

# NEW ANDEAN BEE SPECIES OF *CHILICOLA* SPINOLA (HYMENOPTERA: COLLETIDAE, XEROMELISSINAE) WITH NOTES ON THEIR BIOLOGY

## Especies nuevas de abejas andinas *Chilicola* Spinola (Hymenoptera: Colletidae, Xeromelissinae) con notas sobre su biología

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### ABSTRACT

We describe and illustrate three new bee species of the genus *Chilicola* Spinola that occur at high altitudes in the Eastern Andes of Colombia: *C. (Anoediscelis) paramoides* Gonzalez sp. nov., *C. (Hylaeosoma) bochica* Gonzalez sp. nov., and *C. (Oroediscelis) deborahae* Gonzalez sp. nov. We also provide biological notes and discuss the phylogenetic relationships of *C. deborahae*; nests of this species were found inside dry flower stems of *Espeletia argentea* Humb. & Bonpl. (Asteraceae) and a palynological analysis showed that 80 % of the pollen grains found in brood cells belonged to *Arcytophyllum* aff. *nitidum* and *A. muticum* (Rubiaceae). The ichneumonid wasp *Grotea* sp. is also recorded parasitizing brood cells of *C. deborahae*.

**Key words.** Apoidea, Colombia, *Grotea*, Páramo, Systematics.

### RESUMEN

Describimos e ilustramos tres especies nuevas de abejas del género *Chilicola* Spinola que habitan en grandes alturas en la cordillera Oriental de Colombia: *C. (Anoediscelis) paramoides* Gonzalez sp. nov., *C. (Hylaeosoma) bochica* Gonzalez sp. nov., and *C. (Oroediscelis) deborahae* Gonzalez sp. nov. También presentamos notas biológicas y discutimos la relación filogenética de *C. deborahae*; los nidos de esta especie fueron encontrados dentro de inflorescencias secas de *Espeletia argentea* Humb. & Bonpl. (Asteraceae) y análisis palinológicos mostraron que el 80% de los granos de polen encontrados en las celdas de cría pertenecían a *Arcytophyllum* aff. *nitidum* y *A. muticum* (Rubiaceae). También se registra la avispa ichneumonida *Grotea* sp. parasitando celdas de cría de *C. deborahae*.

**Palabras clave.** Apoidea, Colombia, *Grotea*, Páramo, Sistemática.

### INTRODUCTION

The diverse tropical Andean bee fauna is still poorly known taxonomically and ecologically (Gonzalez & Engel 2004,

Gonzalez *et al.* 2005). Three subgenera (*Anoediscelis* Toro & Moldenke, *Hylaeosoma* Ashmead, and *Oroediscelis* Michener) and 28 species of *Chilicola* Spinola are known to occur in the Andes from Venezuela to

Peru. Michener (2002) revised these species and explored the phylogenetic relationships of the 12 species of *Oroediscelis*, the only subgenus restricted to the Andes. Gonzalez & Michener (2004) described *C. paramo*, a species of *Anoediscelis* that did not show a close relationships to any of the remaining known species of the subgenus; they also provided an updated key to the Andean subgenera.

We describe and illustrate one species of each Andean subgenus of *Chilicola*. These new species occur in Páramos from the Eastern Andes of Colombia, and one of them is closely related to *C. paramo*. We also provide biological notes and discuss the phylogenetic relationships of *C. deborahae* sp. nov.

## MATERIALS AND METHODS

The morphological description and illustrations were made using an Olympus SZ60 stereomicroscope. Morphological terminology follows that of Michener (2007). Abbreviations used in the descriptions are: F, S, T, OD and PD for antennal flagellomere, metasomal sternum and tergum, ocellar diameter, and puncture diameter, respectively. Type specimens are deposited in the Instituto de Ciencias Naturales (ICN), Universidad Nacional de Colombia, Bogotá, Colombia, and the Snow Entomological Museum (SEMC), University of Kansas, Lawrence, Kansas, USA.

To explore the phylogenetic relationships of *C. deborahae* sp. nov., we used the morphological characters and data set of Michener (2002) for the phylogeny of *Oroediscelis* (two outgroup species, 12 ingroup species, and 16 characters). We analyzed this data set using the *wh\** and *max\** commands in *Nona* (Goloboff 1993); all characters were considered

non-additive. Trees were visualized and printed using *Winclada* (Nixon 1999, slow optimization). We only explored the phylogenetic relationships of this species because the phylogeny of the subgenera *Anoediscelis* and *Hylaeosoma* have not yet been studied.

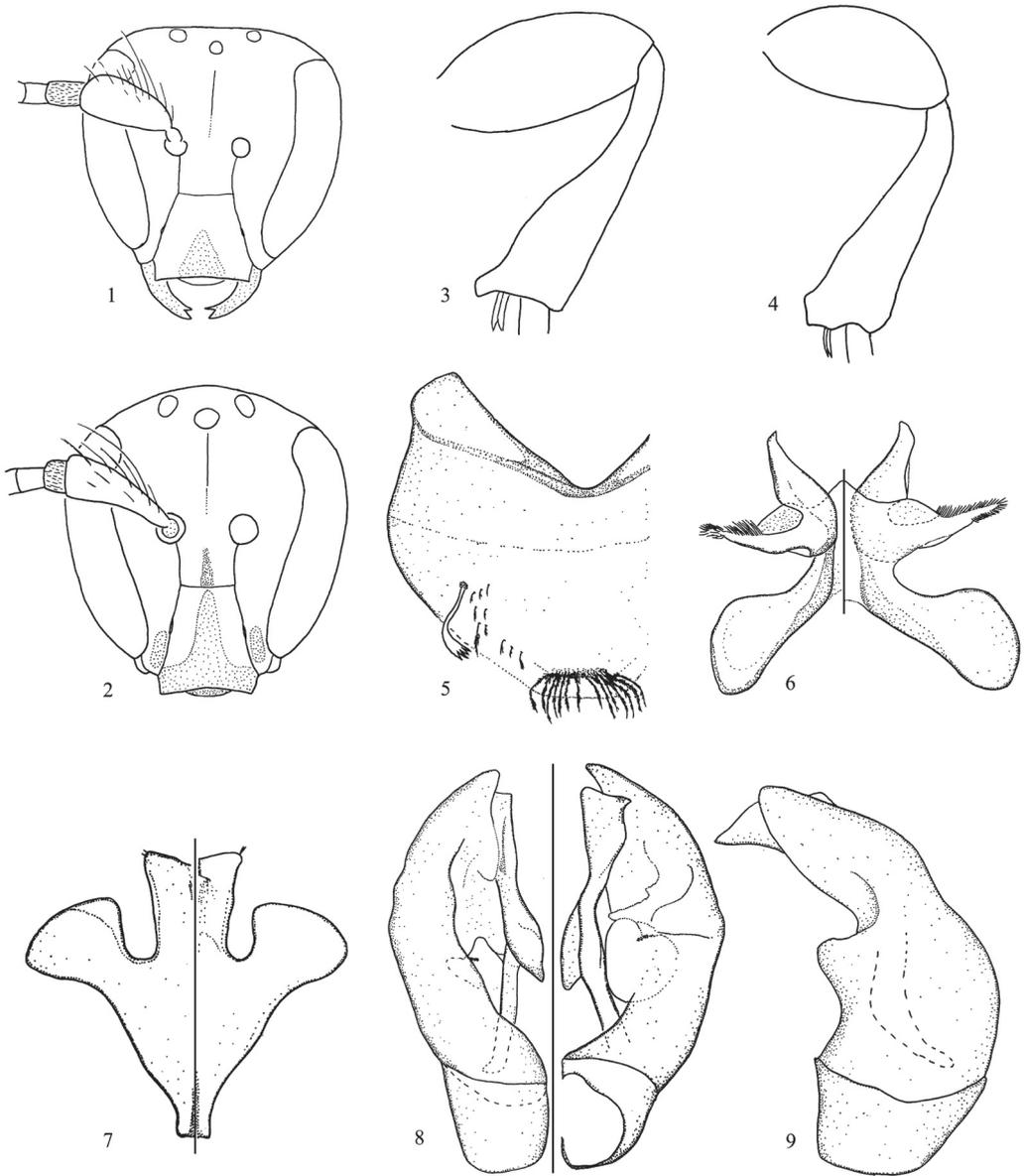
Two nests of *C. deborahae* sp. nov. were found on August 23, 2003, at the Santuario de Fauna y Flora de Iguaque, 5°70'N, 73°46'W (Departamento of Boyacá). The nests were kept inside Ziplock plastic bags containing moistened cotton for about four weeks until adult emergence. Pollen samples from brood cells were treated and preserved following the acetolysis method of Erdtman (1986) and then mounted on glass slides. Pollen samples were examined at 40X and 100X magnification with a Leitz DIALUX 22 EB microscope, and are deposited in the palynological collection of the ICN.

## RESULTS

### *Chilicola (Anoediscelis) paramoides* Gonzalez sp. nov.

Figs. 2, 4–9

**Diagnosis.** This species is closely related to *C. paramo*. Both species differ from the species placed in the *C. ashmeadi* group by Michener (2002) in the distal stigmal perpendicular crossing submarginal cells near first submarginal crossvein, long scape (Figs. 1, 2), and the swollen hind femur of the male (Figs. 3, 4). *C. paramoides* sp. nov. can be separated from *C. paramo* by the shinier integument of frons, more extensive yellow markings on the face and legs, longer and narrower scape, shorter pedicel (Fig. 2), and the broader apical projection of the hind tibia (Fig. 4); the S6–S8 and genital capsule are also different (compare with Figs. 4–8 of Gonzalez & Michener 2004).



**Figures 1–9.** Male of *Chilicola paramo* (1, 3) and *C. paramoides* sp. nov. (2, 4–9). 1, 2 = face showing integumental color. Stippling indicates yellow, remainder black; 3, 4 = hind femur and hind tibia (profile view); 5–9 = S6, S7, S8 and genital capsule. In divided figures, dorsal view is shown on left, ventral on right.

**Description. Male.** Body length 4.6 mm; forewing length 3.6 mm. **Structure.** Head 1.1x longer than broad; interalveolar distance 2x OD, 1.3x greater than alveolorbital distance; interocellar distance 2.4x OD, 1.1x

ocellocular distance; ocelloccipital distance about 1.4x OD; scape 3x longer than broad (Fig. 2); pedicel subcylindrical, 1.1x longer than broad; F1 1.4x longer than broad, narrower and slightly longer than pedicel,

shorter than F2 and F3 individually, F2 and F3 each 1.3x longer than broad, F4–F10 progressively broader and longer, F11 1.5x longer than broad. Hind femur swollen, 1.8x longer than broad, hind tibia with apical projection broader than in *C. paramo* (Fig. 4). S6–S8, and genital capsule as in Figs. 5–9. T7 with distal margin rounded. **Coloration.** Head and mesosoma mainly black except for the following yellow parts: maxillary palpus, hypostomal area, labrum, and mandible, except for red brown apex; clypeus, supraclypeal and lower paraocular areas as in Fig. 2.; antenna dark brown with apex of scape, pedicel, and ventral surface of flagellum yellowish; legs dark brown with apex of femora, outer surface of front tibia, and base and apex of remaining tibiae yellow; tarsi mostly yellow with outer surface of distal segments brownish; tegula dark brown; wings faintly smoky, veins and stigma dark brown; metasoma mainly dark brown, terga and sterna with translucent brownish distal margins. **Pubescence.** Whitish to light yellow; predominantly short and sparse, longer (2.0–2.5x OD) on frons, scape, vertex, anterior surface of mesepisternum, sides of propodeum and terga, and distal margin of sterna; pedicel densely covered by very short (0.5x OD), semierect hairs. **Punctuation.** Head and mesosoma finely imbricate, punctures stronger and denser ( $\leq$  PD) on frons and vertex; basal area of propodeum weakly striate, striae ending before smooth and shiny distal margin, integument otherwise granular; metasoma finely lineolate with minute and sparse punctures.

**Variations.** In all paratypes the supraclypeal area is black; in one of them, the yellow markings of clypeus and lower paraocular area are also slightly more reduced than the holotype.

**Holotype.** ♂, COLOMBIA. *Boyacá*: Arcabuco Prov., Santuario de Fauna y Flora Iguaque, Chaina, 2600 m, 16 March 2001,

V. Gonzalez [ICN]. **Paratypes.** 4♂, two specimens with the same data as the holotype, one of them collected on June 20, 2001; remaining paratypes with the following data: *idem*, Carrizal, 3350 m, June 23 2000 [ICN, SEMC].

**Etymology.** This species is named using the Greek suffix “-oides”, meaning “having the appearance of” or “like”, referring to its resemblance to *C. paramo*.

**Comments.** This species and *C. paramo* represent a distinct group of Andean *Anoediscelis* easily recognized by the characters mentioned in the diagnosis. The group has been fully characterized by Gonzalez & Michener (2004, p. 28).

***Chilicola (Hylaeosoma) bochica* Gonzalez sp. nov.**

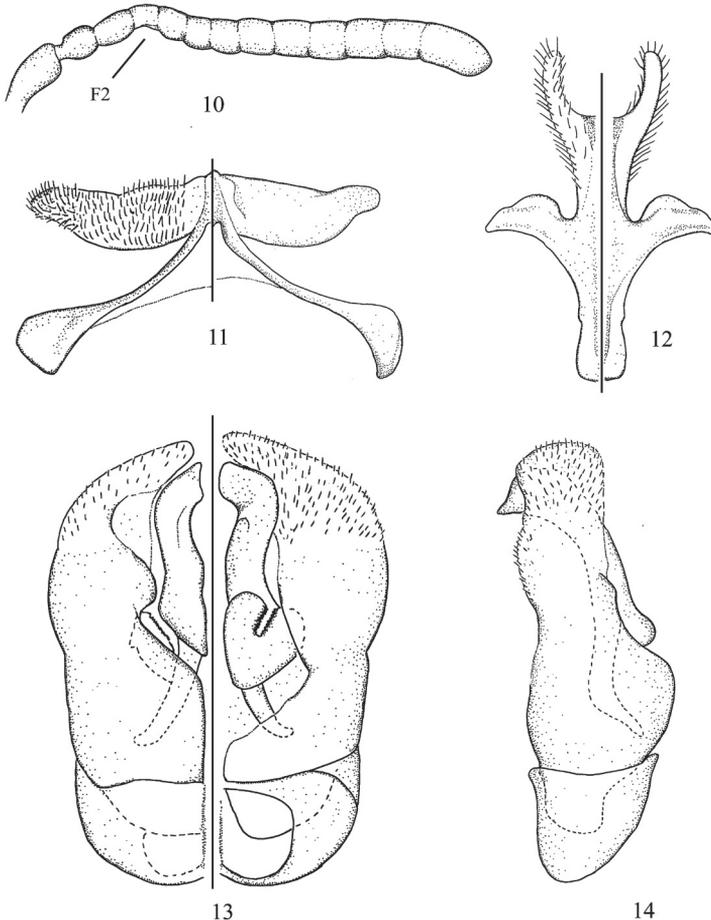
Figs. 10–14

**Diagnosis.** In the key of Michener (2002) to the Andean species of *Hylaeosoma*, this species runs to *C. canei* Michener. However, the male of *C. bochica* sp. nov. can be separated from that species, and any other species of the subgenus, by the shape of S7 and S8 (Figs. 11, 12), and the modified antenna, with F2 ventrally concave and F3–F5 slightly crenulate (Fig. 10); the female can be separated from *C. canei* by the basal area of the propodeum with a strong median stria and the first recurrent vein basad to first submarginal crossvein.

**Description. Male.** Body length 3.9–4.2 mm; forewing length 3.2 mm. **Structure.** Head 1.1x longer than broad; interalveolar distance 2.3x OD, 1.3x greater than alveolorbital distance; interocellar distance 2.1x OD, about as long as ocellocular distance; ocelloccipital distance about 1.2x OD; scape claviform, 1.7x longer than broad; pedicel subcylindrical, 1.3x longer than broad; F2 ventrally concave, F3–F5 slightly crenulate, remaining flagellomeres

normal (Fig. 10); F1 1.1x longer than broad, slightly shorter than pedicel; F2 and F3 each about as long as wide, F4–F10 progressively broader and longer, F11 1.6x longer than broad, distinctly shiny apically. Legs unmodified. S2–S4 with distinct preapical sublateral tubercles, more conspicuous on S4; S5 with strong preapical transverse ridge (clearly visible in lateral view); S7, S8, and genital capsule as in Figs. 11–14. T2 and T3 with pregradular areas distinctly depressed; T7 with distal margin rounded. **Coloration.** Head and mesosoma mainly black except for the following yellow parts: maxillary palpus, mandible (except for red brown apex), and labrum; clypeus with

an inverted T-shaped maculation; antennal flagellum dark brown with ventral surface yellow, except for apex of F11 black; legs dark brown with apex of femora, front tibia, and base and apex of remaining tibiae yellow; tarsi mostly light brown; tegula yellowish; wings faintly smoky, veins and stigma dark brown; metasoma mainly dark brown, terga and S1–S5 with translucent brownish distal margins; S6 yellowish. **Pubescence.** Whitish to light yellow, hairs denser on lower paraocular area, depression for antennal scape, vertex, and gena; scape uniformly covered by short hairs ( $\leq$  OD); mesepisternum and sterna with longer hairs than on head (1.5–2.0x OD). **Punctuation.**



**Figures 10–14.** Male of *Chilicola bochica* sp. nov. 10 = antenna (profile view); 11–14 = S7, S8, and genital capsule. In divided figures, dorsal view is shown on left, ventral on right.

Head and mesosoma dull, finely imbricate between punctures, punctures stronger and denser ( $\leq$ PD) on frons and vertex; lower gena lineolate, hypostomal area shiny; basal area of propodeum with several irregular longitudinal striae, integument otherwise imbricate; metasoma shiny, finely lineolate with minute and sparse (1–2x PD) punctures.

**Female.** As described for the male, except for: **Structure.** Head 1.2x longer than broad; interalveolar distance 3.0x OD, 1.6x greater than alveolorbital distance; interocellar distance 2.3x OD, 0.8x ocellocular distance; ocelloccipital distance about 1.6x OD; scape 2.9x longer than broad; pedicel slightly longer than broad; flagellomere normal, F1 longer than broad, about as long as combined length of F2 and F3. **Coloration.** Labrum and clypeus black; ventral surface of F1–F5 and fore tibia brownish. **Pubescence.** Similar to that of the male but with sparse hairs on lower paraocular area and depression for antennal scape, as in remaining areas of face. **Punctuation.** Face with finer and sparser (1–2x PD) punctures than in the male.

**Variations.** In all paratypes the maxillary palpus and tegula are brownish.

**Holotype.** ♂, COLOMBIA. *Boyacá*: Arcabuco Prov., Santuario de Fauna y Flora Iguaque, Chaina, 2600 m, 15 March 2001, V. Gonzalez [ICN]. **Paratypes.** 1♂ 3♀, one female with the same data as the holotype, remainder paratypes collected on June 20, 2001 [ICN, SEMC].

**Etymology.** The specific name refers to the most influential deities of the Muisca, a pre-Colombian culture that had a well developed political system in South America. The Muisca lived in the highlands of modern-day departments of Boyacá and Cundinamarca (Colombia). Bochica brought law and morals, and taught the Muisca agriculture and manual arts.

***Chilicola (Oroediscelis) deborahae* Gonzalez sp. nov.**

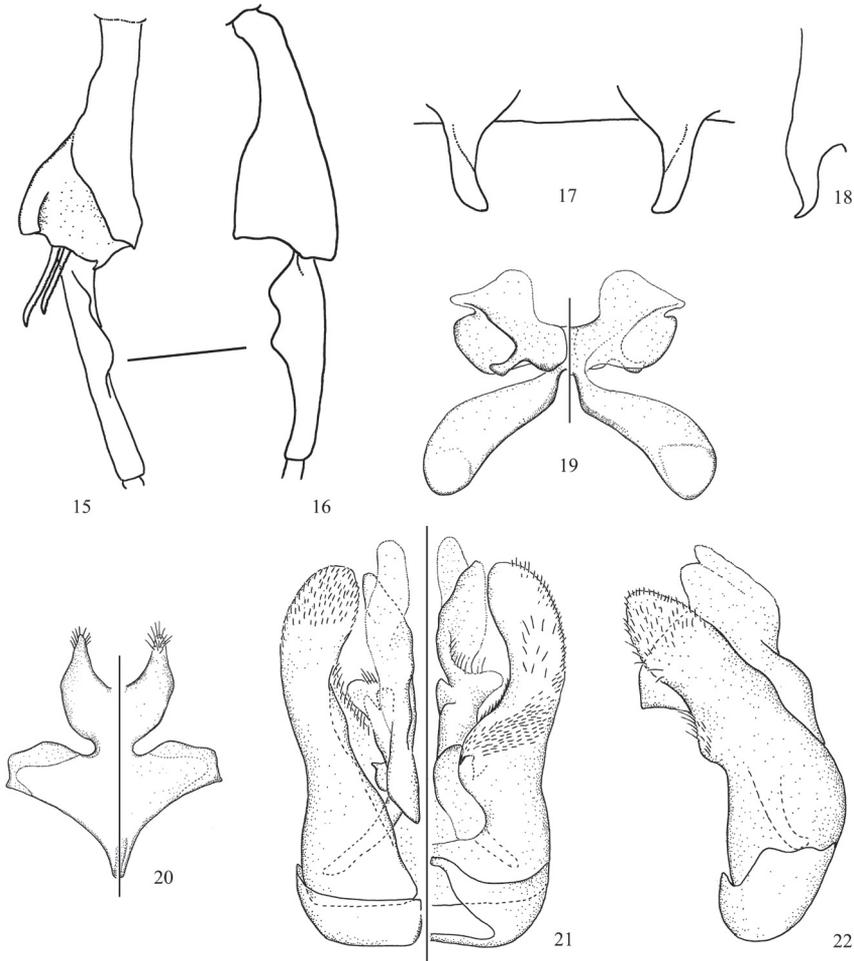
Figs. 15–22

**Diagnosis.** In the key to the species of *Oroediscelis* (Michener 2002), the male of this species runs to *C. bigibbosa* Michener whereas the female runs to *C. benoistiana* Michener and *C. quitensis* Benoist. The male of *C. deborahae* sp. nov. differs from *C. bigibbosa* in the unmodified front and middle femora, longer preapical processes of S4 (Figs. 17, 18), and smaller ventral lobes of the hind basitarsus (Fig. 16). The preapical processes of S4 are flattened apically and pointed in lateral view as in *C. espeleticola* Michener; however, it can be separated by the punctuation of the face, vertex and gena, and the shape of the hind tibia, hind basitarsus, S7 and S8 (compare with Fig. 22 of Michener 2002). The strongly striate vertex and gena of the female of *C. deborahae* sp. nov. distinguishes this species from *C. benoistiana* and *C. quitensis*.

**Description. Male.** Body length 7.2 mm; forewing length 4.8 mm. **Structure.** Head about as long as broad; malar space about two-thirds as long as broad; interalveolar distance 1.5x OD, about as long as alveolorbital distance; interocellar distance 2.1x OD, 0.8x ocellocular distance; ocelloccipital distance about 1.4x OD; scape 3.3x longer than broad; pedicel about as long as broad; F1 2.0x longer than pedicel, 2.1x longer than broad, about as long as F2 and F3 individually; F2 and F3 about as long as wide, F4–F10 progressively shorter. Hind tibia and basitarsus as in Figs. 15, 16. S2 and S3 with low preapical sublateral tubercles; S4 with preapical processes flattened at tips, pointed in lateral view (Figs. 17, 18); S7, S8, and genital capsule as in Figs. 19–22. T7 with distal margin weakly emarginated medially. **Coloration.** Mainly black except for: mandible yellow basally, reddish brown apically; inferior paraocular area yellow; antennal flagellum with dorsal surface light

brown; tarsi dark brown; tegula, wing veins and stigma dark brown; wing membrane faintly smoky. **Pubescence.** Whitish to light yellow; head (including scape) and mesosoma sparsely covered with long, branched hairs ( $\geq 3.0\times$  OD); hind tibia with preapical brush. Distal margin of T1–T6 with weak lateral fasciae; sides of S6 with long tuft ( $\geq 2.5\times$  OD) of yellow hairs, preapical fringe of yellowish hairs surpassed by yellow, transparent lamella. **Punctuation.** Clypeus and

lower supraclypeal area with sparse punctures (1–2x PD), integument shinier on distal half of clypeus, finely imbricate to lineolate between punctures; supraclypeal area duller than clypeus, with few punctures laterally; frons with coarse punctures, as close as they can be; upper paraocular area, vertex, and gena strongly striate; hypostomal area shiny, finely imbricate. Mesosoma finely imbricate with finer and closer punctures on scutum and scutellum; basal half of metepisternum



**Figures 15–22.** Male of *Chilicola deborahae* sp. nov. 15, 16 = hind tibia and hind basitarsus in ventral and profile views. In Fig. 16, tibial spurs are hidden behind basitarsus. The line between figures points to the same projection of basitarsus in both views; 17, 18 = preapical processes of S4 in ventral and profile views; 19–22 = S7, S8, and genital capsule. In divided figures, dorsal view is shown on left, ventral on right.

and sides of propodeum weakly striate; basal area of propodeum with several irregular longitudinal striae. Terga finely lineolate with minute and sparse (1–2x PD) punctures on discs, distal margins smooth and shiny.

**Female.** As described for the male, except for: Body length 6.2–7.1 mm; forewing length 5.1 mm. **Structure.** Head 1.1x longer than broad; interalveolar distance about 2.0x OD, subequal to alveolorbital distance; interocellar distance 2.5x OD, slightly shorter than ocellocular distance; ocelloccipital distance about 1.8x OD; scape about 4.0x longer than broad; pedicel slightly longer than broad; F1 longer than pedicel, 1.8x longer than broad; F2 and F3 each about as long as broad, shorter than F1. **Coloration.** Mandible and lower paraocular area black. **Pubescence.** Metasoma with longer and denser hairs than in male. **Punctuation.** Clypeus dull, with fine and sparse punctures; striae on upper paraocular area, vertex and gena weaker than in the male.

**Holotype.** ♂, COLOMBIA. *Cundinamarca*: PNN Chingaza, La Siberia 4°31'N, 73°45'W, 3170 m, Malaise, 14–31-V-2001. E. Raigoso Leg [ICN]. **Paratypes.** 1♂, 5♀; one female with the same data as the holotype except for: Valle del Fraylejon, Malaise, 02–17-VIII-2000. A. Pérez; remaining paratypes with the following data: **Boyacá:** Arcabuco Prov., Santuario de Fauna y Flora de Iguaque, camino a la laguna, 5°70'N, 73°46'W, 3400–3600 m, Aug 23 2004, in stems of *Espeletia argenteata*. V. H. Gonzalez [ICN, SEMC].

**Etymology.** This species honors Dr. Deborah Smith (University of Kansas), friend and mentor who has provided much inspiration and support to the first author.

**Phylogenetic relationships.** A total of 10 most parsimonious trees (L = 44, CI = 65, RI = 74) were obtained when including *C. deborahae* sp. nov. in the analysis of the data

set of Michener (2002); five nodes collapsed in the consensus tree. The resulting topology was similar to that of Fig. 35 in Michener (2002), except for: *C. maculipes* Michener and *C. bigibbosa* were sister to the remaining species of *Oroediscelis*; *C. deborahae* sp. nov., *C. cuzcoensis* Michener, and *C. espeleticola* were in a polytomy with the clade that includes *C. quitensis* and remaining species. However, we obtained three most parsimonious trees (L = 45, CI = 66, RI = 75) when we included an additional character to Michener's data set (Apex of preapical process of S4: 0 = not apically flattened and pointed in lateral view; 1 = apically flattened and pointed in lateral view, Figs. 17, 18). Only two nodes collapsed in the consensus tree and *C. deborahae* sp. nov. and *C. espeleticola* were sister species. According to this analysis, the preapical processes of S4 with flattened and pointed apex is a synapomorphy that supports such relationship.

**Biological notes.** As described for other Andean species of *Chilicola*, such as *C. espeleticola* and *C. paramo* (Michener 2002, Gonzalez & Michener 2004), nests of *C. deborahae* sp. nov. were also found inside dead, dry, broken, pithy flowering stems of living plants of *Espeletia argentea* Humb. & Bonpl. (Asteraceae). The two stems containing nests had diameters that ranged from 8.0 to 18 mm. Nests consisted of unbranched tunnels through the axes of the stems. The tunnel diameter was 5 mm and length (measured from nest entrance to the upper end of the cell) was 13 mm. Each nest had five cell partitions; one nest had three cells with bee pupae and two with larva and pollen; the other nest had two cells with larva and pollen, two cells with bee pupae, and one long cell (about twice the size of a bee cell) with a pupa of the ichneumonid wasp genus *Grotea* Cresson (Hymenoptera: Ichneumonidae, Labeninae). Cells with bee contents were cylindrical and 8 mm in length.

We found pollen grains from 17 plant species and types (nine genera in 12 families) within cells of *C. deborahae* sp. nov. (Table 1). In both nests, about 80 % of the pollen belonged to *Arcytophyllum* aff. *nitidum* and *A. muticum* (Rubiaceae).

**Table 1.** Spectrum of plant species and pollen types recorded in pollen samples from two nests of *Chilicola deborahae* sp. nov. collected during the transition from the rainy to the dry season in Iguaque on August 23, 2003. N = four cells sampled; \* = pollen grains comprised < 1% of total grains in a cell for all samples.

Family	Species
Asteraceae	<i>Hypochoeris</i> sp. Type 1 Type 2 *Type 3
Fabaceae	<i>Lupinus</i> sp.
Gentianaceae	* <i>Gentiana</i> sp.
Geraniaceae	<i>Geranium</i> sp.
Iridaceae	*Type 1
Melastomataceae	* <i>Miconia</i> sp.
Myrtaceae	<i>Myrcianthes</i> sp.
Poaceae	*Type 1
Rosaceae	* <i>Polylepis/Acaena</i>
Rubiaceae	<i>Arcytophyllum</i> aff. <i>nitidum</i> <i>Arcytophyllum muticum</i>
Solanaceae	*Type 1
Valerianaceae	* <i>Valeriana</i> sp.
Unknown	*Type 1

## DISCUSSION

Given the high abundance of pollen of *Arcytophyllum* aff. *nitidum* and *A. muticum* in the sample, it seems that *C. deborahae* sp. nov. rely heavily on these plants. However, our sample size was very small and restricted to a single locality. It would be interesting to know the role of *C. deborahae* sp. nov. in the pollination biology of *Arcytophyllum*; as far it is known, the tubular white or red flowers of these montane plants are primarily visited by small (1–5 mm in body length) syrphid flies (García-Robledo & Mora 2007).

The *Grotea* specimen, collected from nests of *C. deborahae* sp. nov., is presumably one of the six undescribed species of the genus that occur in Colombia (Herrera 2006). It represents the first record of *Grotea* parasitizing nests of *Chilicola* subgenus *Oroediscelis*, although it has also been recorded from nests of other stem nesting bees, including *Chilicola* (*Chilicola*) *venticola* Packer, *Ceratina*, *Manuelia* (Apidae), and *Megachile* (Megachilidae) (Gauld 2000, Packer 2004).

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**LITERATURE CITED**

- ERDTMAN, G. 1986. The acetolysis method in a revised description. *Svensk Botanisk Tidskrift Lund* 54: 561–564.
- GARCÍA-ROBLEDO, C. & F. MORA. 2007. Pollination biology and the impact of floral display, pollen donors, and distyly on seed production in *Arcytophyllum lavarum* (Rubiaceae). *Plant Biology* 9(4): 453–461.
- GAULD, I. D. 2000. The Ichneumonidae of Costa Rica, 3. *Memoirs of the American Entomological Institute* 63:1–453.
- GOLOBOFF, P. A. 1993. *NoName (NONA), version 1.5.1*. Program and documentation, Fundación and Instituto Miguel Lillo, Tucumán.
- GONZALEZ, V. H. & M. S. ENGEL. 2004. The Tropical Andean bee fauna (Insecta: Hymenoptera: Apoidea), with examples from Colombia. *Entomologische Abhandlungen* 62 (1): 65–75.
- GONZALEZ, V. H. & C. D. MICHENER. 2004. A new *Chilicola* from Colombian Páramo (Hymenoptera, Colletidae, Xeromelissinae). *Journal of Hymenoptera Research* 13(1): 24–30.
- GONZALEZ, V. H., OSPINA, M., & D. BENNETT. 2005. Abejas altoandinas de Colombia: Guía de campo. Instituto de Investigación de Recursos Biológicos Alexander von Humboldt, Bogotá, D.C.
- HERRERA, A. 2006. Labeninae (Hymenoptera: Ichneumonidae) de Colombia. Estudio taxonómico preliminar. Undergraduate thesis. Universidad de Antioquia, Medellín.
- MICHENER, C. D. 2002. The bee genus *Chilicola* in the tropical Andes, with observations on nesting biology and a phylogenetic analysis of the subgenera (Hymenoptera: Colletidae, Xeromelissinae). *Scientific Papers, Natural History Museum, The University of Kansas* 26: 1–47.
- MICHENER, C. D. 2007. *The Bees of the World*. Johns Hopkins University Press, Baltimore, MD, 2<sup>nd</sup> Edition.
- NIXON, K. C. 1999. *WINCLADA, version 0.9.99tuc.13, beta*. Cornell University, Ithaca, New York.
- PACKER, L. 2004. Taxonomic and behavioural notes on the Patagonian Xeromelissinae with the description of a new species (Hymenoptera: Colletidae). *Journal of the Kansas Entomological Society* 77(4): 805–820.

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