop Sao Paulo.* 2011;53(1):19-24. Doi: <https://doi.org/10.1590/S0036-46652011000100004>
- Freitas MTS, Costa Jr CRL, Silva LG, Leal-Balbino TC, Brazil RP, Balbino VQ. Genetic Structure of Sympatric Populations of Female *Lutzomyia longipalpis* (Diptera: Psychodidae) In Sobral and Caririáçu, Ceará State, Brazil. *Vector Biol J.* 2018;3(1):1-5. Doi: <https://doi.org/10.4172/2473-4810.1000125>
- Galati EAB, Fonseca MB, Marassá AM, Bueno EFM. Dispersal and survival of *Nyssomyia intermedia* and *Nyssomyia neivai* (Diptera: Psychodidae: Phlebotominae) in a cutaneous leishmaniasis endemic area of the speleological province of the Ribeira Valley, state of São Paulo, Brazil. *Mem Inst*

- Oswaldo Cruz. 2009;104:1148-1158. Doi: <https://doi.org/10.1590/S0074-02762009000800012>
- Golczer G, Arrivillaga J. Use and trends of molecular markers in sandflies (Diptera: Psychodidae). *Bol Mal Salud Amb*. 2015;55(1):19-40.
- Gomes AC, Santos JLF, Galati EAB. Ecological aspects of American cutaneous leishmaniasis: 4. Observations on the endophilic behavior of the sandfly and the vectorial role of *Psychodopygus intermedius* in the Ribeira Valley region of the S. Paulo State, Brazil. *Rev de Saude Publica*. 1986;20(4):280-287. Doi: <https://doi.org/10.1590/S0034-89101986000400003>
- Gonçalves AL, Luz E, Castro EA, Klisiowicz DR, Gazda TL, Melo ALA, *et al*. Abundance and diversity of vectors (Diptera: Psychodidae) in an old transmission area of cutaneous leishmaniasis in the new world after Bolivia-Brazil gas pipeline construction. *Mem Inst Investig Cienc Salud*. 2019;17(2):16-23. Doi: <https://doi.org/10.18004/mem.iics/1812-9528/2019.017.02.16-023>
- González R, Wilkerson R, Suárez MF, García F, Gallego G, Cárdenas H, *et al*. A population genetics study of *Anopheles darlingi* (Diptera: Culicidae) from Colombia based on random amplified polymorphic DNA-polymerase chain reaction and amplified fragment length polymorphism markers. *Mem Inst Oswaldo Cruz*. 2007;102(3):255-262. Doi: <https://doi.org/10.1590/S0074-02762007005000037>
- Hiragi C, Simões K, Martins E, Queiroz P, Lima L, Monnerat R. Variabilidade Genética em Populações de *Aedes aegypti* (L.) (Diptera: Culicidae) Utilizando Marcadores de RAPD. *Neotrop Entomol*. 2009;38(4):542-547. Doi: <https://doi.org/10.1590/S1519-566X2009000400018>
- Ipardes (Instituto Paranaense de Desenvolvimento Econômico e Social). *Cadernos municipais*. Curitiba; 2021. Available in: <http://www.ipardes.pr.gov.br/Pagina/Cadernos-municipais> Cited: 13 may 2021.
- Jaccard P. Nouvelles recherches sur la distribution florale. *Bull Soc Vaudoise Sci Nat*. 1908;44:223-270.
- Lima Costa CR, Freitas MTS, Figueirêdo Jr CAS, Aragão NS, Silva LG, Marcondes CB, *et al*. Genetic structuring and fixed polymorphisms in the gene *period* among natural populations of *Lutzomyia longipalpis* in Brazil. *Parasit Vectors* 2015;8:193. Doi: <https://doi.org/10.1186/s13071-015-0785-6>
- Loxdale HD, Lushai G. Molecular markers in entomology. *Bull Entomol Res*. 1998;88(6):577-600. Doi: <https://doi.org/10.1017/S0007485300054250>
- Lozano-Sardaneta YN, Paternina LE, Sánchez-Montes S, Quintero A, Ibáñez-Bernal S, Sánchez-Cordero V, *et al*. DNA barcoding and fauna of phlebotomine sand flies (Diptera: Psychodidae: Phlebotominae) from Los Tuxtlas, Veracruz, Mexico. *Acta Trop*. 2020;201:105220. Doi: <https://doi.org/10.1016/j.actatropica.2019.105220>
- Mantel N. The Detection of Disease Clustering and a Generalized Regression Approach. *Cancer Res*. 1967;27(2):209-220.
- Marcondes CB. A Redescription of *Lutzomyia (Nyssomyia) intermedia* (Lutz & Neiva, 1912), and Resurrection of *L. neivai* (Pinto, 1926) (Diptera, Psychodidae, Phlebotominae). *Mem Inst Oswaldo Cruz*. 1996;91(4):457-462. Doi: <https://doi.org/10.1590/S0074-02761996000400012>
- Marcondes CB. Morfometria e DNA mitocondrial de populações sul americanas de *Lutzomyia (Nyssomyia) intermedia* (Lutz & Neiva, 1912) (Diptera, Psychodidae, Phlebotominae). *Rev Soc Bras Med Trop*. 1997;30(6):533-534. Doi: <https://doi.org/10.1590/S0037-86821997000600017>
- Mathis A, Depaquit J, Dvořák V, Tuten H, Bañuls AL, Halada P, *et al*. Identification of phlebotomine sand flies using one MALDI-TOF MS reference database and two mass spectrometer systems. *Parasit Vectors*. 2015;8:266. Doi: <https://doi.org/10.1186/s13071-015-0878-2>
- Meneses CR, Cupolillo E, Monteiro F, Rangel EF. Microgeographical variation among male populations of the sandfly, *Lutzomyia (Nyssomyia) intermedia*, from an endemic area of American cutaneous leishmaniasis in the state of Rio de Janeiro, Brazil. *Med Vet Entomol*. 2005;19(1):38-47. Doi: <https://doi.org/10.1111/j.0269-283X.2005.00535.x>
- Mukhopadhyay J, Ghosh K, Braig HR. Identification of cutaneous Leishmaniasis vectors, *Phlebotomus papatasi* and *P. duboscqi* using random amplified polymorphic DNA. *Acta Trop*. 2000;76(3):277-283. Doi: [https://doi.org/10.1016/S0001-706X\(00\)00130-3](https://doi.org/10.1016/S0001-706X(00)00130-3)
- Pinedo-Cancino V, Sheen P, Tarazona-Santos E, Oswald WE, Jeri C, Vittor AY, *et al*. Limited diversity of *Anopheles darlingi* in the Peruvian Amazon region of Iquitos. *Am J Trop Med Hyg*. 2006;75(2):238-245. Doi: <https://doi.org/10.4269/ajtmh.2006.75.238>
- Posso CA, González R, Cárdenas H, Gallego G, Duque MC, Suarez MF. Random Amplified Polymorphic DNA Analysis of *Anopheles nuneztovari* (Diptera: Culicidae) from Western and Northeastern Colombia. *Mem Inst Oswaldo Cruz*. 2003;98(4):469-476. Doi: <https://doi.org/10.1590/S0074-02762003000400007>
- Rocha LS, Falqueto A, Santos CB, Grimaldi Jr G, Cupolillo E. Genetic structure of *Lutzomyia (Nyssomyia) intermedia* populations from two ecologic regions in Brazil where transmission of *Leishmania (Viannia) braziliensis* reflects distinct eco-epidemiologic features. *Am J Trop Med Hyg*. 2007;76(3):559-65. Doi: <https://doi.org/10.4269/ajtmh.2007.76.559>
- Rohlf FJ. NTSYS-pc: Numerical taxonomy and multivariate analysis system [computer program]. Version 2.1. New York: Exeter Software; 2000.
- Salomón OD, Mastrángelo AV, Santini MS, Liotta DJ, Yadón ZE. Retrospective eco-epidemiology as a tool for the

- surveillance of leishmaniasis in Misiones, Argentina, 1920-2014. *Rev Panam Salud Publica*. 2016;40(1):29-39.
- Saraiva L, Carvalho GM, Sanguinette CC, Carvalho DAA, Andrade Filho JD. Biogeographical aspects of the occurrence of *Nyssomyia neivai* and *Nyssomyia intermedia* (Diptera: Psychodidae) in a sympatric area of the Brazilian savannah. *Mem Inst Oswaldo Cruz*. 2012;107(7):867-872. Doi: <https://doi.org/10.1590/S0074-02762012000700005>
- Savoia D. Recent updates and perspectives on leishmaniasis. *J Infect Dev Ctries*. 2015;9(6):588-596. Doi: <https://doi.org/10.3855/jidc.6833>
- Seblova V, Volfova V, Dvorak V, Pruzinova K, Votypka J, Kassahun A, et al. *Phlebotomus orientalis* sand flies from two geographically distant Ethiopian localities: biology, genetic analyses and susceptibility to *Leishmania donovani*. *PLoS Negl Trop Dis*. 2013;7(4):e2187. Doi: <https://doi.org/10.1371/journal.pntd.0002187>
- Sharma A, Kumar S, Tripathi P. Assessment of *Achyranthes aspera* induced toxicity and molecular analysis of RAPD-PCR profiles of larval genomic DNA of *Aedes aegypti* L. (Diptera: Culicidae). *J Parasit Dis*. 2017;41(4):1066-1073. Doi: <https://doi.org/10.1007/s12639-017-0935-1>
- Silva MH, Nascimento MD, Leonardo FS, Rebêlo JM, Pereira SR. Genetic Differentiation in Natural Populations of *Lutzomyia longipalpis* (Lutz & Neiva) (Diptera: Psychodidae) with Different Phenotypic Spot Patterns on Tergites in Males. *Neotrop Entomol*. 2011;40(4):501-506. Doi: <https://doi.org/10.1590/S1519-566X2011000400015>
- Sneath P, Sokal RR. Numerical taxonomy: the principles and practice of numerical classification. San Francisco: W. H. Freeman; 1973. 573 p.
- Souza CM, Fortes-Dias CL, Linardi PM, Dias ES. Phenetic studies on randomly amplified polymorphic DNA-polymerase chain reaction-variability of four geographical populations of *Lutzomyia whitmani* (Diptera: Psychodidae) in Brazil. *Rev Soc Bras Med Trop*. 2004a;37(2):148-153. Doi: <https://doi.org/10.1590/S0037-86822004000200007>
- Souza NA, Brazil RP, Araki AS. The current status of the *Lutzomyia longipalpis* (Diptera: Psychodidae: Phlebotominae) species complex. *Mem Inst Oswaldo Cruz*. 2017;112(3):161-174. Doi: <https://doi.org/10.1603/0022-2585-41.5.906>
- Souza NA, Vigoder FM, Araki AS, Ward RD, Kyriacou CP, Peixoto AA. Analysis of the Copulatory Courtship Songs of *Lutzomyia longipalpis* in Six Populations from Brazil. *J Med Entomol*. 2004b;41(5):906-913. Doi: <https://doi.org/10.1603/0022-2585-41.5.906>
- Sudia WD, Chamberlain RW. Battery-operated light trap, an improved model. *Mosq News*. 1962;22:126-129.
- Suesdek L. Microevolution of medically important mosquitoes - A review. *Acta Trop*. 2019;191:162-171. Doi: <https://doi.org/10.1016/j.actatropica.2018.12.013>
- Tibayrenc M, Neubauer K, Barnabé C, Guerrini F, Skarecky D, Ayala FJ. Genetic characterization of six parasitic protozoa: parity between random-primer DNA typing and multilocus enzyme electrophoresis. *Proc Natl Acad Sci USA*. 1993;90(4):1335-1339. Doi: <https://doi.org/10.1073/pnas.90.4.1335>
- Vigoder FM, Souza NA, Brazil RP, Bruno RV, Costa PL, Ritchie MG, et al. Phenotypic differentiation in love song traits among sibling species of the *Lutzomyia longipalpis* complex in Brazil. *Parasit Vectors* 2015;8:290. Doi: <https://doi.org/10.1186/s13071-015-0900-8>
- Young DG, Duncan MA. Guide to the identification and geographic distribution of *Lutzomyia* sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae). *Mem Am Entomol Inst*. 1994;54:1-881. Doi: <https://doi.org/10.21236/ADA285737>