CASE PRESENTATION

Disseminated toxoplasmosis in a heart transplant patient despite co-trimoxazole prophylaxis: A case report

Victoria Dávila1, Gustavo Roncancio-Villamil1,2, Luis Alfonso Correa3,4, Catalina Restrepo4, Camilo Alberto Madrid1, Javier Mauricio González1

1 Servicio de Enfermedades Infecciosas, Clínica Cardio VID, Medellín, Colombia
2 Escuela de Ciencias de la Salud, Universidad Pontificia Bolivariana, Medellín, Colombia
3 Servicio de Patología, Laboratorio Clínico VID, Medellín, Colombia
4 Sección de Dermatología, Departamento de Medicina Interna, Facultad de Medicina, Universidad de Antioquia, Medellín, Colombia

We report the case of a 61 year-old male who underwent heart transplantation eight months before developing a systemic condition with central nervous system, lung, kidney, colonic, cutaneous, and hematologic involvement, found to be secondary to a systemic toxoplasmosis despite co-trimoxazole prophylaxis in a previous-to-transplant seronegative patient receiving a heart from a seropositive donor. A review of prophylactic options in our environment is discussed.

Key words: Toxoplasmosis; heart transplantation; immune tolerance; seroconversion.

doi: https://doi.org/10.7705/biomedica.v34i2.3189

Toxoplasmosis diseminada en un paciente con trasplante de corazón a pesar de la profilaxis con trimetoprim-sulfametoxazol

Se reporta el caso de un paciente de sexo masculino, de 61 años de edad, quien ocho meses después de someterse a un trasplante de corazón presentó una enfermedad sistémica con compromiso del sistema nervioso central y del sistema inmunológico, así como de pulmón, riñón, colon y piel, y a quien finalmente se le diagnosticó toxoplasmosis diseminada, a pesar de haber recibido profilaxis con trimetoprim-sulfametoxazol, debido a que el órgano provenía de un donante positivo para toxoplasmosis siendo él un receptor negativo. Se discuten las opciones de profilaxis en nuestro medio.

Palabras clave: toxoplasmosis; trasplante de corazón; tolerancia inmunológica; seroconversión.

doi: https://doi.org/10.7705/biomedica.v34i2.3189

Toxoplasmosis is an infection caused by the protozoan *Toxoplasma gondii*. It is prevalent worldwide, affecting approximately one third of the world’s population (1). Seroprevalence increases with age and there is no gender difference.

Toxoplasmosis is transmitted via oral intake of food contaminated with the parasite’s cysts or oocysts. However, there are reports of transmission through transplanted organs and blood transfusions, as well as vertical transmission (2).

It evolves to a systemic disease, and only 10% of the affected population is symptomatic. The clinical course is usually benign, but it can be lethal, especially in immunosuppressed patients (3). Systemic clinical signs include generalized adenopathies, myocarditis, polymyositis, pneumonitis, encephalitis and chorioretinitis. Skin manifestations are rare, though usually present in immunosuppressed patients. Given the great variability in the appearance of cutaneous toxoplasmosis, histopathological studies are required for diagnosis.

Although direct observation of the parasite is possible in most tissue samples, diagnosis should be confirmed by immunohistochemistry and polymerase chain reaction (PCR) (4).

Medical therapy is indicated when there is active skin or ocular infection, or congenital disease, and in all immunosuppressed patients. Sulfadiazine in combination with pyrimethamine is the treatment...
Seronegative heart transplant patients should receive prophylaxis with pyrimethamine and sulfadiazine for the first three months followed by co-trimoxazole for the rest of their lives (5,6).

**Case report**

We report the case of a 61-year-old male patient with a past medical history of hypertension, dyslipidemia, hypothyroidism, heart failure with preserved ejection fraction and severe coronary artery disease, in whom surgical and percutaneous revascularization were performed. After surgery, the patient developed refractory angina despite optimal anti-ischemic medication which led to heart transplantation. Laboratory tests previous to transplant were discordant for *T. gondii* and human herpes viruses type I and II. The donor had positive immunoglobulin M (IgM), negative immunoglobulin G (IgG) for *T. gondii*, and positive IgG for herpes viruses (HV) type I and type II. The receptor was negative for both.

After eight months of transplant the patient was admitted with persistent diarrhea lasting 20 days, abdominal pain and malaise. The immunosuppression regimen included mycophenolate mofetil, 1 g every 12 hours, prednisolone, 5 mg per day, and cyclosporine, 150 mg every 12 hours. Prophylaxis with co-trimoxazole, 160/800 mg per day, was also given.

On physical examination the patient presented with tachycardia, normal blood pressure and respiratory rate, and no fever was documented. Vesicular lesions with clear content and erythematous base with dermatome distribution were seen in the right gluteus.

Initial laboratory tests showed anemia, leukopenia with lymphopenia, renal dysfunction and a positive Tzank test (table 1). Viral load for cytomegalovirus (CMV) was positive with 404,000 copies/ml; therapy with ganciclovir, 5 mg/kg every 12 hours, was started with initial improvement of the diarrhea.

However, two weeks later the patient persisted with leukopenia, the anemia worsened requiring red blood cell (RBC) transfusion and he developed fever as high as 39.8º C. The only additional finding was four 5 mm to 1 cm erythematous, mildly painful lesions, some in legs and thighs with white scaling in the center, and a similar lesion in the upper right eyelid (figure 1). The diarrhea reappeared, renal dysfunction worsened, as well as the complete blood count (CBC), now with pancytopenia (anemia, leukopenia, lymphocytopenia, neutropenia and thrombocytopenia).

Empirical therapy with anidulafungin plus cefepime was started for febrile neutropenia. *Toxoplasma* antibody seroconversion was documented. Therapy with clindamycin plus pyrimethamine was started and the skin lesions were biopsied.

**Table 1. Diagnostic aids**

<table>
<thead>
<tr>
<th>Laboratory tests on admission</th>
<th>Creatinine 2.7 mg/dl, BUN 53 mg/dl</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>CBC (leucocytes 3100, PMN 83%, lymphocytes 13%, hemoglobin 9.4, platelet count 250,000)</td>
</tr>
<tr>
<td></td>
<td>AST 37 ALT 30 FA 80 TB 0.4 DB 0.3</td>
</tr>
<tr>
<td></td>
<td>Normal urinalysis</td>
</tr>
<tr>
<td></td>
<td>Coprological examination with scarce leucocytes</td>
</tr>
<tr>
<td>Microbiology</td>
<td>Negative coproculture</td>
</tr>
<tr>
<td>Day 5-30</td>
<td>Negative blood cultures for fungus, bacteria, and mycobacteria</td>
</tr>
<tr>
<td>Serological and molecular</td>
<td>Latex agglutination test negative for Cryptococcus neoformans</td>
</tr>
<tr>
<td>tests</td>
<td>Hystoplasma capsulatum urinary antigen: Negative</td>
</tr>
<tr>
<td>Day 5-30</td>
<td>Serum galactomannan: Negative (Platelia)</td>
</tr>
<tr>
<td></td>
<td>Serum CMV viral load: 404,000 copies/ml</td>
</tr>
<tr>
<td>Histopathology</td>
<td>Colonoscopy: Active chronic inflammation with erosive component. Negative histochemistry colorations</td>
</tr>
<tr>
<td>Day 20</td>
<td>Bone marrow biopsy: Hypocellular hematopoietic bone marrow without granuloma or crythrocytic changes. Special colorations negative for organisms. Cultures were negative for fungus, bacteria and mycobacteria.</td>
</tr>
<tr>
<td>Other tests</td>
<td>Normal IgA, IgE, and IgM levels. Low IgG levels (593 mg/dl N: 700.6-1600 mg/dl).</td>
</tr>
<tr>
<td>Day 20</td>
<td>CSF studies: PCR for CMV, Epstein-Barr virus (EBV) and mycobacteria were negative. PCR for Toxoplasma was positive</td>
</tr>
<tr>
<td></td>
<td>Low serum ADAMS-13 levels</td>
</tr>
<tr>
<td>Laboratory tests on discharge</td>
<td>Creatinine 1.4 mg/dl, BUN 24 mg/dl</td>
</tr>
<tr>
<td></td>
<td>CBC (leucocytes 5.300, PMN 75%, lymphocytes 20%, hemoglobin 9.4, platelet count 229,000)</td>
</tr>
<tr>
<td></td>
<td>PCR for Toxoplasma was positive.</td>
</tr>
</tbody>
</table>

BUN: Blood urea nitrogen; PMN: Polymorphonuclear; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; AP: Alkalin phosphatase; TB: Total bilirubin; DB: Direct bilirubin
Histopathology of skin lesions was positive and immunohistochemistry for toxoplasmosis confirmed the presence of the parasite (figures 2 and 3). Molecular studies with blood and cerebrospinal fluid (CSF) PCR were also positive for *Toxoplasma* (7). Central nervous system magnetic resonance (MR) did not show focal lesions.

The initial course was unfavorable, the patient persisted with fever, headache, encephalopathy, oliguric renal failure, hepatitis, pancytopenia, metabolic acidosis and increased lactate dehydrogenase (LDH) up to 2,500 IU/L. Pulmonary patchy infiltrates were found. He was admitted to the intensive care unit for mechanical ventilation. Plasma exchange was started for suspected thrombotic thrombocytopenic purpura.

After a few days the patient’s status improved, especially renal function and blood counts, but he persisted with hypogammaglobulinemia. CMV infection relapsed with poor response to ganciclovir. Gammaglobulin, 30 g intravenous (IV) weekly, was started with complete remission.

He was released after six weeks of therapy with clindamycin and pyrimethamine upon suspicion of co-trimoxazole resistance. Suppression with co-trimoxazole plus valganciclovir and gammaglobulin was maintained and the patient has been symptom-free for 28 months.

**Discussion**

This is a heart transplant patient who developed systemic toxoplasmosis with central nervous system, pulmonary, renal, colonic, cutaneous, and hematologic compromise secondary to disseminated toxoplasmosis. *Toxoplasma gondii* infection after heart transplant is often a severe disease. The clinical presentation ranges from chorioretinitis to disseminated infection (pulmonary, myocardial, hepatic and CNS infection) (8).

Toxoplasmosis in immunocompromised patients is usually secondary to a reactivation of latent infection from the donor (9). In this case, previous to transplantation the donor’s toxoplasmosis serology was discordant with the recipient’s. *Toxoplasma* cysts may persist in the donor’s myocardium, thus, hosts are at higher risk of developing the disease. With immunosuppression, the bradyzoites become tachyzoites, quickly proliferating and consequently destroying cells.

Before the use of prophylaxis with co-trimoxazole there was a higher risk of disseminated toxoplasmosis (10). Studies of seronegative heart

---

**Figure 1.** A 5 mm erythematous papule with white scaling in the center

**Figure 2.** Skin histology from the lower limb showing *T. gondii* tachyzoites forming a pseudocyst. Giemsa stain, 40X.

**Figure 3.** Special immunohistochemical staining for toxoplasma from the skin biopsy showing a pseudocyst. Toxoplasma RB-9423-P rabbit monoclonal antibodies (Thermo®) were used.
recipients have found that the risk of *T. gondii*-associated clinical disease ranges from 0 to 82% if patients receive organs from seropositive donors without antimicrobial prophylaxis (6,8).

Seronegative heart transplant patients should receive prophylaxis. Effective regimens include one single-strength co-trimoxazole tablet daily, one double-strength three times a week, 1,500 mg atovaquone daily or pyrimethamine 25 mg/day. This patient had received prophylaxis with cotrimoxazole. Results from observational studies have shown that this drug is effective in preventing toxoplasmosis, even in cardiac recipients (6,11).

The patient may have developed severe disease despite co-trimoxazole prophylaxis due to resistance to sulfonamide (12,13). In this sense, more-virulent *Toxoplasma* strains have been found in Colombia and transmission of toxoplasmosis by renal transplant was reported in two local cases (14,15). This suggests that the risk could be higher in tropical zones of the Americas where a high genetic diversity of *Toxoplasma* has been found.

Co-trimoxazole is provided as a 1/5 ratio combination of trimethoprim and sulfamethoxazole. This ratio was chosen in order to provide synergistic ratio against bacteria, but due to the poor efficacy of trimethoprim against *T. gondii* trophozoites (100 times lower than pyrimethamine), the optimal ratio of the two compounds against this infection is probably closer to 1/1 (16). High doses of co-trimoxazole are needed to cure experimental acute infection in mice, and relapses are observed after cessation of therapy, because it is ineffective on the cyst form of the parasite. However, it has proved remarkably efficient in the treatment and prophylaxis of toxoplasmosis in transplant patients with AIDS (11).

Considering that a high genetic diversity, gene polymorphisms in the dihydrofolate reductase and dihydropteroate synthase genes, more-virulent strains of *T. gondii*, and several transplanted patients with disseminated toxoplasmosis have been reported in Colombia, the recommended doses of co-trimoxazole could be insufficient, or another agent (e.g., dapsone plus pentamidine or pyrimethamine and sulfadiazine) could be necessary to prevent toxoplasmosis in high risk transplant patients.

It is also important to point out that this patient developed symptoms after eight months of transplantation. Previous reports have described how heart transplant patients receiving the organ from seropositive donors usually develop infections four to twelve weeks after transplantation. According to previous reports, the organs most commonly affected by systemic toxoplasmosis in immunosuppressed patients are the lungs and the central nervous system (17,18).

To our knowledge, this is the first heart transplant patient reported in the literature presenting toxoplasmosis with cutaneous manifestations. Such cutaneous involvement is quite rare, and only a few cases have been reported in immunocompromised patients with great variability in the appearance of skin lesions. Generalized rash, erythema multiforme-like lesions, nodular lesions and vesicular varicella-like lesions have been described (19-21). Skin biopsies confirmed the presence of the parasite, but histopathological findings are variable. The involvement of the colon is also unique in this patient.

We can conclude, then, that diagnosing toxoplasmosis in immunosuppressed patients is challenging, as any organ may be affected. Thus, toxoplasmosis should always be suspected as a differential diagnosis in patients with systemic symptoms. Patients must be studied thoroughly, using all possible diagnostic aids to rule out the disease, including serology, histology, special immunohistochemical staining and PCR (22-23). Co-trimoxazole prophylaxis does not exclude the diagnosis and resistance studies are necessary.

**Conflict of interests**

The authors declare no conflict of interests.

**Funding**

No funding was received.

**References**


