

A new species of Long-eared Brown Bat of the genus *Histiotus* (Chiroptera) and the revalidation of *Histiotus colombiae*

Una nueva especie de murciélago pardo de orejas largas del género *Histiotus* (Chiroptera) y revalidación de *Histiotus colombiae*

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ABSTRACT

The South American bats of the genus *Histiotus* comprise between four and eight species, but their taxonomy has been controversial and the limits between species and their distribution are not well understood. In Colombia, *Histiotus humboldti* and *H. montanus colombiae* have been recorded, but undescribed species has been suggested. We evaluated the species richness and distribution of Colombian *Histiotus* using morphological, molecular, and acoustic traits. Our results evidence three species in Colombia, the two previously recorded taxa and a new species from the Cordillera Central of Colombia and northern Ecuador that we describe here. We also revalidated *H. colombiae* as a full species. *H. humboldti* is widely distributed in the Colombian and Ecuadorean Andes and can be sympatric with the other two species. *H. colombiae* is restricted to the Colombian Cordillera Oriental. Finally, we highlight the potential hidden diversity within *Histiotus* in the Peruvian and Bolivian Andes, the need to resolve the evolutionary relationships of the genus, and its implications to the understanding of the processes that have structured the Andean mammal fauna.

Keywords. Andes, Cytochrome b, echolocation calls, morphometry, Vespertilionidae.

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RESUMEN

Los murciélagos suramericanos del género *Histiotus* comprenden entre cuatro y ocho especies, pero su taxonomía ha sido controversial y los límites entre sus especies y sus áreas de distribución no son bien entendidos. En Colombia, se han registrado a *Histiotus humboldti* y a *H. montanus colombiae*, pero se ha propuesto que hay especies por describir. Evaluamos la riqueza de especies de murciélagos *Histiotus* colombianos incluyendo caracteres morfológicos, moleculares y acústicos. Nuestros resultados evidencian la presencia de tres especies en Colombia, los dos taxones previamente registrados y una especie nueva de la Cordillera Central de Colombia y el norte de Ecuador que describimos aquí. Además, revalidamos *H. colombiae* a nivel de especie. *H. humboldti* presenta la distribución más amplia en los Andes colombianos y ecuatorianos, y puede estar en simpatria con las otras dos especies. *H. colombiae* es sólo conocida de la Cordillera Oriental de Colombia. Finalmente, resaltamos la diversidad potencial no descrita de *Histiotus* en los Andes peruanos y bolivianos, la necesidad de resolver las relaciones evolutivas del género y sus implicaciones en el entendimiento de los procesos que han estructurado la fauna Andina de mamíferos.

Palabras clave. Andes, citocromo b, ecolocalización, morfometría, Vespertilionidae.

INTRODUCTION

The Big-eared Brown Bats of the genus *Histiotus* are endemic to South America, occurring along the Andean chains in a variety of mountainous ecosystems, from Venezuela and Colombia to the most southern continental lands of Argentina and Chile, the coastal Atlantic Forest of eastern Brazil, and the semiarid regions of Argentina and Brazil (Handley and Gardner 2008, Feijó et al. 2015, Moratelli et al. 2019). *Histiotus* are easily differentiated from other South American Vespertilionids by their very long pinnae and large tympanic bullae (Handley and Gardner 2008). Recently, molecular phylogenetic analyses found that *Histiotus* was nested within the clade of the New World *Eptesicus* (Hofer and Van Den Bussche 2003, Roehrs et al. 2010, Amador et al. 2018). However, the relationships among worldwide *Eptesicus* remain unresolved and no taxonomic decisions have been taken. Despite that, based on the unique morphology within the South American vespertilionid bats, *Histiotus* has been recognized as a valid genus in recent taxonomic treatments (Simmons 2005, Handley and Gardner 2008, Burgin et al. 2018, Moratelli et al. 2019).

The alpha taxonomy of *Histiotus* has been also controversial and the species limits unclear, due to the similar interspecific skull morphology, few specimens available

in collections, and scarce available molecular data with limited taxonomic and geographic representativeness (Feijó et al. 2015, Giménez et al. 2019). The number of recognized species has varied between four and eight (Handley and Gardner 2008, Giménez et al. 2019, Moratelli et al. 2019). For example, Simmons (2005) recognized seven species: *H. alienus* Thomas, 1916, *H. humboldti* Handley, 1996, *H. laeophotis* Thomas, 1916, *H. macrotus* (Poeppig, 1835), *H. magellanicus* Philippi, 1866, *H. velatus* (I. Geoffroy, 1824), and *H. montanus* (Philippi and Landbeck, 1861), the latter with two additional subspecies, *H. m. colombiae* Thomas, 1928, and *H. m. inambarus* Anthony, 1920. In contrast, Handley and Gardner (2008) considered *H. alienus*, *H. magellanicus*, *H. laeophotis* as subspecies of *H. montanus*. Recently, Moratelli et al. (2019) recognized eight species adding *H. diaphanopterus* Feijó, Da Rocha and Althoff, 2015 to the seven species recognized by Simmons (2005).

Contributions on the taxonomy and distribution of *Histiotus* are few and mostly dedicated to southern South American forms. For instance, Feijó et al. (2015) described *H. diaphanopterus* from northeastern Brazil, and Giménez et al. (2019) showed molecular evidence of specific level differentiation between *H. montanus* and *H. magellanicus* in Argentina and Chile. In contrast, the northern forms, *H. humboldti* and *H. m. colombiae*, distributed along the Andes of Venezuela, Colombia, and Ecuador (Handley and

Gardner 2008, Moratelli *et al.* 2019), remain poorly studied (Rodríguez-Posada 2010).

In a taxonomic account on *Histiopus* of the Cordillera Central of the Andes of Colombia – one of the three roughly parallel chains of the Andes of the country (namely Cordilleras Occidental, Central, and Oriental), extending northeastward almost to the Caribbean Region – Rodríguez-Posada (2010) raised two important issues on the taxonomy of the Colombian *Histiopus*. First, the author considers that due to the wide distribution of *H. montanus* and the recognition of the validity of three of its historical subspecies as full species, *H. m. colombiae* should be considered as a full species too. Second, the author notices morphological differences between specimens identified as *H. montanus* (assigned to *H. m. colombiae*) from the Central and Oriental cordilleras that suggest possible cryptic diversity within *Histiopus*. To evaluate both issues, we reviewed the alpha taxonomy and distribution of *Histiopus* in Colombia using morphological, morphometric, genetic, and acoustic evidence.

MATERIALS AND METHODS

Morphological analyses

We studied 103 specimens belonging to eight proposed taxa of *Histiopus* according to Simmons (2005) and Moratelli *et al.* (2019): *H. montanus montanus*, *H. m. colombiae*, *H. m. inambarus*, *H. diaphanopterus*, *H. humboldti*, *H. laephotis*, *H. macrotus*, and *H. velatus*. We also included specimens from two independent undescribed taxa, both from Peru. One of them is based on one specimen from Cajamarca housed at the American Museum of Natural History (AMNH 268090) that could not be assigned to any *Histiopus* species known to date. The second has been mentioned in the literature as *Histiopus* sp. by Giménez *et al.* (2019) based on two specimens (AMNH 278521, 278524) from Piura (Supplementary material, Appendix 1). We recorded the sex, the locality, and five external measurements from the specimen's labels (in mm). Thirteen craniodontal and mandibular measurements were taken from specimens with calipers accurate to the nearest 0.01 mm. The eighteen standard measurements, including external, craniodontal and mandibular, are based on Handley (1996), Moratelli *et al.* (2013), and Feijó *et al.* (2015). These measurements are described in appendix 2 of the supplementary material: total length (TL), tail length (TaL), hindfoot length (HFL), ear length (Ear), forearm length (FA), greatest

length of skull (GLS), condylo-incisive length (CI), zygomatic breadth (ZB), postorbital breadth (PO), braincase breadth (BCB), mastoid breadth (MB), braincase height (BCH), length of the maxillary tooth row (MTR), palatal length (PAL), post-palatal length (PPAL), breadth across upper molars (M-M), breadth across the canines (C-C), mandibular tooth row (Dent-L).

To evaluate morphometric variation within *Histiopus* from Colombia, we performed a Principal Component Analysis with the thirteen cranial measurements using the statistical package PAST version 4 (Hammer *et al.* 2001). We used the covariance matrix to preserve the information about the relative scale among variables and all data were transformed $\log_{10}$. Due to the completeness of our dataset, only 60 adult specimens (Appendix 1 in supplementary material) with all the variables representing eight taxa (*H. humboldti*, *H. laephotis*, *H. velatus*, *H. macrotus*, *H. montanus montanus*, and *H. montanus colombiae*) were included in our morphometric analyses. For *H. m. colombiae* we included samples from both the cordilleras Oriental and Central of Colombia and from northern Ecuador. To find discrete characters supporting morphometric groups we also explored the diagnostic external and craniodontal traits available in the literature for all studied specimens.

Molecular methods

We extracted whole genomic DNA from two individuals of *H. montanus colombiae* and four individuals of *H. humboldti* from the Cordillera Oriental of Colombia, using standard phenol-chloroform methods (Sambrook *et al.* 1989) on muscle tissues preserved in 96 % ethanol. Unfortunately, we could not access to tissues of specimens identified as *H. montanus colombiae* from the Cordillera Central. Preservation in formalin of ICN specimens refrain us from successful DNA amplification. Amplification of cytochrome-b (Cyt b) was performed using primers glo7L and glo6H (Hoffmann and Baker 2001). A partial sequence between 900 and 1100 base pairs of all individuals was amplified and sequenced following Porter *et al.* (2007), although we increased the annealing temperature from 45–48 °C to 50.5 °C. We carried out purification and sequencing of both strands with the amplification primers on an ABI 3500 sequencer (Applied Biosystems, Waltham, MA, USA) at the Servicio de Secuenciación y Análisis Molecular SSiGMol at the Universidad Nacional de Colombia, Colombia, Bogotá. We submitted genetic sequences to GenBank under accession numbers presented in Table 1 in the

supplementary material. Sequences were manually checked and aligned using BioEdit 7.2.6 software (Hall 1999). We checked for discontinuities and stop codons in each sequence using the ExPASy translate web tool (<https://web.expasy.org/translate/>).

We retrieved two additional sequences from BoldSystems of *H. m. colombiae* provided by Instituto de Recursos Biológicos Alexander von Humboldt, and 17 complete Cyt b gene (1140 bp) sequences from GenBank representing four species of *Histiotus* (*H. m. montanus*, *H. magellanicus*, *H. macrotus*, and *H. sp.* from Peru), and the outgroups *Eptesicus fuscus* (Palisot de Beauvois, 1796) and *Myotis riparius* Handley, 1960, to be used in phylogenetic analyses and genetic distance comparisons (accession numbers available in Table 1 in supplementary material). The complete data set consisted of 26 sequences. We inferred a phylogenetic tree using Maximum Likelihood analyses in RAxML 1.5 beta software (Stamatakis 2014), and Bayesian Inference analyses using MrBayes v.3.1 software (Ronquist and Huelsenbeck 2003), running 10×10^6 generations with one cold and three incrementally heated Markov chains, random starting trees for each chain, and trees sampled every 100 000 generations. We discarded 20 % of the resulting trees as burn-in, and 85 % of the trees were used for generating a 50 % majority-rule consensus. Statistical support for resulting phylogenies was measured in bootstrap support values (of 500 iterations) and Bayesian posterior probabilities. Genetic distance values were estimated using the p-distance method.

Acoustic characterization

We recorded echolocation calls of *H. montanus colombiae* from the Cordillera Oriental using an ultrasound bat-detector (Anabat Swift, Titley Scientific) with a sampling rate of 500 kHz. The bat detector was placed at a height of two meters above ground level in an open space, close to a roost in an abandoned rural house roof (Cundinamarca: Guachetá, Gachetá Alto: 5°27'39.47"N; 73°39'44.57"W; 2893 m). We collected four voucher specimens to confirm the species identification and deposited them at the Colección de Mamíferos Alberto Cadena, Instituto de Ciencias Naturales - ICN collections (ICN 24845-24848). We analyzed only the recordings with at least five pulses of good signal to noise ratio. Sequences of echolocation calls were displayed simultaneously as spectrograms and oscillograms using the software Raven Pro 1.6.1 (Center for Conservation Bioacoustics 2019). Spectrograms were

made of consecutive fast Fourier transformations with an 85 % overlap and visualized on a Hamming type window. For each pulse, we manually measured the following parameters: 1) duration (time between start and end of a call, measured in milliseconds); 2) peak frequency (frequency in kHz corresponding with maximal intensity in the power spectrum), 3) start frequency, 4) end frequency, and 5) interpulse interval (measured from the beginning of a call to the start of next call). We measured these variables to compare them with the data from *H. montanus montanus* from Chile reported by Ossa *et al.* (2015). All the values of acoustic parameters are given as mean $\pm$ 1 SD, since the data sets were not normally distributed, we used nonparametric statistics (Kolmogorov-Smirnov test). We carried out the tests using the software PAST version 4 (Hammer *et al.* 2001).

Nomenclatural statement

This article has been registered in the Official Registry of Zoological Nomenclature (Zoobank) as 3AEB5C6F-509B-4BB3-AB7E-E5C81A946571. In this way, we comply with the requirements of the International Code of Zoological Nomenclature (Article 8.5.3).

RESULTS

Morphological traits

Based on exclusive combinations of discrete characters, for Colombia, we identified three morphological groups of *Histiotus* including *H. humboldti*, *H. m. colombiae* from the Cordillera Oriental and *H. m. colombiae* from the Cordillera Central and northern Ecuador; the latter was considered as a new species described below (see comparisons below and Table 2 in supplementary material). In addition, there is morphological evidence to differentiate the nominal *H. montanus* from southern South America from the three morphological forms found in Colombia (see description and comparisons below and Table 2 of the supplementary material).

The PCA analysis of the morphometric data including specimens from all South America shows that the first two principal components (PC) accounted for 78.8 % of the total variance (PC1 = 70.9 %, PC2 = 7.9 %). The length of the maxillary toothrow (MTR), the palatal length (PAL), the breadth across upper molars (M-M), and breadth across canines (C-C) were related with PC1 and the postorbital breadth (PO) with PC2.

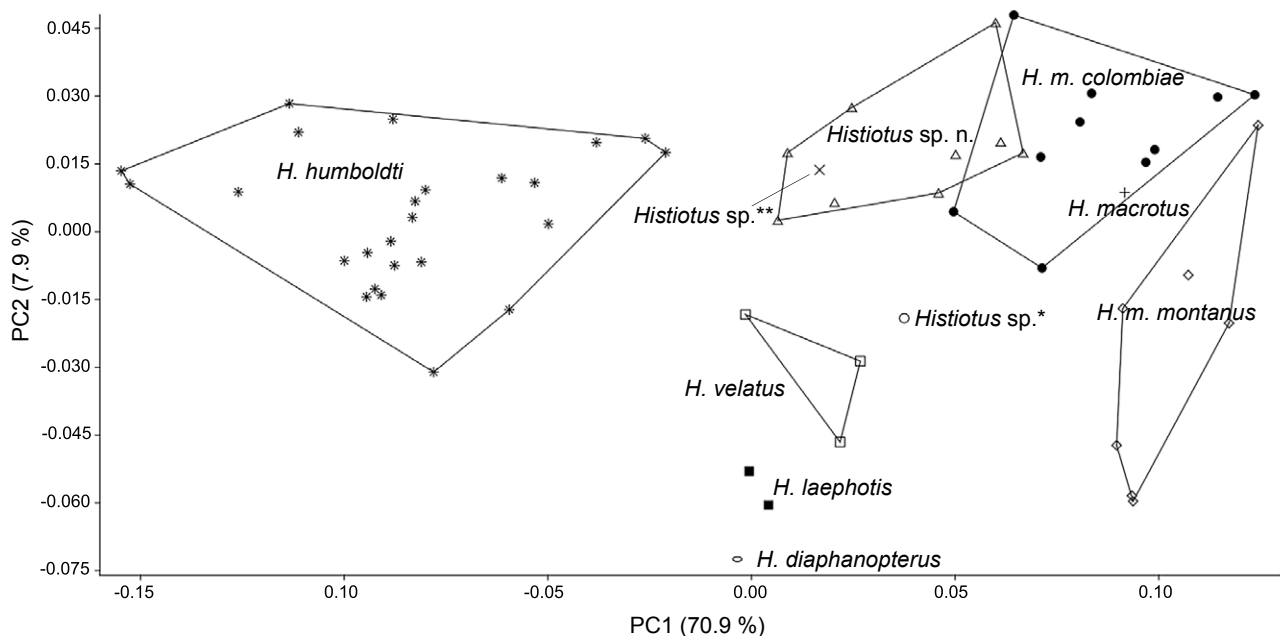


Figure 1. Scatter plot of the first and second principal component scores from the principal component analysis. The analysis was based on 13 cranial and mandibular measurements from seven species of *Histiopus* including *H. humboldti*, *H. laephotis*, *H. velatus*, *H. laephotis*, *Histiopus* sp. n. (described here as *H. cadenai*), *H. colombiae*, *H. macrotus*, and *H. montanus*; and two *H. sp.* which could not be assigned to any *Histiopus* species. *H. sp.**: AMNH 278521 from Peru, Piura. *H. sp.***: AMNH 268090 from Peru, Cajamarca.

The projection of the first two components of the PCA (Fig. 1) showed five morphological groups. *H. humboldti* is observed in the negative extreme of the PC1. *H. m. montanus* from southern South America and *H. m. colombiae* are separated in the morpho-space in two distinct discrete groups in the positive axis of the PC1. Specimens identified as the new species from Colombia and Ecuador appeared also separated from *H. montanus colombiae*, although have a small overlapping. Other morphological distinctive taxa such as *H. velatus* are located at the center of the PC1, and two specimens of *H. laephotis* and one specimen of *H. diaphanopterus* are isolated in the negative extreme of the PC2 (center of the PC1). One specimen (AMNH 268090) identified as *Histiopus* sp. is immersed within the new species of *Histiopus* from Colombia and Ecuador. A second specimen identified as *Histiopus* sp. (AMNH 278521) is isolated between *H. velatus* and the *H. montanus* group. Finally, an individual of *H. macrotus* is immersed in an extreme of the *H. montanus colombiae* group.

Molecular analyses

Sequence alignment of the 23 cytochrome-b sequences of *Histiopus* was unequivocal and without internal stop

codons. Four sequences of *H. m. colombiae* from the Cordillera Oriental of Colombia were recovered as a monophyletic group with strong support (Bootstrap = 100, Posterior probability = 1, Fig. 2) and separated for *H. m. montanus* from Argentina, the latter forming a clade with *H. macrotus*.

The mean genetic distances between *H. m. colombiae* from the Cordillera Oriental and other species of *Histiopus* range between 11.26 % and 12.75 %. *Histiopus m. colombiae* differs about 12.71 % from *H. m. montanus* from southern South America. Genetic distances from other species range from 0.18 % between *H. m. montanus* and *H. macrotus*, and 11.96 % between *H. m. montanus* and *H. magellanicus*. The intraspecific genetic distance value within *H. m. colombiae* from Cordillera Oriental was 1 % between four sequenced specimens (Table 3 in supplementary material).

Acoustic results

We obtained a total of seven wave files from *H. m. colombiae* from individuals flying in the surroundings of their roost. The echolocation calls from *H. m. colombiae* displays the

downward frequency-modulated sweep design. *Histiotus m. colombiae* emitted search calls of a single harmonic. Calls are characterized by durations shorter than 6 ms emitted at intervals of 175.23 ± 54.5 ms (Fig. 3). The comparisons with *H. m. montanus* showed that although both species have a quasi-constant frequency component, the results from Kolmogorov-Smirnov test indicated significant differences for each one of the parameters ($P < 0.005$, Fig. 3). *H. m. colombiae* presented lower start (40.96 ± 4.0 ; N= 100 pulses), end (20.36 ± 0.8 ; N= 100 pulses) and peak frequencies (26.12 ± 1.7 ; N= 100 pulses) than *H. m. montanus* (start frequency (46.26 ± 4.4); end frequency (25.47 ± 1.9) and peak frequency (32.30 ± 1.8) N=90 pulses), and also displayed larger duration pulses (5.01 ± 1.3 ; N = 100 pulses) respect to *H. m. montanus* (3.10 ± 1.1 ; N=90 pulses; see Table 4 in supplementary material).

Taxonomic remarks

We recognize the bats previously identified as *H. montanus colombiae* from the Cordillera Central of Colombia and northern Ecuador as a new species based on a unique combination of morphological traits (Table 2 in supplementary

material) that did not match those of any known species of *Histiotus*. Additionally, we suggest the validity at species level (and discard clinal or subspecific variation) of *H. colombiae* based on a set of morphological traits (Table 2 in supplementary material), the wide genetic distance in the Cyt b gene (around 12.71 %; Table 3 in supplementary material) with the nominal *H. montanus* and the differences in the acoustical parameters between them. We conclude that the nominal *H. montanus* is distributed in southern South America in Argentina, Chile, and Uruguay.

SYSTEMATICS

CLASS MAMMALIA

ORDER CHIROPTERA

Family Vespertilionidae Gray, 1821

Genus *Histiotus* I. Geoffroy Saint-Hilaire, 1824

Histiotus cadenai Rodríguez-Posada, Ramírez-Chaves and Morales-Martínez, new species

Cadena's Long-Eared bat.

Figs. 4-5b, f.

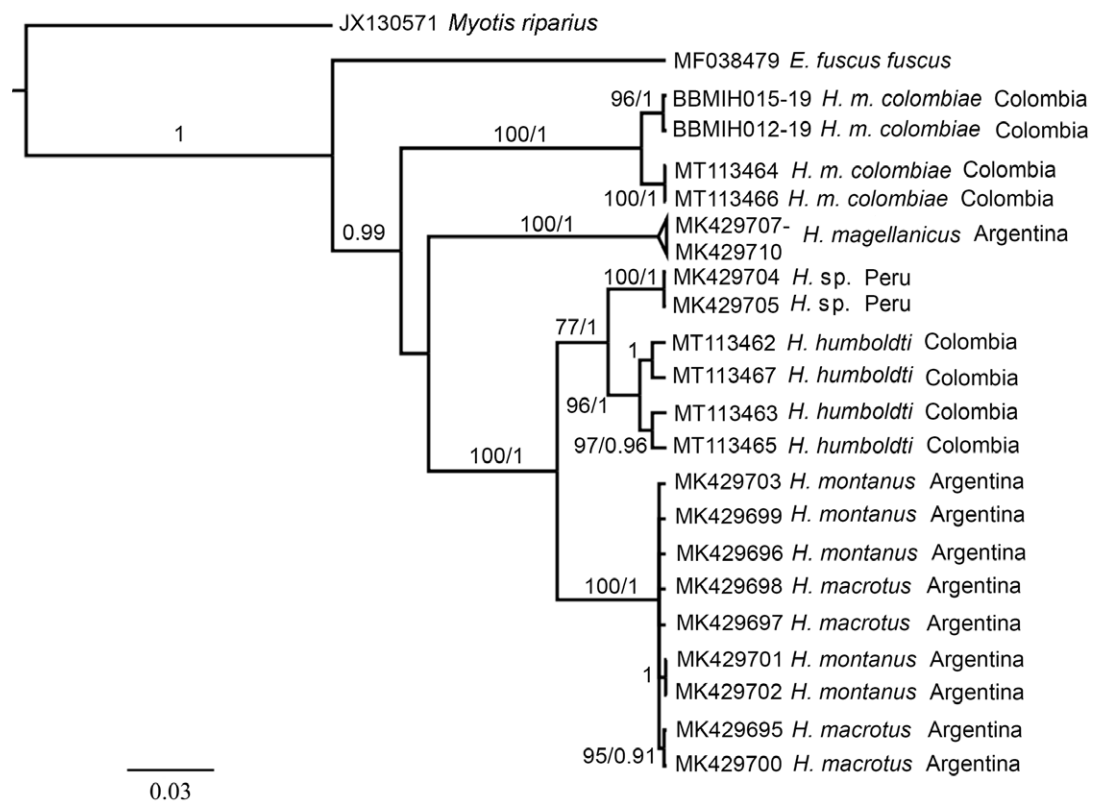


Figure 2. Combined tree from Bayesian and maximum likelihood analysis based on 23 cytochrome-b gene sequences of six species of *Histiotus*. Numbers in parentheses designate individual specimens listed in Appendix 3. Branch support is given for supports above 0.90 in the Bayesian posterior probability and over 75 in bootstrap support values (percentage of 500 iterations) for maximum likelihood.

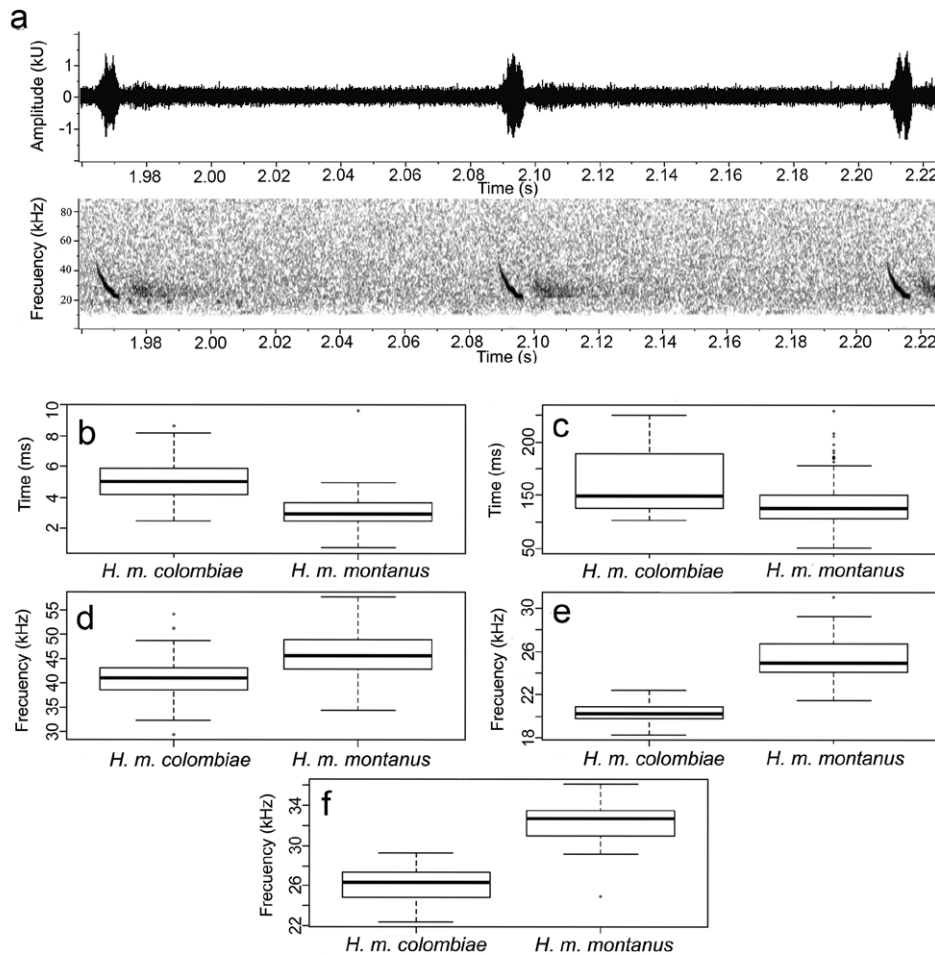


Figure 3. Characteristic of echolocation call of *Histiotus m. colombiae* and differences in call parameters with *H. m. montanus*. a. Oscillogram (above) and spectrogram (below) of the echolocation calls emitted by *H. m. colombiae*. Boxplots for each variable between *H. m. colombiae* from Colombia and *H. m. montanus* from Chile. b. Duration, c. Inter-pulse interval, d. Start frequency, e. End frequency, f. Peak frequency, Data of *H. m. montanus* were provided by Gonzalo Ossa.

Histiotus montanus: Bejarano-Bonilla et al. (2007): 300, Rodríguez-Posada (2010): 176, not *Vespertilio montanus* Philippi and Landbeck, 1861.

Holotype. COLOMBIA. Caldas: ♀ fixed in formalin, preserved in alcohol with the skull removed and cleaned, Manizales, vereda el Desquite, corregimiento 7, Reserva Hidrográfica Río Blanco, finca Martinica, 5° 4'9.40"N, 75°22'38.28"W, 3500 m a.s.l., 6 Feb 2004, col: M. E. Rodríguez-Posada/ field number MRP 171/ICN 16980 (ICN). The external and cranial measurements of the holotype are: TL: 110, TaL: 50, HFL:12, Ear: 36, FA:48, Weight: 11.25, GLS: 17.45, CIL: 16.98, ZB: 10.73, PO: 4.68, BCB: 8.14, MB: 8.98, BCH: 5.99, MTR: 6.57, PPAL: 7.07, PAL: 7.76, M-M: 7.2, C-C: 4.88, Dent-L: 12.17.

Paratypes. COLOMBIA. Caldas: ♀ fixed in formalin, preserved in alcohol with the skull removed and cleaned,

Manizales, vereda el Desquite, corregimiento 7, Reserva Hidrográfica Río Blanco, finca Martinica, 5° 4'9.40"N, 75°22'38.28"W, 3500 m a.s.l., 6 Feb 2004, col: M. E. Rodríguez-Posada /field number MRP 172/ICN 16981 (ICN). Quindío: ♂ skin and cleaned skull, Génova, predio Juntas, 04°08'N, 75°44'W, 3400 m a.s.l., 3 Dec 2004, col: P. X. Roa-A/ field number 20/ Universidad del Valle - UV 13274 (UV). Risaralda: ♀ skin and cleaned skull, Laguna del Otún, bosque del Oso, 3 km N, 1 km W, 4°46'N, 75°24'W, 3560 m a.s.l., 20 Aug 1980, col: M.S. Alberico/ field number 751/ UV 2524 (UV). Valle del Cauca: ♀ skin and cleaned skull, La Florida, Hacienda El Sinaí, 03°19'N, 76°11'W, 2900 m a.s.l., 21 Mar 1993, col: M.S. Alberico/ field number 2235 / UV 11031 (UV). ECUADOR. Pichincha: Sex unknown and ♂ skins and cleaned skulls, Piedra Blanca, ladera norte del volcán Rumiñahui, 0°34'S, 78°30'W, 4000 m a.s.l., 14 and 24 Oct 1992, col: G. Zapata et al./ No field number/ Pontificia Universidad Católica de Ecuador - QCAZ 690 and

QCAZ 691 (QCAZ). Tungurahua: ♀ skin and cleaned skull, Patate, San Francisco, East of Ambato, 01°18'S, 78°30'W, 2438 m a.s.l., 15 Jan 1924, col: G. H. H. Tate / 2703/ AMNH 67648 (AMNH). Napo: ♀ skin and cleaned skull, El Chaco, Oyacachi, 5 km al este, confluencia del río Oyacachi con el río Chalpi, 00°25'S, 78°00'W, 2550 m a.s.l., 18 Feb 2007, col: P. Jarrín/ 18 Feb 1997/ QCAZ 2303 (QCAZ). The external and cranial measurements of the paratypes are in the [Table 5 in supplementary material](#).

Other specimens. COLOMBIA. Tolima: ♂ preserved in alcohol with the skull removed and cleaned, Ibagué, vereda La Cueva, 4°37'N, 75°19'W., 3550 m a.s.l., 2 Aug 2002, No collector data/ Colección Zoológica Universidad del Tolima - CZUT 259 (CZUT). A ♂ preserved in alcohol, Anzategui, vereda El Palomar, 04°36'S, 75°11'W, 3297 m a.s.l., 9 Jul 2007, No collector data/ CZUT 674 (CZUT).

Distribution: *Histiotus cadenai* sp. n. is currently known to occur in 10 localities ([Appendix 1 in Supplementary material](#)) along the northern Andes in a latitudinal range from 01°18'S to 05°04'N. In Colombia, the species has been recorded in six localities in the Cordillera Central (departments of Caldas, Quindío, Risaralda, Tolima, and Valle del Cauca). In Ecuador, the species has been recorded in four localities of the northern portion of the Ecuadorean Andes (provinces of Napo, Pichincha, and Tungurahua). The known altitudinal distribution encompasses a gradient between 2550 to 4000 m a.s.l., including highland ecosystems such as Paramo, Andean montane forest, and anthropized areas. The distance between the extreme localities is approximately 797 km.

Etymology. The species epithet honors the Colombian pioneering mammalogist Alberto Cadena-García. He trained several generations of researchers currently working around the world on topics such as biology and conservation of mammals, continuing his legacy for new generations of biologists.

Diagnosis. This species is distinguished from all other species of the genus *Histiotus* by the following combination of characters: a medium-sized bat (total length: 102-116 mm; forearm length: 45-50 mm; greatest length of skull: 17.12-18.13 mm), with long and silky dorsal fur light brown, ventral coloration is yellowish-brown. Ears are very long (length: 31-36 mm) and not connected by a band. Ears and patagium dark brown, darker than fur coloration. Pinnae are triangular with rounded points and a notch on

the outer edge near the tip. The tragus is ensiform, with a sword-like shape with parallel edges and an acute tip. Calcar is well developed and keeled without lappet. The skull is robust, has a globular braincase with a continuous slope in lateral view, but the posterior border is rounded. Sagittal crest present but form a triangular plate of bone at the intersection with the nuchal crests. Nuchal crests are well developed. Paraoccipital process well developed wide and blunt. The anterior region of the braincase has the parietals straight and highly convergent, forming an abrupt angle with the frontal borders in a conspicuous waist shape towards the postorbital constriction. The supraorbital region is swollen without marked postorbital ridges. The upper toothrow is straight. The position of I2 is lateral to I1. Upper premolar without an anterior projection. M1 has a well-developed but blunt protocone. The angular process has a laminar shape, and it is longer than the condylar process, projected outward, and outside of the condylar process plane. The anterior edge of the coronoid process is straight and forms a 90° angle with the dentary.

Description. *Histiotus cadenai* is a medium-sized bat (total length: 102-116 mm; forearm length: 45-50 mm; greatest length of skull: 17.12-18.13 mm). The dorsal fur is long, silky, and light brown with a brown base (3/4), and light brown tips (1/4). Ventral coloration is yellowish-brown with a dark brown base (2/3) and creamy yellowish tips (1/3). The ears are very long (31-36 mm) and not connected by a band. Pinnae are deltate (equilateral triangle form) with rounded points and a notch on the outer edge near the tip. The tragus is ensiform with parallel edges and an acute tip, the edge is crenulated, but it appears smooth without magnification. Ears and patagia are dark brown, darker than fur coloration. Wings are attached to inferior extremities on the metatarsal. Calcar is well developed and keeled at 2/3 basal of its length. The tail protrudes beyond the uropatagium (ca. 5 mm).

The skull is robust, has a globular braincase with a continuous slope from the parietals to the rostrum making it look flattened in lateral view, but the posterior border is rounded. Sagittal crest present, from the middle of the sagittal suture to the interparietal, where it forms a triangular plate of bone at the intersection of the nuchal crests. Nuchal crests are well developed along the suture occipitoparietalis and between the supraoccipital and the mastoid exposure of the petrosal. Paraoccipital process well developed, wide and blunt. At the basicranium, the basisphenoid has a flattened

surface, the basioccipital has a wide longitudinal ridge with two deep fosses on each side, the lateral edge of these structures forms a laminar process curved up towards the bullae reaching the posterior basicochlear commissure. Bullae are well developed. Mastoid region bulges outward from the braincase. Zygomatic arches are straight on the squamosal ramus and convergent anteriorly towards the maxillary ramus. Glenoid fossa has a rectangular shape where the width is greater than the length, with a recurved posterior edge, and anterior edge well developed. Postorbital processes of the zygomatic arches are triangular, and their base breadth is greater than their height. In dorsal view, the anterior region of the braincase has straight parietals highly convergent towards the postorbital constriction, forming an abrupt angle with the frontal borders in a conspicuous waist shape. The rostrum is shorter than the braincase ($1/3$ of the total length of the skull). The supraorbital region is swollen without marked postorbital ridges. The external nasal fossa has a “U” shape with a conspicuous emargination of the nasal bones. Premaxillae are robust and continuous regarding the braincase and rostrum profile. Palatine processes of the maxillae of the hard palate are visible from the dorsal view. In the ventral view, the hard palate is deeply convex, and the posterior edge of the hard palatal has a spine forming an “M” shape. The hamular processes are thickened but straight. The dental formula is $I\ 2/3\ C\ 1/1\ P\ 1/2\ M\ 3/3$, with a total of 32 teeth. The upper tooth row is straight. The inner upper incisors have convergent tips, each tooth has two asymmetrical cuspids. The position of I2 is beside or lateral to I1. The height of I2 reaches $2/3$ of I1 height. Each upper incisor has a conspicuous cingulum around the tooth crown. Upper canine well developed, simple without accessory cusps, but with a cingulum rounded the crown base. The upper premolar has a continuous cingulum around the crown base, without an anterior projection. Upper molars dilambodont. The M1 has a well-developed protocone but blunt, the paracone is shorter than the metacone, reaching $2/3$ of the size of the metacone. No styler cusps are present. The M2 has a similar pattern and size as M1. The M3 has an “N” form without metastyle. Mandibular ramus is straight with an upward inflection below the coronoid process. The angular process has a laminar form, it is longer than the condylar process, projected outward, outside of the condylar process plane. The coronoid process is taller than wide with a rounded tip curved forward. The anterior edge of the process is straight and forms a 90° angle with the dentary. Lower incisors are trilobed and crowded one after the other.

Canine is curved inward; it has an evident cingulum around the crown that is obscured by the incisors and the anterior premolar cingulum. The lower canine has an accessory cusp on the postero-lingual extreme of the cingulum, near to the contact zone with the anterior lower premolar. The anterior lower premolar is shorter than the posterior lower premolar. The posterior lower premolar has a well-developed cingulum around the crown that forms a small accessory cusp on the antero-lingual region, and a convex emargination on the labial side near the middle of the tooth. Molars height is similar to that of the posterior premolar, but the cusps of the last molar are narrower. Anterior lower molar overlaps with the posterior cingulum of the posterior lower premolar.

Comparisons. *Histiopus cadenai* can be differentiated from *H. velatus* and *H. diaphanopterus* in having a rounded ear tip (fine and acute in the last two species). Additionally, *H. cadenai* is distinct from *H. diaphanopterus* in having black ears and wing membrane (vs. white translucent ears and wings in *H. diaphanopterus*). Cranially, *H. cadenai* has a highly robust cranium and swollen rostrum, and short hard palate being subequal to the size of M2, whereas in *H. velatus* and *H. diaphanopterus* the skull is narrower in dorsal view because the supraorbital region is poorly developed, and the hard palate is long projecting more than two times the size of M2. *H. cadenai*, *H. magellanicus*, *H. colombiae*, and *H. montanus* lack a connective band between ears that is characteristics of *H. velatus*, *H. diaphanopterus*, *H. alienus*, *H. macrotus*, and *H. laeophotis*. *Histiopus cadenai* is differentiated from *H. magellanicus* by its light brown color (dark brown dorsal and ventral coloration of *H. magellanicus*). In addition, *H. cadenai* has a developed sagittal crest only between the posterior side of interparietal bones and the lateral view of the cranium forms a continuous slope, whereas in *H. magellanicus* the cranium has a highly developed sagittal crest extending from the postorbital constriction to the posterior edge of the interparietal bones, and in lateral view, the rostrum and the braincase form an angle. *Histiopus cadenai* is differentiated from *H. montanus* by triangular pinnae with rounded tips compared with the ovate pinnae with pointed tips with the two sides unequal of *H. montanus*. Cranially, in *H. cadenai* the braincase and the rostrum form a continuous slope; the supraorbital region is swollen without marked postorbital ridges; the paraoccipital process is well developed, wide and blunt; the position of I2 is lateral to I1; the angular process has a laminar form and projected

outward, outside of the condylar process plane; whereas in *H. montanus* the braincase and the rostrum form an obtuse (around 160-170°) angle; the supraorbital region is swollen with marked postorbital ridges; paraoccipital process is very well developed but projecting further back from in dorsal view; the position of I2 is behind to I1; the angular process has a laminar form, and, projected upward in the same plane of the condylar process.

Histiotus cadenai only co-occurs with *H. colombiae* and *H. humboldti*. All three species can be externally differentiated by the shape of the pinnae which is triangular with rounded tips and a notch on the outer edge in *H. cadenai*, triangular with rounded tips and smooth outer edge near to the tip in *H. humboldti*, and ovate with asymmetric tips (two sides unequal), and the posterior edge of the tip of the pinna with a notch in *H. colombiae*. Additionally, *H. cadenai* can be differentiated from *H. humboldti* by the tragus shape that is ensiform (sword shape with parallel edges and acute tip) in *H. cadenai*, whereas the tragus of *H. humboldti* is eccentric with the principal axis in one side at the middle point of the structure. Furthermore, *H. cadenai* has a keeled calcar without lappet (lappet present in *H. humboldti*).

Cranially, *H. cadenai* is easily distinguished from *H. humboldti* because the braincase and the rostrum of *H. cadenai* form a continuous slope in lateral view, whereas in *H. humboldti* the braincase and the rostrum form an angle (Figs. 4 and 5). *H. cadenai* has a developed sagittal crest that forms a triangular plate of bone at the intersection with the nuchal crests (not developed sagittal crest and the braincase is rounded without a bone plate in *H. humboldti*). The paraoccipital processes are well developed in *H. cadenai*, but wide and blunt (tubular in form and recurved downward) in *H. humboldti* (Figs. 4 and 5). Furthermore, *H. cadenai* can be distinguished from *H. colombiae* by the upper premolar without an anterior projection in the former and the upper premolar with a well-developed anterior projection in the latter. The supraorbital region of *H. cadenai* is swollen without marked postorbital ridges whereas *H. colombiae* has marked postorbital ridges. In *H. cadenai* the I2 is beside or lateral to I1, whereas in *H. humboldti* and *H. colombiae* the I2 is behind the I1. Finally, the angular process in *H. cadenai* has a laminar form and is projected outside the condylar process plane, whereas in *H. humboldti* the angular process has a tubular form and is projected upward in the same plane of the condylar process, and in *H. colombiae* has a laminar form, projected upward and outside, but in the same plane of the condylar process (Figs. 4 and 5).

Table 1. Craniodental measurements (mm), including the mean, the standard deviation, the observed range, and the sample size of *H. humboldti*, *H. cadenai*, *H. colombiae*, and *H. montanus*. Measurement acronyms are in the main text.

	<i>Histiotus humboldti</i>	<i>Histiotus cadenai</i> sp. n.	<i>Histiotus colombiae</i>	<i>Histiotus montanus</i>
GLS	16.50±0.39;15.45-17.07 (25)	17.62±0.40;17.12-18.13 (9)	18.16±0.38;17.67-18.72 (10)	17.80±0.16;17.60-18.03 (7)
CIL	15.88±0.44;14.43-16.65 (25)	16.89±0.37;16.39-17.48 (9)	17.34±0.43;16.31-17.96 (10)	17.46±0.13;17.30-17.66 (7)
ZB	9.59±3.60;9.00-10.08 (21)	10.74±5.38;10.03-11.17 (6)	11.20±3.55;10.78-11.52 (9)	11.40±4.32;10.93-11.64 (6)
PO	4.44±0.13;4.19-4.65 (25)	4.68±1.56;4.50-4.95 (8)	4.71±0.15;4.49-4.93 (10)	4.30±0.25;4.00-4.61 (7)
BCB	8.04±0.17;7.76-8.43 (25)	8.29±0.25;8.00-8.83 (9)	8.50±0.14;8.23-8.68 (10)	8.28±0.17;8.00-8.52 (7)
MB	8.69±0.16;8.36-9.03 (25)	9.04±0.17;8.86-9.33 (9)	9.24±0.13;9.10-9.55 (10)	9.35±0.24;9.05-9.69 (7)
BCH	5.78±0.19;5.10-6.10 (25)	6.05±0.15;5.85-6.33 (9)	6.23±0.22;5.86-6.63 (10)	5.93±0.29;5.58-6.32 (7)
MTR	5.62±0.26;5.29-6.58 (25)	6.48±0.24;6.12-6.94 (9)	6.48±0.16;6.25-6.68 (10)	6.61±0.11;6.48-6.76 (7)
PPAL	6.66±1.85;6.40-7.06 (23)	6.64±0.29;6.13-7.07 (9)	7.15±0.20;6.84-7.39 (10)	7.01±0.22;6.75-7.38 (7)
PAL	6.90±1.41;6.29-7.53 (24)	7.53±3.33;7.11-7.76 (7)	8.05±0.30;7.59-8.47 (10)	8.36±0.41;7.85-9.03 (7)
M-M	6.13±0.36;5.41-7.01 (25)	6.95±3.07;6.42-7.22 (7)	7.19±0.14;6.98-7.43 (10)	7.21±0.38;6.81-7.91 (7)
C-C	4.32±0.20;3.99-4.68 (25)	4.91±0.21;4.70-5.26 (9)	4.92±0.15;4.74-5.23 (10)	5.21±0.19;5.02-5.56 (7)
Dent-L	10.96±2.21;10.47-11.49 (24)	11.84±3.97;11.15-12.48 (8)	12.35±0.26;12.07-12.76 (10)	12.83±4.86;12.44-13.33 (6)

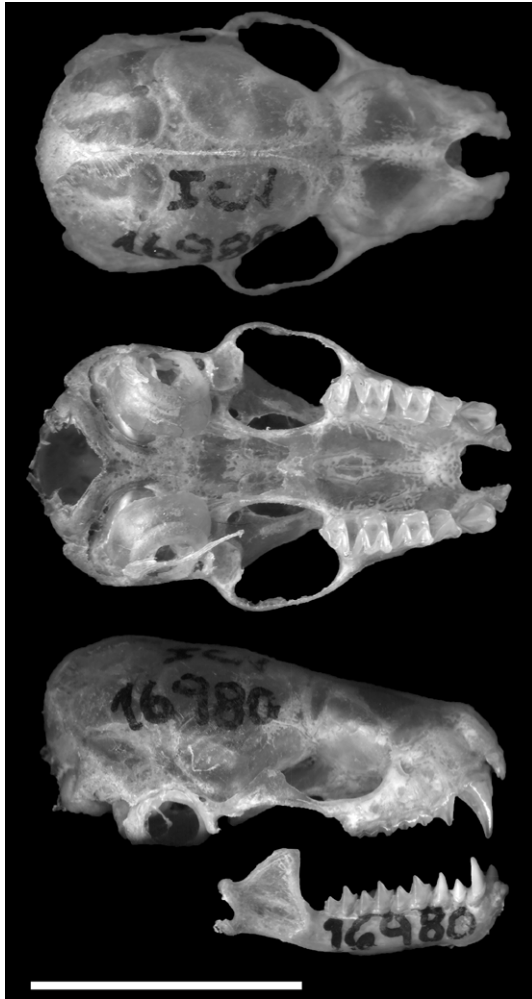


Figure 4. Dorsal and ventral views of skull and mandible of the holotype of *Histiotus cadenai* sp. n. (ICN 16980). Scale bar = 10 mm.

The specific recognition of *Histiotus colombiae*

Our analyses clearly showed the validity of at species level of *H. colombiae* based on acoustic, morphological, and genetic characters and clarified that the nominal *H. montanus* is distributed in Argentina, Chile and Uruguay. For these reasons, we present the information on the systematics of this taxon:

Family Vespertilionidae Gray, 1821

Genus *Histiotus* I. Geoffroy Saint-Hilaire, 1824

Histiotus colombiae Thomas, 1916

Histiotus montanus colombiae: Cabrera, 1958:109; name combination.

Histiotus montanus colombiae: Handley and Gardner, 2008: part.

Histiotus montanus: Rodríguez-Posada 2010: 176, not *Vespertilio montanus* Philippi and Landbeck, 1861.

Type locality: “Choachi, near Bogota,” Cundinamarca, Colombia.

Holotype: British Museum of Natural History (BMNH 99.11.4.1.) ♀ preserved as skin and cleaned skull (skull broken), collected on 20 August 1895.

Description. *H. colombiae* has a dorsal light brown color with the tips lighter than the base. The ventral coloration also presents two phases and is much lighter than the dorsal. The band connecting the ears is not evident; the shape of the pinnae is triangular ovate with asymmetric tips (two sides unequal), and the posterior edge of the tip of the pinna with a notch. The skull is robust (GLS: 17.67-18.72 mm)

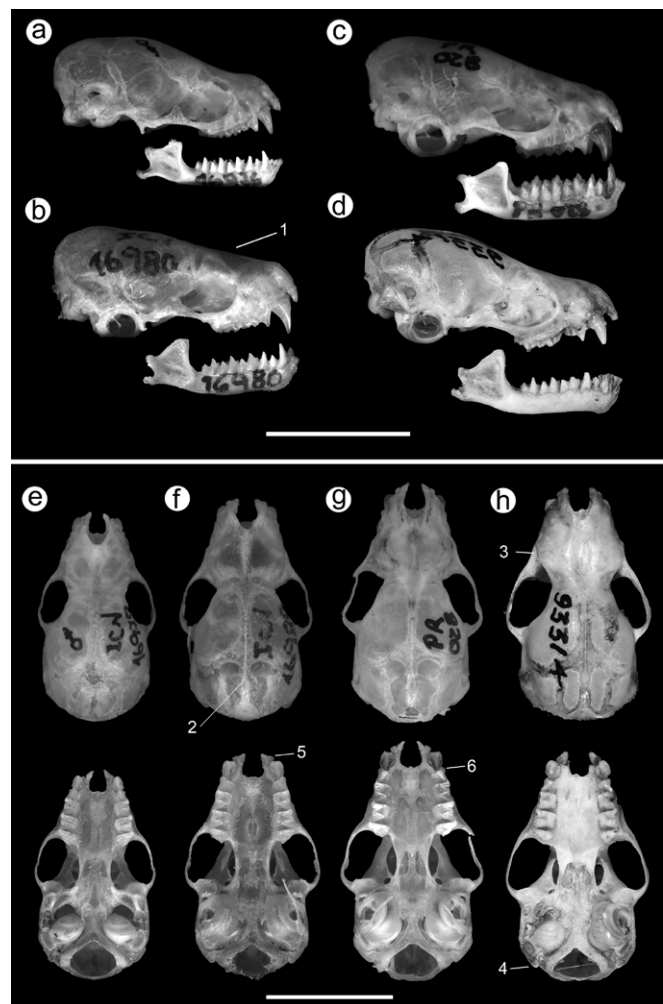


Figure 5. Dorsal and ventral views of skull and lateral views of skull and mandible of **a, e.** *H. humboldti* (ICN 16979). **b, f.** *H. cadenai* sp. n. (ICN 16980). **c, g.** *H. colombiae* (IAVH 8598). **d, h.** *H. montanus* (MHNG 93314). Scale bar 10 mm. 1. The braincase and the rostrum form a continuous slope. 2. Sagittal crest. 3. Supraorbital region. 4. Paraoccipital process. 5. I2 is beside or lateral to I1. 6. Upper premolar with a well-developed anterior projection. ICN: Instituto de Ciencias Naturales; IAVH: Instituto Alexander von Humboldt; MHNG: Muséum d'Histoire Naturelle de la Ville de Genève.

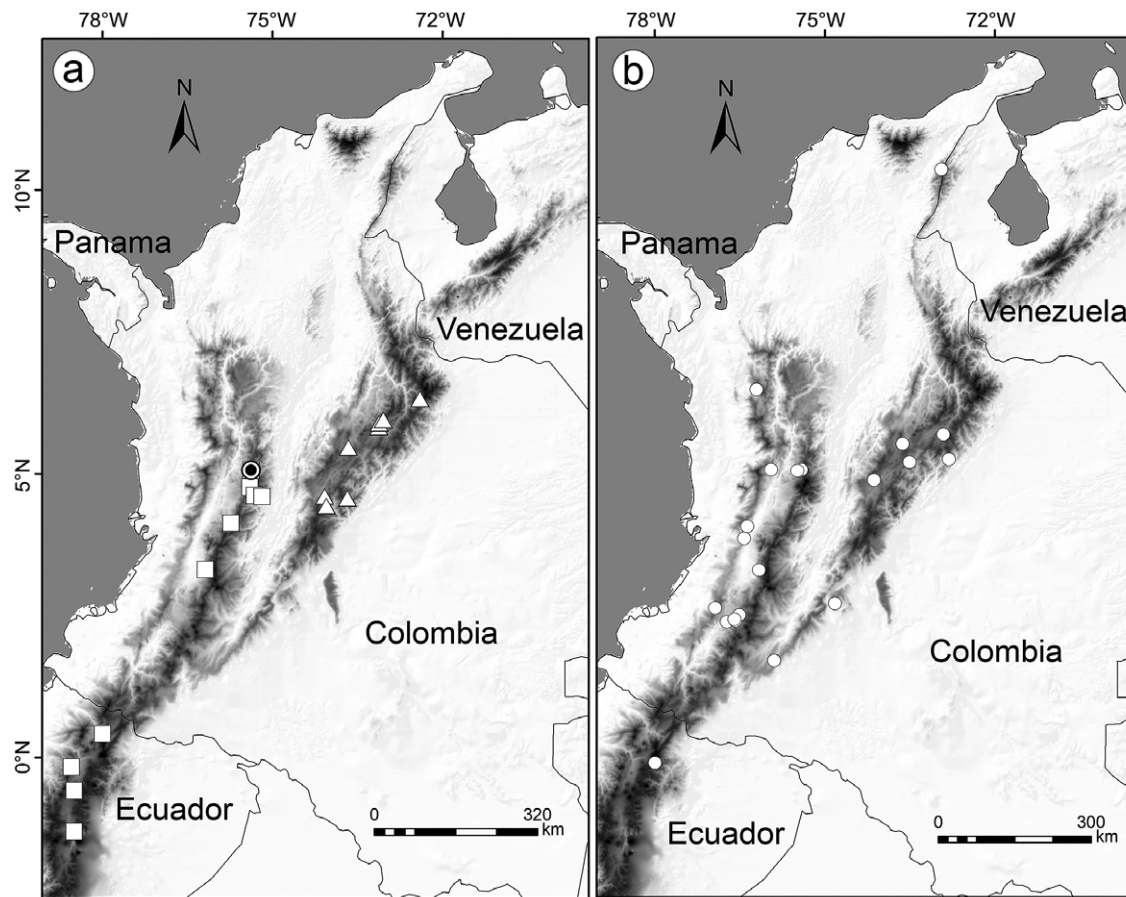


Figure 6. Geographic distribution of three species of *Histiotus* found in Colombia. **a.** *H. cadenai* sp. n. (Squares and black-white circle = type locality) and *H. colombiae* (Triangles). **b.** *H. humboldti* (White circles).

and has a wider interorbital zone than other species such as *H. laephotis* and *H. alienus*. The upper premolar of *H. colombiae* has a well-developed anterior projection (see Thomas 1916). The supraorbital region of *H. colombiae* has marked postorbital ridges. The I2 is behind the I1. The angular process has a laminar form, projected upward and outside, but in the same plane of the condylar process (see Table 2 in the supplementary material for morphological comparison with additional *Histiotus* species).

Distribution of *Histiotus* in Colombia

Histiotus cadenai is distributed from the Cordillera Central in Colombia to the north of Ecuador, over 2550 m (Fig. 6a). The distribution of *H. colombiae* encompasses the high plains of the Cordillera Oriental in the departments of Boyacá and Cundinamarca in an elevational range between 2600 and 3100 m (Fig. 6a). Finally, *H. humboldti* has the widest distribution covering the three Cordilleras of the Colombian Andes and the north of Ecuador in an elevational range between 1700 and 3000 m (Fig. 6b).

DISCUSSION

The new species and the recognition of *H. colombiae* increase the number of recognized species of *Histiotus* from eight (Moratelli *et al.* 2019) to ten, with three of them occurring in Colombia. Despite that, we suggest that number of species in *Histiotus* is larger because the samples from Peru and Bolivia remain unrevised. In ours and in previous assessments of the diversity of *Histiotus* (Gimenez *et al.* 2019) one unidentified specimen from Peru (AMNH 278521) is included as a potential undescribed species. In addition, our review of the holotype of *H. m. inambarus* from Peru showed a particular cranial morphology and might represent a distinct valid species. The validation of these hypotheses should be subject of further studies with the inclusion and comparison of new material.

The inclusion of a taxonomic framework based on different sources of evidence is needed to clarify the diversity of

the genus and will facilitate species delimitation. Unfortunately, taxonomic assessments are challenging because acoustic or genetic data are not available for most *Histiotus* species, and there are few specimens in natural history collections. For instance, the northern South American forms had not been studied since Thomas (1916) and Handley (1996) descriptions, perhaps due to the scarcity of specimens. Although Rodríguez-Posada (2010) called attention to the potential novelties on the diversity of Colombian *Histiotus*, it took a decade to complete morphological, genetic, and acoustic data to compare northern specimens with the southern form (Ossa et al. 2015, Gimenez et al. 2019). The easiest way to solve this situation is the collaborative work, joining resources and capacities of Neotropical mammalogists that work along with the countries where these species are distributed.

Finally, our findings increase the number of bat species of Colombia to 217 species (23 vespertilionids), including recent records for the country (Basantes et al. 2020, Morales-Martínez et al. 2020), and a new species, *Vampyressa voragine*, described recently (Morales-Martínez et al. 2021). We highlight the urgent need for systematic reviews of poorly sampled groups of bats, considering that they have shown an increase in the number of validated taxa based on morphological (Moratelli et al. 2013, Tavares et al. 2014, Mantilla-Meluk and Montenegro 2016), and molecular data (Loureiro et al. 2018). This is particularly important in areas like the northern Andes that are considered a highly threatened biodiversity hotspot (Myers et al. 2000).

AUTHOR'S CONTRIBUTIONS

MRP collected the holotype and one paratype, DMMM processed DNA samples and conducted the Maximum-likelihood analyses, MRP, DMMM, and HERC examined collection specimens, describe the species and did the morphological and morphometric analyses. DMM analyzed the acoustic data. CACA Conducted Bayesian analyses. All authors contributed to the theoretical and conceptual framework, wrote the paper, prepared figures, and tables and reviewed drafts of the paper.

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CONFLICT OF INTEREST

The authors declares that they do not have conflict of interest

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SUPPLEMENTARY MATERIAL

Supplementary material of the article: A new species of Long-eared Brown Bat of the genus *Histiotus* (Chiroptera) and the revalidation of *Histiotus colombiae*

Material suplementario del artículo: Una nueva especie de murciélago pardo de orejas largas del género *Histiotus* (Chiroptera) y revalidación de *Histiotus colombiae*

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Appendix 1. Specimens examined. Acronyms: American Museum of Natural History, New York, USA (AMNH); Natural History Museum, London, UK (BMNH); Colección Zoológica Universidad del Tolima, Ibagué, Colombia (CZUT); Instituto Alexander von Humboldt, Villa de Leyva, Colombia (IAvH); Colección de mamíferos Alberto Cadena García, Instituto de Ciencias Naturales, Universidad Nacional de Colombia, Bogotá, Colombia (ICN); Muséum d'Histoire Naturelle de la Ville de Genève, Geneva, Switzerland (MHNG); Museo de Historia Natural Universidad del Cauca, Popayán, Colombia (MHNUC); Museo de Historia Natural Universidad de Caldas, Manizales, Colombia (MHN-UCa); Museo de La Salle, Bogotá, Colombia (MLS); Museo Javeriano de Historia Natural Lorenzo Uribe, Bogotá, Colombia (MPUJ-MAMM); Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, Lima, Perú (MUSM); Sección Mastozoología, Museo de Zoología, Pontificia Universidad Católica del Ecuador, Quito, Ecuador (QCAZ), and Colección de mamíferos, Universidad del Valle, Cali, Colombia (UV).

***Histiotus cadenai* sp. nov.:** **Colombia:** Caldas, Manizales, vereda el Desquite, corregimiento 7, finca Martinica, Reserva hidrográfica Río Blanco (05°04'N, 075°23'W, 3500 m, ICN 16980♀ holotype, ICN 16981♀ paratype). **Quindío,** Municipio Génova, Predio Juntas (04°08'N, 075°44'W, 3400 m, UV 13274♂ paratype). **Risaralda,** Laguna del Otún, bosque del Oso 3 KM N, 1 Km O (04°08'N, 075°44'W, 3560 m, UV 2524♀ paratype). **Tolima,** Anzoategui, vereda El Palomar (04°36'N, 075°11'W, 3297 m, CZUT 674♂), Ibagué, vereda La Cueva (04°36'N, 075°11'W, 3550 m, CZUT 259♂). **Valle del Cauca,** La Florida, Hacienda El Sinaí (03°19'N, 076°11'W, 2900 m, UV 11031♀). **Ecuador:** **Napo,** El Chaco, Oyacachi, 5

Km al este, confluencia del río Oyacachi con el río Chalpi (00°25'S, 078°00'W, 2550 m, QCAZ 2303♀); **Pichincha,** Piedra Blanca, ladera norte del volcán Rumiñahui (00°34'S, 078°30'W, 4000 m, QCAZ 690 paratype, QCAZ 691♂ paratype); **Tungurahua,** Patate, San Francisco, E of Ambato (01°18'S, 078°30'W, 3000 m, AMNH 67648♀, paratype).

Histiotus colombiae: **Colombia:** **Boyacá,** Duitama, Páramo de la Rusia entre Peñas Blancas y Peñas Negras (ICN 11169♂); La Uvita (06°19'N, 072°24'W, 2700 m, MLS 1652♀, MLS 1686♂); Paipa, vereda Los Medios, Finca Las Quebraditas (05°49'13"N, 073°07'32"W, 2882 m; IAvH 8596♀, IAvH 8598♀); Paipa, vereda Peña Amarilla, Finca El Aguaco (05°53'13"N, 073°07'17"W, 2964 m; IAvH 8597♀). **Cundinamarca,** Bogotá (04°36'N, 074°05'W, 2600 m, AMNH 75235♂, MLS 797♀, 798♀); Bogotá, barrio Usaquén, La Floresta (04°36'N, 074°05'W, 2600 m, MLS 1536♀); Chipaque, Iglesia de Chipaque (04°26'N, 074°02'W, 2400 m, MLS 1536♀); Choachí, near to Bogotá (BMNH 99.11.4.1 ♀, holotype), Guachetá, Vereda Guachetá Alto, Reserva Forestal Protectora El Robledal (05°27'39.47"N, 073°39'44.57"W, 2893 m, ICN 24845♀, ICN 24846♀, ICN 24847♀, ICN 24848♀); Junín, PNN Chingaza, retén la Paila (04°34'N, 073°41'W, 3100 m, MPUJ-MAMM 228, MPUJ-MAMM 230, MPUJ-MAMM 235).

Histiotus diaphanopterus: **Bolivia: Santa Cruz**, 5.5 km NW of Valle Grande (18°25'S, 64°08'W, 1800 m, AMNH 264086).

Histiotus humboldti: **Colombia: Antioquia**, Urrao, vereda Pabón, Inspección de policía Llano Grande, sitio Rioarriba (2550 m, ICN 12879♂); **Boyacá**, Ráquira, Iglesia de Ráquira (05°32'N, 073°38'W, 2150 m, MHNG 1910.074♀), Sogamoso (ICN 21919♂), Úmbita, vereda Tásvita, parte media quebrada La Gacha, Sector El Infierno (05°12'56"N, 073°30'46"W, 3077m, ICN 24841♂). **Caldas**, Manizales (2150 m, MHN-UCa 261♂), Manizales, sitio casa La Y, Reserva Rio Blanco (2350 m, ICN 15160♂), Manizales, vereda Las Palomas Reserva Hidrográfica Rio Blanco, Fundación Ecológica "Gabriel Arango Restrepo" (2400 m, ICN16975♀, ICN16976♀), Manizales, vereda El Desquite, corregimiento 8, finca La Navarra, Reserva hidrográfica Rio Blanco (3000 m, ICN 16979♂). **Caquetá**, Belén de los Andaquíes, Parque Natural Andakí, Divisoria de aguas (01°43'05"N, 075°54'12"W, 1890 m, ICN 23809♂). **Casana-re**, Chámeza, vereda El Jordán (05°15'59"N, 072°49'13"W, 1400 m, ICN 24842♀, ICN 24843♀), **Cauca**, Cerro Munchique, Hacienda "Mirador" Propiedad Cartón Colombia (2400 m, UV 4397♂), El Tambo, corregimiento San Joaquín (MH-NUC uncataloged [F. J. López 01], Popayán (1700 m, MLS 2571♀, MLS 2581♀, MLS 2570♀, MLS 2580♀), Popayán, El Hogar, Bocatoma (1880 m, MHNUC 1395♀), Totoró, Resguardo Novirao (1720 m, MHNUC 1058♀). **Cesar**, Serranía del Perijá, Municipio Manaure, vereda El Cinco (10°22'17"N, 072°57'15.5"W, ICN 2295♂). **Cundinamarca**, Tenjo, vereda Churuguaco alto, predio Buenavista (04°53'45"N, 074°81'22"W, 3180 m, ICN 24844♂). **Risaralda**, Municipio Santuario, vereda El Campamento (2480 m, ICN 11865♂). **Valle del Cauca**, La Florida, Hacienda los Alpes (2450 m, UV 10847♂, UV 10846♂, UV 10516♀, UV 10845♂, UV 10848♂, UV 10849♀, UV 12069♂), Rio Frio, vereda San Ignacio, corregimiento Fenicia, quebrada El Duende (2500 m, UV 12069♂),

Yotoco, Inspección de Policía Puente Tierra; Reserva Natural de Yotoco Alt: 1700 msnm (1700 m, ICN 21778♀, ICN 21779♀). **Ecuador: Napo**, Sierrazul, 10 Kms S, 10 Km W, Río Aragón (00°05'S, 78°00'W, QCAZ 7601♂).

Histiotus laephotis: **Argentina, Jujuy**, Yuto (AMNH 181527♂, AMNH 181528♂).

Histiotus macrotus: **Chile**, Ñuble, Termas de Chillán (AMNH 238025♀). **Perú**, (MUSM 5257).

Histiotus montanus inambarus: **Perú: Madre de Dios**, Rio Inambari (AMNH 37194).

Histiotus montanus montanus: **Chile, Araucanía**, Cautín, Temuco (AMNH 33253♀, AMNH 33255♀), Malleco, Angol (96 m AMNH 93314♀), Ñuble, Termas de Chillan (AMNH 238027♀); **Biobío**, Arauco, Res. Nahuelbuta, Sur de Concepción (37°47'S, 072°59'W, 1200 m, MHNG 1,744,061♂); Lago Risopatron, Km 240 S. Lago Rosselot (MHNG 1744.60♂). **Uruguay: Canelones**, Jaureguyberry (AMNH 188780♀); **Flores**, Ciudad de Trinidad (AMNH 188781♀); **San José**, Chamizo, Estancia Santa Clara (AMNH 183876♀); **Tacuarembó**, Tacuarembó, 40 Km NW (AMNH 205649♀).

Histiotus sp.: **Perú: Piura**, Talara, Quebrada Pariñas, 9.6 km NE of Talara (AMNH 278521♀).

Histiotus sp.: **Perú: Cajamarca**, Las Ashitas, 4lm w of Pachapirana (AMNH 268090).

Histiotus velatus: **Paraguay: Villarica**, Caroveny (AMNH 217565♀). **Perú**: (MUSM 24554, MUSM 39437).

Appendix 2. Descriptions of measurements taken in *Histiotus* specimens.

1. *Total length* (TL): Distance from the tip of the snout to the tip of the last caudal vertebra. Taken from specimen label.
2. *Tail length* (TaL): Measured from the point of dorsal flexure of the tail with the sacrum to the tip of the last caudal vertebra. Taken from specimen label.
3. *Hindfoot length* (HFL): Measured from the anterior edge of the base of the calcar to the tip of the claw of the longest toe. Taken from specimen label.
4. *Ear length* (Ear): Measured from the notch to the fleshy tip of the pinna. Taken from specimen label.
5. *Forearm length* (FA): Taken from the elbow (tip of the olecranon process) to the wrist (including the carpals). This measurement is taken with the wing at least partially folded.
6. *Greatest length of skull* (GLS): Measured from the posteriormost point on the occiput to the anteriormost point on the premaxillae (excluding the incisors).
7. *Condylolincisive length* (CIL): Taken from the posteriormost point on the occipital condyles to the anteriormost point on the upper incisors.
8. *Zygomatic breadth* (ZB): Greatest breadth across the zygomatic arches.
9. *Postorbital breadth* (PO): Least breadth across the frontals, posterior to the postorbital processes or bulges.
10. *Braincase breadth* (BCB): Greatest breadth of the globular part of the braincase
11. *Mastoid breadth* (MB): Greatest cranial breadth across the mastoid region
12. *Braincase height* (BCH): Greatest height of the skull measured between medioventral surface of basioccipital and the highest dorsal point of braincase without include the sagittal crest.
13. *Length of the maxillary tooth row* (MTR): From the anteriormost edge of the canine crown to the posteriormost edge of the crown of M3.
14. *Palatal length* (PAL): Distance between the posterior most point of palatal including the palatal spine and the posterior edge of central incisive alveolus.
15. *Postpalatal length* (PPAL): Distance between the anterior most point of mesopterygoid fossa (without including the palatal spine) and the anterior edge of foramen magnum
16. *Breadth across upper molars* (M-M): Greatest breadth across the outer edges of the crowns of the upper molars.
17. *Breadth across the canines* (C-C): Greatest breadth across the outer edges of the crowns of the upper canines.
18. *Mandibular tooth row* (Dent-L): Greatest distance between anterior most point of dentary and the posterior edge of the condylar process.

Table 1. Genbank and Bold Systems accessions, and voucher numbers used in this study. Museum acronyms in annex 1. LIEB-M Universidad Nacional de la Patagonia San Juan Bosco, Esquel, Chubut, Argentina.

Number	Species	Accession number	Museum voucher	Locality
1	<i>Myotis riparius</i>	JX130571	TTU 102811	Ecuador
2	<i>Eptesicus fuscus fuscus</i>	MF038479	FMNH 214994	USA: Illinois
3	<i>Histiotus colombiae</i>	BBMIH012-19	IAvH 9902	Colombia: Boyacá
4	<i>Histiotus colombiae</i>	BBMIH015-19	IAvH 10013	Colombia: Boyacá
5	<i>Histiotus colombiae</i>	MT113464	ICN 24847	Colombia: Cundinamarca
6	<i>Histiotus colombiae</i>	MT113466	ICN 24846	Colombia: Cundinamarca
7	<i>Histiotus magellanicus</i>	MK429709	LIEB-M 855	Argentina: Chubut
8	<i>Histiotus magellanicus</i>	MK429708	LIEB-M 859	Argentina: Chubut
9	<i>Histiotus magellanicus</i>	MK429710	LIEB-M 860	Argentina: Chubut
10	<i>Histiotus magellanicus</i>	MK429706	LIEB-M 1105	Argentina: Chubut
11	<i>Histiotus magellanicus</i>	MK429707	LIEB-M 1109	Argentina: Chubut
12	<i>Histiotus</i> sp.	MK429704	AMNH 278521	Perú: Piura
13	<i>Histiotus</i> sp.	MK429705	AMNH 278524	Perú: Piura
14	<i>Histiotus humboldti</i>	MT113463	ICN 24841	Colombia: Boyacá
15	<i>Histiotus humboldti</i>	MT113465	ICN 24843	Colombia: Casanare
16	<i>Histiotus humboldti</i>	MT113467	ICN 24844	Colombia: Cundinamarca
17	<i>Histiotus humboldti</i>	MT113462	ICN 23809	Colombia: Caquetá
18	<i>Histiotus macrotus</i>	MK429700	LIEB-M 762	Argentina: Chubut
19	<i>Histiotus macrotus</i>	MK429698	LIEB-M 852	Argentina: Chubut
20	<i>Histiotus macrotus</i>	MK429697	LIEB-M 853	Argentina: Chubut
21	<i>Histiotus macrotus</i>	MK429695	LIEB-M 1100	Argentina: Chubut
22	<i>Histiotus montanus</i>	MK429699	LIEB-M 1113	Argentina: Chubut
23	<i>Histiotus montanus</i>	MK429703	LIEB-M 1115	Argentina: Chubut
24	<i>Histiotus montanus</i>	MK429701	LIEB-M 1116	Argentina: Chubut
25	<i>Histiotus montanus</i>	MK429702	LIEB-M 1117	Argentina: Chubut
26	<i>Histiotus montanus</i>	MK429696	LIEB-M 1118	Argentina: Chubut

Table 2. Comparison among four species of *Histiotus*.

<i>Histiotus humboldti</i>	<i>Histiotus cadenai</i> sp. n.	<i>Histiotus colombiae</i>	<i>Histiotus montanus</i>
Pinnae are triangular with rounded tips and smooth outer edge near to the tip.	Pinnae are triangular with rounded tips and a notch on the outer edge.	Pinnae are ovate. Tips are asymmetric with the two sides unequal. Posterior edge of the pinna with a notch near to the ear tip.	Pinnae are ovate. Tips are asymmetric with the two sides unequal. Posterior edge of the pinna with a notch near to the ear tip.
Tragus eccentric with the principal axe in one side of the middle point of the structure.	Tragus ensiform as a sword form with parallel edges and acute tip.	Tragus ensiform as a sword form with parallel edges and acute tip.	Tragus ensiform as a sword form with parallel edges and acute tip.
Keeled calcar with a lappet.	Keeled calcar without lappet.	Keeled calcar without lappet.	Keeled calcar without lappet.
Braincase and rostrum form an obtuse angle.	Braincase and rostrum form a continuous slope.	Braincase and rostrum form a continuous slope.	Braincase and rostrum form an angle.
No sagittal crest developed, braincase rounded, nuchal crest developed.	Sagittal crest developed a triangular plate of bone at the intersection with the nuchal crests.	Sagittal crest present, a triangular plate of bone at the intersection with the nuchal crests.	Sagittal crest present, a triangular plate of bone at the intersection with the nuchal crests.
Supraorbital region swelled without marked postorbital ridges.	Supraorbital region swollen without marked postorbital ridges.	Supraorbital region swollen with marked postorbital ridges.	Supraorbital region swollen with marked postorbital ridges.
The paraoccipital process is tubular in form and recurved downward.	The paraoccipital process is well developed, wide and blunt.	The paraoccipital process is well developed.	Paraoccipital process very well developed.
Upper premolar without an anterior projection.	Upper premolar without an anterior projection.	Upper premolar with a well-developed anterior projection.	Upper premolar without an anterior projection.
The position of I2 is behind to I1.	The position of I2 is lateral to I1.	The position of I2 is behind to I1.	The position of I2 is behind to I1.
The angular process has a tubular form and projected upward in the same plane of the condylar process.	The angular process has a laminar form and projected outward, outside of the condylar process plane.	The angular process has a laminar form, and projected upward and outside, but in the same plane of the condylar process.	The angular process has a laminar form and projected upward in the same plane of the condylar process.

Table 3. Cytochrome b sequence divergence (%) between and within six *Histiotus* species with p-distance method.

	<i>H. colombiae</i>	<i>H. humboldti</i>	<i>H. macrotus</i>	<i>H. magellanicus</i>	<i>H. montanus</i>	<i>H. sp.</i>
<i>H. colombiae</i>	1.00					
<i>H. humboldti</i>	12.71	1.00				
<i>H. macrotus</i>	12.75	6.46	0.00			
<i>H. magellanicus</i>	11.26	11.39	11.94	0.00		
<i>H. montanus</i>	12.71	6.44	0.18	11.96	0.00	
<i>H. sp.</i>	11.93	3.48	6.67	11.15	6.56	0.00

Table 4. Measurements of different acoustic parameters of *H. montanus montanus* from Chile (Ossa *et al.* 2014) and *H. montanus colombiae* from Cordillera Oriental of Colombia.

	<i>H. m. montanus</i>	<i>H. m. colombiae</i>	Kolmogorov-value	p-Value
N	90	100		
Duration (ms)	3.10 ± 1.1 (0.72-9.65)	5.01 ± 1.3 (2.46-7.55)	0.58842	< 0.005
Interval (ms)	135.80 ± 54.8 (51.66-308.89)	175.23 ± 54.5 (103.02-299.77)	0.31502	< 0.005
Start Frequency (kHz)	46.26 ± 4.4 (37-51.6)	40.96 ± 4.0 (29.31-48.15)	0.53368	< 0.005
End Frequency (kHz)	25.47 ± 1.9 (21.5-31)	20.36 ± 0.8 (18.28-22.41)	0.95895	< 0.005
Peak Frequency (kHz)	32.30 ± 1.8 (24.9-36.1)	26.12 ± 1.7 (22.4-29.24)	0.92789	< 0.005

Data are presented as the mean ± SD, minimum – maximum. N, number of utilized pulses

Table 5. External and craniodental measurements of the type series of *Histiotus cadenai* sp. nov. For acronyms of measurements, see "Appendix 2".

Measurement	*** ICN 16980	** ICN 16981	** UV 13274	** UV 2524	** UV 11031	** QCAZ 2303	** QCAZ 690	** QCAZ 691	** AMNH 67648
Sex	Female	Female	Male	Female	Female	Female	-	Male	Female
TL	110	116	[160]	113	111	-	-	-	-
TaL	50	52	42	47	47	-	-	-	-
HFL	12	10	9	11	9	-	-	-	-
Ear	36	36	[25]	33	31	-	-	-	-
FA	48	47.8	44.98	48.87	46.61	47.64	48	50	-
Weigtht	11.25	11.5	10	12	-	-	-	-	-
GLS	17.45	17.17	17.12	17.86	17.73	17.13	18.13	17.95	18
CIL	16.98	16.83	16.43	17.02	16.57	16.39	17.48	17.27	17.08
ZB	10.73	-	10.46	11.11	10.95	10.03	-	11.17	-
PO	4.68	4.5	4.81	4.53	4.68	4.74	4.69	4.95	4.56
BCB	8.14	8.14	8	8.24	8.17	8.18	8.52	8.83	8.35
MB	8.98	8.9	8.86	9.13	8.88	8.92	9.18	9.33	9.15
BCH	5.99	5.89	6.09	5.97	6.16	6.06	6.33	6.08	5.85
MTR	6.57	6.58	6.4	6.2	6.49	6.36	6.94	6.62	6.12
PPAL	7.07	6.94	6.6	6.86	6.55	6.13	6.36	6.69	6.55
PAL	7.76	7.11	7.69	7.64	7.72	7.44	8.78	8.09	7.35
M-M	7.2	6.76	6.88	7.14	7	7.22	7.58	7.58	6.42
C-C	4.88	4.76	4.75	4.82	4.89	4.85	5.24	5.26	4.7
Dent-L	12.17	11.15	11.7	-	11.98	11.82	12.48	11.33	12.05

*** Holotype, ** Paratypes. [] measurements copied from specimen tag but probably erroneous. - No data.