



Image analysis for the evaluation of soybean seeds vigor

Análisis de imágenes para la evaluación del vigor de semillas de soja

Erich Barros Brandani ^{1,4}, Nara Oliveira Silva Souza ^{1,5}, Nilson Matheus Mattioni ^{1,4}, Flívia Fernandes de Jesus Souza ^{1,7}, Michelle Souza Vilela ^{1,8}, Érica Alves Marques ^{3,9}, Wallas Felippe de Souza Ferreira ^{1,10}.

¹Universidade de Brasília. Asa Norte, Brasília, Brasíl. ²Universidade Federal de Santa Maria. Santa Maria, Brasil. ³Empresa Tbit Sistemas e Tecnologias. Lavras, Brasil. ⁴ ⊠ erich_bb@hotmail.com, ⁵ ⊠ narasouza@unb.br. ⁶ ⊠ nilsonmattioni@ufsm.br. ⁷ ⊠ fliviafdejesus@gmail.com. ⁸ ⊠ michellevilelaunb@gmail.com. ⁹ ⊠ ericagroeng@yahoo.com.br. ¹⁰ ⊠ wallasfelippe@gmail.com

e0989 https://doi.org/10.15446/acag.v70n3.92093

2021 | 70-3 p 311-316 | ISSN 0120-2812 | e-ISSN 2323-0118 | Rec.: 02-12-2020. Acep.:09-02-2022

Abstract

The search for time and human subjectivity reduction in the analysis of the physiologic potential of seeds has brought forward the use of computerized systems as an alternative. The goal of this research was to assess the effectiveness of the GroundEve equipment in the characterization of soybean seeds vigor by using the image analysis technique in comparison to the information provided by vigor tests recommended to soybean seeds. Sixteen lots of two soybean varieties were used - variety 98Y12 and P98Y70, characterized by the following tests: germination test, accelerated ageing test, cold test, tetrazolium test, and seedling emergence in the field. The image analysis was performed using the GroundEye software to obtain the vigor index and the average length of seedlings and roots for a later comparison with the results of the accelerated ageing test and the manual measurement of the average length of seedlings and roots. GroundEye has demonstrated efficiency in the determination of vigor of soybean seeds and has reduced the time needed to obtain the results in six days, being indicated in the quality control programs established by soybean seeds production companies.

Keywords: Glycine max L., physiological quality, GroundEye software.

Resumen

La búsqueda de la reducción del tiempo y de la subjetividad humana en el análisis del potencial fisiológico de las semillas ha planteado como alternativa el uso de sistemas informáticos. El objetivo de esta investigación es evaluar la efectividad del equipo GroundEye en la caracterización del vigor de semillas de soja mediante la técnica de análisis de imágenes en comparación con la información proporcionada por las pruebas de vigor recomendadas para semillas de soja. Se utilizaron dieciséis lotes de dos variedades de soja, la variedad 98Y12 y P98Y70, caracterizaron por las siguientes pruebas: prueba de germinación, ensayo de envejecimiento acelerado, prueba de frío, prueba de tetrazolio, y emergencia de plántulas en el campo. El análisis de imágenes se realiza mediante el software GroundEye para obtener el índice de vigor y la longitud promedio de plántulas y raíces para su posterior comparación con los resultados de la prueba de envejecimiento acelerado y la medición manual de la longitud promedio de plántulas y raíces. GroundEye ha demostrado eficiencia en la determinación del vigor de semillas de soja y ha reducido el tiempo necesario para obtener los resultados en seis días, siendo indicado en los programas de control de calidad establecidos por las empresas productoras de semillas de soja.

Palabras clave: Glycine max L., calidad fisiológica, software GroundEye.

Introduction

The commercial production of soybean seeds aims to obtain healthy seeds with a high physiological potential. In order to aid the decisions process regarding the destination of seed lots and to identify differences in performance of the same, it is necessary an evaluation of the physiological potential of the lot. Such evaluation is usually performed through germination tests and measurement of the length of the seedlings and their parts (Medeiros *et al.*, 2019).

Vigor tests require a strict control of the test variables, a more comprehensive interpretation of results and a more specific training by analysts (Marcos-Filho, 2015). These tests are susceptible to human subjectivity of the assessment and require a longer period of time for results to be obtained. Aiming to minimize such limitations, efforts in the development of specific computerized equipment have been made in search of standardizing vigor tests (Marcos-Filho, 2015). In this sense, image analysis techniques associated with vigor tests of seeds have an immense use potential.

In Brazil, the company Tbit Tecnologia e Sistemas has developed a system called GroundEye (GE), which evaluates the vigor of soybean seeds through the development of seedlings. This evaluation is made by analysing high resolution images obtained by the equipment, which allow the extraction of information on colour, texture and geometry, serving as a parameter for the provision of growth indexes, uniformity and vigor of seedlings.

The goal of this work is to verify the efficiency of the GroundEye equipment in characterizing the vigor of soybean seed lots in comparison to other tests recommended in the evaluation of soybean seeds, in search of a methodology which grants less subjectivity and that might be performed in a smaller period of time.

Material and Methods

Characterization of lots. Sixteen lots of soybean seeds with a different physiological quality were used, being eight lots of the variety 1 (98Y12) and eight lots of the variety 2 (P98Y70). Both were provided by the company DuPont[®] –Pioneer Seeds Division (Pioneer[®]) – having a late cycle, determined growth habits and Roundup Ready[®] gene.

The study has been conducted at the Seed Laboratories of the DuPont Company, Pioneer Seeds Division, and at the Faculty of Agronomy and Veterinary Medicine of the Darcy Ribeiro Campus, University of Brasília, UnB. Field experiments were carried out at Fazenda Água Limpa, UnB, located in the rural area of Vargem Bonita, Federal District, Brazil (DF).

Water content. We determined the seed water content by the greenhouse method at 105 °C \pm 2 °C for 24 hours using two replicates and 5 g of seeds per treatment. The results were expressed as percentage (wet basis) (Brasil, 2009).

.

Germination test. For the germination test, we used four replications with 50 seeds per treatment. They were germinated in germitest paper in rolls, moistened with distilled water with a volume equivalent to 2.5 times the weight of the dry paper, and placed in a germinator at 25 °C ± 2 °C, according to the criteria described in the Rules for Seed Analysis. A single reading was performed on the fifth day after sowing, where we counted the number of normal seedlings (Brasil, 2009).

Accelerated aging test. The accelerated aging test was performed using the plastic germination box method (AOSA, 2009). Seeds of each treatment were distributed over an internal stainless-steel screen in a single layer, so as to cover it completely and uniformly, and 40 ml of distilled water were added at the bottom of the box. The boxes were capped and kept in BOD (biochemical oxygen demand) regulated at 41 °C \pm 2 °C for 48 hours for the subsequent germination test. A single reading was performed on the seventh day after sowing by determining the percentage of normal seedlings (Marcos-Filho *et al.*, 2009).

Cold test. For the cold test, four replications of 50 seeds were used for each treatment. Sowing was performed in plastic trays using a 2:1 mixture of soil and sand moistened at 60 % of its water retention capacity. After sowing, the boxes were kept in a cold room at 10 °C \pm 2 °C for five days. Subsequently, the boxes were conditioned in a plant growth chamber at 25 °C \pm 2 °C for five days, when we performed the counting of emerged seedlings (Barros *et al.*, 1999).

Tetrazolium test. To conduct the tetrazolium test, the methodology described by França Neto *et al.* (1998) has been applied, using samples of 100 seeds (two subsamples with 50 seeds) for each variety, treatment and repetition, in which the samples had been preconditioned between sheets of moistened paper towel, with a volume of water equivalent to 2.5 times the mass of the paper, for 16 hours at 25 °C. The seeds were then placed in plastic containers and submerged in a 0.075 % solution of 2,3,5-triphenyl tetrazolium chloride at 40 °C, in the dark, for 150 minutes. After this period, the seeds were washed under running water and analysed individually.

Seedling emergence in the field test. The sowing has been performed manually with four repetitions of 50 seeds per treatment. The parcels were randomly distributed with a 10 cm space between the lines and the counting of the seedlings has been done seven days after the installation (Brasil, 2009).

Image analysis. To obtain seedling images, we performed a germination test with four replications and 50 seeds per treatment using as a substrate a

paper towel in the form of a roll moistened with a water content equivalent to 2.5 times its dry weight. Then, the rolls were conditioned in a germinator at 25 °C \pm 2 °C for three days, when images were captured (Pinto *et al.*, 2015).

Capturing the images. To capture images, the seedlings were carefully removed from the paper towel and placed on a transparent acrylic tray so that seedlings did not touch each other. Subsequently, the tray was inserted into the capture chamber of the equipment for image capture and analysis. Germination test images were captured by a professional high-resolution camera contained in the GroundEye capture chamber (Pinto *et al.*, 2015).

The background colour calibration was HSV colour space, hue between 198.4 and 278.4, saturation between 0.389 and 1.000, and brightness between 0.000 and 1.000. The type of recognition parameter was seedlings with three days, depth fill not selected, and minimum object discard size of 0.3 cm².

Seedling and root length. After the image capturing of seedlings of the germination test on the third day, we carried out a manual measurement of seedling length (SL) and root length (CR) (Nakagawa, 1999) to later compare them with the results obtained by the software. To carry out measurements, a ruler fixed to the table was used (reading in centimetres). For dead or non-germinated seeds, a zero value was assigned. The result was expressed in mean seedling and root length, by manual measurement and software measurement (Nakagawa, 1999).

Automatic calibration. The automatic calibration process performed by the GroundEye (GE) software consisted of capturing images of soybean seedlings three days after sowing in the germination test and the later association with vigor value obtained by accelerated ageing test for the same lots.

We used 38 lots with approximately 100 seedlings per lot for image capturing. Variation in vigor was 44-97 %, according to the accelerated ageing test. These lots were different from those adopted for data collection.

After this process, the value obtained by the accelerated ageing test was associated by the system to its respective image, so that, by multivariate regression, the weight of the parameters contained in the formulas described below was defined, according to Pinto *et al.* (2015):

Where: l_h and l_r : sample means of hypocotyl and root length; s_h , s_r , s_{total} and $s_{r/h}$: standard deviation of hypocotyl and root length, total length and ratio between root and hypocotyl; p_c and p_u : weight of growth and uniformity indexes; *w*: weight of the parameters.

Experimental design and statistical analysis. The experiment has been conducted in a completely random design with four repetitions, with each sample containing 50 seeds, both in the first stage of the characterization of the lots (water content, germination, emergence of seedlings in the field, accelerated ageing, cold test and tetrazolium), and in the second stage of the evaluation of efficiency of the GroundEye. In the second stage new tests have been conducted (accelerated ageing, seedling length and root length), following the same methodology of characterization of the lots, denominated as manual tests, and its results were compared to the results obtained through the GroundEye (automatic vigor, seedling length and root length), denominated as automatic tests. Thus, such evaluation has been accomplished in an 8x2 factorial scheme for each of the varieties isolated, being 8 lots of seeds and 2 types of tests (manual and automatic). We performed an analysis of variance, and means were compared by a Scott-Knott test at 5 % probability. The software used was ASSISTAT version 7.7 (Silva & Azevedo, 2009).

Results

The water content of the eight lots of both soybean varieties was similar. For the lots of the variety 1, there was a variation from 10.6 % to 11.9 %, and for lots of variety 2, there was a variation from 11.2 % to 12.2 %.

In all lots of the variety 1, the values were above 80 % by the germination test. The lots 1, 2, 5, 6 and 7 had the highest values, and the lots 3, 4, and 8 had the lowest values (Table 1).

For seedling emergence in the field test, the lots 3 and 8 differed statistically from the others, obtaining the lowest emergence in the field percentages. By the accelerated ageing test, there was a potential of experimental lots, allowing the classification of lots into two levels: lots 1, 2, 5, and 7 were the most vigorous lots, and the lots 3, 4, 6, and 8 were the less vigorous lots. Such result is consistent with the high germination level under laboratory conditions and with field emergence observed for the lots 1, 2, 5, 6, and 7.

The results of the tetrazolium test (TZViab and TZVig) and the cold test corroborate those of the other tests: the lots 1, 2, 5, 6, and 7 had the best results, although they did not differ statistically

- Growth Index = min $(w_h * l_h + w_r * l_r, 1,000)$	Eq. 1
- Uniformity index = max (1,000 - ($w_{sh} * s_{h} + w_{sr} * s_{r} + s_{total} + w_{sr/h} * s_{r/h}$) - $W_{m} *$ no dead seeds, 0)	Eq. 2
- Vigor Index = p_c growth + p_u uniformity	Eq. 3

from the lot 8 in the tetrazolium test. Such results demonstrate the high performance of such lots in a wide range of environmental conditions.

The results for lot vigour of this variety by accelerated ageing test, cold test and tetrazolium test were higher than 80 %, with the exception of the lots 4 and 8 in the accelerated ageing test and the lots 3 and 4 in the tetrazolium test. We were able to classify lots at different vigour levels (Table 1).

In the lots of the variety 2, by germination test, we could verify that all lots presented values above 80 %. In the same way as for the variety 1, it was possible to classify the 8 lots of the variety 2 into two levels, where the best results were observed for the lots 4 and 7. Such result is similar to the seedling emergence in the field test. The lots 4, 6, 7, and 8 obtained the best results. However, the emergence rate was lower, and all lots were below an 80 % average (Table 2).

By the accelerated ageing test, we classified the lots into two levels, where the lots 3, 4, 7, and 8 obtained the best results. In this test, the lots 1, 2, 5, and 6 presented a mean below 80 %, thus classifying it as presenting medium/low vigor.

For the cold test, we observed that the lots 3, 4, and 7 obtained the best vigor indexes. In the tetrazolium test, there was a great variation among results, both for vigor index and for viability, allowing a classification into four vigor levels, by which the lot 4 obtained the best result.

By a characterization of the physiological quality of the lots of soybean varieties, we could determine that (a) for the variety 1, the lots 1, 2, 5, 6, and 7 were classified as presenting a high vigour in all

Table 1. Mean results obtained by evaluation of the physiological qualityof soybean seeds from eight lots of the variety 1

		0		1		
Treatment	G	FE	AA	СТ	TZ Vig	TZ Viab
V1L1	93.00	94.00	92.00	92.50	93.00	95.00
	a	a	a	a	a	a
V1L2	99.00	90.00	93.00	96.00	88.00	95.00
	a	a	a	a	a	a
V1L3	85.00	85.50	82.00	88.50	72.00	75.00
	b	b	b	b	b	b
V1L4	81.50	92.00	79.50	88.00	71.00	75.00
	b	a	b	b	b	b
V1L5	97.50	91.50	96.00	92.50	91.00	93.00
	a	a	a	a	a	a
V1L6	95.50	91.00	86.50	95.00	92.00	95.00
	a	a	b	a	a	a
V1L7	97.00	89.50	97.50	95.50	96.00	97.00
	a	a	a	a	a	a
V1L8	84.00	83.50	76.00	84.00	84.00	87.00
	b	b	b	b	a	a
CV	4.33	4.17	7.57	6.08	6.86	3.89

Means followed by the same lowercase letter in rows belong to the same group, according to the Scott-Knott test at 5 % probability; V1: variety 1; L(n): lots from 1 to 8; G: germination standard test; FE: field emergence; AA: accelerated aging; CT: cold test; TZ Vig: Tetrazolium vigor; TZ Viab: Tetrazolium viability.

characterization tests, with the exception of the accelerated ageing test, and the lots 3, 4, and 8 were classified as presenting medium vigor; (b) for the variety 2, the lots 4 and 7 were classified as presenting a high vigor in all characterization tests, with the exception of the tetrazolium test for the lot 7. The lots 1, 3, 5, 6, and 8 presented a medium vigor and the lot 2 presented a low vigor.

The means of the results by manual tests and by the GroundEye software for variety 1 are shown in Table 3. As a result of the accelerated ageing test, it was possible to classify the lots into two groups, where the highest values were found for the lots 1, 2, 5, and 7. The result of the automatic vigor index also indicated lot 6 as more vigorous. This result was similar to that verified for the characterization of lots, where the lots 1, 2, 5, 6, and 7 were characterized as presenting a high vigor.

Comparing the results of the accelerated ageing test with the AV values of the software, we noted that there were no differences between the two methods of evaluating lot vigor, that is, the evaluation performed by the software can be statistically compared with accelerated ageing test data (Table 3).

The results for seedling and root length, by manual measurement and GroundEye software, indicated the lots 2, 5, and 6 as the most vigorous. We observed that there is no statistical difference between the two types of analysis for all lots. This evidences the efficiency of the analysis performed by the GroundEye software.

The means of the results by manual tests and by the GroundEye software for variety 2 are shown in Table 4. The results of automatic vigor index indicate

Table 2. Mean results obtained by evaluation of the physiological quality of soybean seeds from eight lots of the variety 2 $\,$

Treatment	G	FE	AA	ст	TZ Vig	TZ Viab
V2L1	87.00	70.00	76.00	83.00	84.00	84.00
	b	b	b	b	b	c
V2L2	85.00	71.50	73.00	81.00	67.00	71.00
	b	b	b	b	d	d
V2L3	87.00	67.50	84.00	87.00	78.00	83.00
	b	b	a	a	c	c
V2L4	97.50	77.00	92.00	91.00	97.00	99.00
	a	a	a	a	a	a
V2L5	88.00	70.00	79.00	84.00	77.00	80.00
	b	b	b	b	c	c
V2L6	80.00	78.50	78.00	84.50	85.00	86.00
	b	a	b	b	b	c
V2L7	94.00	79.00	90.00	93.00	90.00	91.50
	a	a	a	a	b	b
V2L8	84.50	76.50	86.50	84.50	84.00	86.00
	b	a	a	b	b	c
CV	5.01	5.16	7.66	5.67	4.83	4.58

Means followed by the same lowercase letter in rows belong to the same group, according to the Scott-Knott test at 5 % probability; V2: variety 2; L(n): lots from 1 to 8; G: germination standard test; FE: field emergence; AA: accelerated aging; CT: cold test; TZ Vig: Tetrazolium vigor; TZ Viab: Tetrazolium viability. the lots 4, 7, and 8 as the most vigorous. We observed the same result for seedling and root length using the GroundEye software and for root length by manual measurement. These results were different from the results of the accelerated ageing test, as it also classified lot 3 as the most vigorous. By the manual measurement of seedling length, the lots 4, 5, 6, 7, and 8 obtained the best results.

Despite a greater variation between the results of the lots of the variety 2, all tests evidenced that the lots 4 and 7 presented the best results, confirming their superiority as described in the characterization of lots of the variety 2. This variation contributed positively to the evaluation of GroundEye efficiency, since its efficiency was observed even for medium/ low vigor lots.

In addition to the comparison between lots, a comparison was made between the types of test (manual and automatic). No statistical differences were shown variety 2.

Discussion

The uniformity of water content among seed lots is of paramount importance to obtain consistent results, since differences greater than two to three percentage points may interfere with the results of vigor tests (Marcos-Filho *et al.*, 2009). This is because the rate of water uptake by seeds exerts an influence on such tests.

In both varieties, the results above 80 % for germination are within the minimum standards for the commercialization of soybeans in Brazil (Brasil, 2013).

Table 3. Mean values obtained by traditional tests and via GroundEye for eight lots of variety 1 $\,$

Lots	AA	AV	SL	GE SL	RL	GE RL
V1L1	92.00 aA	90.50 aA	7.03 bA	6.86 bA	4.63 bA	3.84 bA
V1L2	93.00 aA	96.00 aA	8.87 aA	8.93 aA	5.92 aA	5.05 aA
V1L3	82.00 bA	79.00 bA	6.62 bA	6.74 bA	4.27 bA	4.13 bA
V1L4	79.50 bA	76.00 bA	7.45 bA	7.07 bA	4.46 bA	4.13 bA
V1L5	96.00 aA	97.00 aA	9.61 aA	9.01 aA	6.19 aA	5.44 aA
V1L6	86.50 bA	92.50 aA	9.32 aA	9.36 aA	5.92 aA	5.27 aA
V1L7	97.50 aA	94.50 aA	7.21 bA	7.01 bA	4.99 bA	4.07 bA
V1L8	76.00 bA	74.50 bA	7.01 bA	6.37 bA	4.63 bA	3.78 bA
CV	7.57	6.69	6.91	10.91	7.76	14.57

Means followed by the same lowercase letter in rows and upper-case letters in lines belong to a same group according to the Scott-Knott test at 5 % probability level; V1: variety 1; L(n): lots from 1 to 8; AA: accelerated aging; AV: automatic vigor; SL: seedling length by manual measurement; RL: root length by manual measurement; GE SL: seedling length by GroundEye; GE RL: root length by GroundEye.

Due to the proximity of field emergence and germination results (Table 1), it is possible to infer that the lots of the variety 1, besides having a high vigor, did not encounter adverse field conditions, expressing their full potential.

According to França-Neto and Krzyzanowski (2018), in the tetrazolium test, values above 85 % are for lots with a very high vigor, between 84 % and 75 % for lots with a high vigor, between 74 and 60 % for lots with a medium vigor, between 59 % and 50 % for lots with a low vigor, and below this, for lots with a very low vigor. In this context, the lots 1, 2, 5, 6, and 7 are very vigorous lots, the lot 8 has a high vigor and the lots 3 and 4 have a medium vigor (Table 1).

Considering the results obtained for the variety 1 in all tests, the lots with the lowest physiological quality values are lots 3, 4 and 8. These lots had the greatest variation among the results. This variation is more visible in lots with a medium and low physiological potential, since the process of deterioration of a seed lot affects each individual seed, making this variation more visible (Pinto *et al.*, 2015).

For variety 2, the percentage of emergence of seedling in the field was smaller than the germination, which can be associated to the lower vigor of the lot once the means of the vigor tests of such variety were smaller and varied more when compared to variety 1. This data corroborates the documented information in the literature that associates the emergence of the seedlings in the field as a vigor test (Rodrigues *et al.*, 2020). In the evaluation of the physiological potential of soybean seeds, Yagushi *et al.* (2014) found similarity between the data of emergence of seedlings in the field and the data obtained in the tetrazolium and accelerated ageing tests. Similarly, Wendt *et al.* (2017) found connection between the

Table 4. Mean values obtained by traditional tests and via GroundEye for eight lots of variety 2 $\,$

0						
Lots	AA	AV	SL	GE SL	RL	GE RL
V2L1	76.00 bA	73.00 bA	6.54 bA	6.10 bA	4.50 cA	3.69 bA
V2L2	73.00 bA	72.00 bA	6.74 bA	6.60 bA	4.32 cA	3.70 bA
V2L3	84.00 aA	81.00 bA	7.25 bA	6.86 bA	5.02 bA	3.85 bA
V2L4	92.00 aA	94.00 aA	8.28 aA	7.70 aA	5.52 aA	4.53 aA
V2L5	79.00 bA	77.00 bA	8.02 aA	6.88 bA	5.26 bA	3.96 bA
V2L6	78.00 bA	80.00 bA	7.99 aA	6.87 bA	5.04 bA	3.95 bA
V2L7	90.00 aA	94.00 aA	8.23 aA	8.04 aA	5.78 aA	4.66 aA
V2L8	86.50 aA	92.00 aA	8.04 aA	7.86 aA	5.64 aA	4.68 aA
CV	7.66	7.78	6.72	15.36	7.67	18.34

Means followed by the same lowercase letter in rows and upper-case letters in lines belong to a same group according to the Scott-Knott test at 5 % probability level. V2: variety 2; L(n): lots from 1 to 8; AA: accelerated aging; AV: automatic vigor; SL: seedling length by manual measurement; RL: root length by manual measurement; GE SL: seedling length by GroundEye; GE RL: root length by GroundEye.

results of emergence of the seedlings in the field and the results of the tetrazolium test, accelerated ageing test, electric conductivity and the analysis of carbon dioxide concentration.

In spite of the great variation among the results of characterization of the lots of variety 2, we may observe that lots 4 and 7 had the best general performance, both by statistical analysis and by determination of the numerical value of each test results (Table 2), evidencing their superior physiological quality.

Rodrigues *et al.* (2020), using the Vigor-S equipment (Seed Vigor Automated Analysis System), concluded that such equipment is efficient in the evaluation of vigor of soybean seeds. Carvalho *et al.* (2020), also using the Vigor-S equipment, successfully obtained information of the root length and the hypocotyls, seedlings length, vigor index and uniformity to the evaluation of the phyto-toxicity in soybean seeds treated with phytosanitary products in different moments of application.

The use of the GroundEye, when compared to the accelerated ageing test and the measurement tests of seedling and root length, demonstrates the precision of the equipment and the speed in which results can be obtained. According to the methodology of the accelerated ageing test (AOSA, 2009), nine days are needed for obtaining the results. As for the vigor results from the GroundEye, the analyses of the images are accomplished in seedlings from the germination test with only three days, representing a great contribution to the quality control programs of seed producing companies once there is a constant need for rapid and accurate information regarding the quality of the seed during the production process.

Conclusions

The results obtained in this study show that the computer analysis of seedling images using the GroundEye is efficient in determining the physiological potential of soybean seeds. Such evaluation can be performed on the third day after the germination test, thus reducing in six days the time needed to obtain the results, being indicated in the use of quality control programs of seed production companies.

References

- AOSA. (2009). Seed vigor testing handbook. Association of Official Seed Analysts. Lincoln, USA.
- Barros, A.S.R., Dias M.C.L.L., Cicero, S. M., & Krzyzanowski, F.C. (1999). Teste de frio. In F.C Krzyzanowski, R.D. Vieira, & J.B. França-Neto (Eds.), Vigor de Sementes: Conceitos e Testes (p. 218). Abrates, Londrina.
- Brasil. (2009). Regras para Análises de Sementes (1 edition). Ministério da Agricultura, Pecuária e Abastecimento.

Brasil. (2013). Instrução Normativa, N° 45, de 17 de setembro de 2013. Seção 1. Ministério da Agricultura, Pecuária e Abastecimento.

- Carvalho, E.R., Rocha, D.K., Andrade, D.B., Pires, R.M.O., Penido, A.C., & Reis. L.V. (2020). Phytotoxicity in soybean seeds treated with phytosanitary products at different application times. Journal of Seed Science, 42(e202042036), 1-12. https:// doi.org/10.1590/2317-1545v42237847
- França-Neto, J.B., Krzyzanowski, F.C., & Costa, N.P. (1998). O teste de tetrazólio em sementes de soja. Embrapa-CNPSo.
- França-Neto, J.B., & Krzyzanowski, F.C. (2018). Metodologia do teste de tetrazólio em sementes de soja. Documento 406. Embrapa-Soja.
- Marcos-Filho, J. (2015). Seed vigor testing: an overview of the past, present and future perspective. *Scientia Agricola*, 72(4), 363-374. https://doi.org/10.1590/0103-9016-2015-0007
- Marcos-Filho, J., Kikuti, A.L.P., & Lima, L.B. (2009). Métodos para avaliação do vigor de sementes de soja, incluindo a análise computadorizada de imagens. Revista Brasileira de Sementes, 31(1), 102-112. https://doi.org/10.1590/S0101-31222009000100012
- Medeiros, A.D., Pereira, M.D., Silva, I.R.F., Capobiango, N.P., & Flores, M.E.P. (2019). Vigor of maize seeds determined by a free image analysis system. *Revista Ciência Agronômica*, 50(4), 616-624. https://doi.org/10.5935/1806-6690.20190073
- Nakagawa, J. (1999). Testes de vigor baseados na avaliação das plântulas. In F.C. Krzyzanowski, R.D. Vieira, & J.B. França-Neto (Eds.), Vigor de Sementes: Conceitos e Testes (pp.4-26). Abrates.
- Pinto, C.A.G., Carvalho, M.L.M., Andrade, D.B., Leite, E.R., & Chalfoun, I. (2015). Image analysis in the evaluation of the physiological potential of maize seeds. Revista Ciência Agronômica, 46(2), 319-328. http://www.ccarevista.ufc.br/seer/ index.php/ccarevista/article/view/3503
- Rodrigues, M., Gomes-Junior, F.G., & Marcos-Filho, J. (2020). Vigor-S: System for automated analysis of soybean seed vigor. Journal of Seed Science, 42(e202042039), 1-12. https:// doi.org/10.1590/2317-1545v42237490
- Silva, F.A.S., & Azevedo, C.A.V. (2009). Principal components analysis in the software assistat-statistical assistance. In 7th World Congress on Computers in Agriculture, Reno. Proceedings of the 7th World Congress on Computers in Agriculture (pp.1-5). ASABE.
- USDA. (2020). World Agricultural Production. Foreign Agricultural Service. Office of Global Analysis, Circular Series WAP.
- Wendt, L., Malavasi, M.M., Lopes Dranski, J.A., Malavasi, U.C., & Gomes-Junior, F.G. (2017). Relação entre testes de vigor com a emergência a campo em sementes de soja. Revista Brasileira de Ciências Agrárias, 12(2), 166-171. https://doi.org/10.5039/ agraria.v12i2a5435
- Yagushi, J.T., Costa, D.S., & França-Neto, J.B. (2014). Saturated salt accelerated aging and computerized analysis of seedling images to evaluate soybean seed performance. *Journal of Seed Science*, 36(2), 213-221. https://doi.org/10.1590/2317-1545v32n2935