Biophysical resources

Scientific and technological research article

Characterization of vegetation and the arbuscular mycorrhizal fungi community in soils impacted by gold mining in Colombia

Caracterización de la vegetación y la comunidad de hongos micorrízicos arbusculares en suelos impactados por la minería de oro en Colombia



¹ Servicio Nacional de aprendizaje SENA. Espinal, Colombia.
² Universidad del Tolima. Ibagué, Colombia.
³ Corporación Colombiana de Investigación Agropecuaria [Agrosavia]. Mosquera, Cundinamarca, Colombia.

*Corresponding author: Maryeimy Varón Lopez. Universidad del Tolima. Calle 42 #1B-1, Ibagué, Colombia. mvaronl@ut.edu.co

> Received: December 22, 2020 Approved: August 25, 2022 Published: December 02, 2022

Subject editor: Pedro Rodríguez Hernández, (Corporación Colombiana de Investigación Agropecuaria [AGROSAVIA], Bogotá, Colombia.

How to cite this article: Guzmán Lasso, H. J., Perdomo Useche, Y. L., Mosquera Mosquera, H. R., Pérez-Moncada, U. A., & Varón-Lopez, M. (2022). Characterization of vegetation and the arbuscular mycorrhizal fungi community in soils impacted by gold mining in Colombia. *Ciencia y Tecnología Agropecuaria*, 23(3), e2394. https://doi.org/10.21930/rcta.vol23_num3 [_art:2394 Abstract: Mining extraction has a substantial impact on soil and vegetation cover. However, little is known about its effect on the flora and microorganisms associated with these environments. Arbuscular Mycorrhizal Fungi (AMF) form associations with plants, improving their development in environments contaminated with heavy metals, among other benefits. Native plants and AMF found in the soil degraded by mining play an essential role in establishing programs dealing with recovering areas influenced by these activities. Therefore, this study aimed to characterize vegetation and AMF abundance and diversity in an area affected by gold mining in Colombia. Vegetation was evaluated in two areas, one adjacent to mining deposits with native vegetation (SI-N) and the other in an intermediate area with mining waste and initial natural plant successions (SI-SV). AMF was assessed in the SI-N and the mining waste area (SI-M). Vegetation was quantified using the RAP method, and AMF was assessed using wet sieving from direct soil samples and trap crops. Vegetation evaluations identified 605 individuals belonging to 29 families, 46 genera, and 54 species, and AMF assessment identified 11 species and five genera. Diversity was established using Hill numbers, finding that SI-N was more diverse for vegetation (Q1: 11.94) and SI-M for AMF (Q1: 3.07). Heavy metals and vegetation decreased the diversity and abundance of AMF in the SI-M. Plant species of the genera *Baccharis, Agnostis, Miconia,* and the AMF *Glonus microcarpum* were the most frequent species with potential for recovery in degraded areas.

Keywords: Diversity index, botanical composition, soil, spore abundance, spore richness.

Resumen: La extracción minera tiene un fuerte impacto sobre el suelo y la cobertura vegetal. Sin embargo, poco se conoce sobre el efecto de esta actividad en la flora y los microrganismos asociados a este tipo de ambientes, los cuales son fundamentales para la cadena trófica de muchos organismos. Los hongos formadores de micorrizas arbusculares (HFMA) forman asociaciones con las plantas y pueden avudar a mejorar su desarrollo en ambientes contaminados con metales pesados, entre otros beneficios. Las plantas nativas y los HFMA de los suelos degradados por la minería tienen un papel importante en el establecimiento de programas de recuperación de áreas influenciadas por estas actividades; por lo tanto, el objetivo de este estudio fue caracterizar la abundancia y diversidad de la vegetación y HFMA en una zona afectada por la minería de oro. La vegetación se evaluó en dos áreas: una adyacente a depósitos mineros y con vegetación nativa (SI-N) y otra intermedia con desechos mineros e inicio de sucesión vegetal natural (SI-SV) y los HFMA en el área SI-N y el área de desechos de minería (SI-M). La vegetación se cuantificó mediante el método de RAP y los HFMA por medio de tamizado en húmedo a partir de suelo directo y cultivos trampa. En las plantas, se identificaron 605 individuos, pertenecientes a 29 familias, 46 géneros y 54 especies vegetales y de HFMA, 13 especies y 11 géneros; la diversidad, se determinó mediante los números de Hills, donde el área SI-N fue mayor en la vegetación (Q1: 11,94) y el área SI-M en los HFMA (Q1: 3.07). Los metales pesados y la vegetación fueron factores que disminuyeron la diversidad y abundancia de HFMA en SI-M. Se destacan las especies vegetales de los géneros Agrostis, Baccharis y Miconia y el HFMA Glomus microcarpum como las especies más frecuentes y con potencial para la recuperación de áreas degradadas.

Palabras clave: abundancia de esporas, composición botánica, índices de diversidad, riqueza de esporas, suelo.



Introduction

Diverse anthropic activities such as deforestation, mining, indiscriminate burning, crops, and livestock, among others, cause alterations in the soil's physical, chemical and biological composition, modifying the landscape and its biodiversity (Casas, 2012). Of these, mining is one of the harshest activities due to the impact on the land surface. It causes deforestation, erosion, and deposits of mining waste, where heavy metals accumulate, producing toxicity in the soil, water sources, and nutrients and biodiversity loss (Gill, 2010).

Soil is one of the environments with the most considerable effect on ecosystem stability, as it is a complex system that supports thousands of organisms and their biological interactions. One of the most important is the relationship between plants and arbuscular mycorrhizal fungi (AMF), which are associated with 72 % of plants (Brundrett & Tedersoo, 2018). In this association, they act as plant establishment promoters that benefit the plant nutritionally and increase its tolerance to pathogens (Ramírez & Rodríguez, 2012). Moreover, they counteract soil compaction thanks to the secretion of glomalin, which acts as a binder of minerals and organic matter, i.e., a feature related to soil stability and structure (Genre et al., 2008; Grümberg et al., 2010).

For this reason, it is necessary to conduct studies that evaluate changes in the microbiota's current state, vegetation cover, and physicochemical characteristics of soils degraded by mining activities. They allow comparing plant and microbiological communities according to their richness for aspects of their ecology, structural components, and responses at the plant-environment level that allow plant and microbiological communities to be compared according to their richness for aspects of their ecology, structural components and responses at the plant-environment level, and to apply these comparisons to the identification of organisms that can be used in programs or plans to restore degraded areas (Marrugo-Negrete et al., 2016).

The evaluation of the plant community and AMF has focused mainly on the study of native forests used as reference areas (Quiroga et al., 2019; Rodríguez-Rodríguez et al., 2021) or the effect of anthropic activities such as agroecosystems agroforestry, and livestock (Hernández-Acosta et al., 2021; Restrepo et al., 2019; Sandoval-Pineda et al., 2020; Segura-Madrigal et al., 2020). Worldwide, studies report the effects of gold mining on biological communities, showing that it effectively decreases the diversity of vegetation and AMF (Takarina et al., 2021; Wei et al., 2015). However, few studies in Colombia have analyzed the responses of communities altered by mining, such as the work by Ramirez and Rangel-Ch (2019) that show changes in plant succession processes after gold, platinum, and silver open-pit mining extraction activities in the Choco rainforest. Medina et al. (2009) reported decreases in the abundance and diversity of AMF in areas altered by alluvial mining in the lower Antioqueño River basin.

Therefore, phytoremediation and mycoremediation arise from previous characterization processes in degraded areas and become sustainable alternatives to reduce this impact (Hakeem et al., 2015). The use of plants and fungi with the capacity to detoxify and stabilize contaminated environments, together with the accumulating effect of pioneer plants found in the areafurther

enhanced with the association with AMF, is a strategy used worldwide (Audet & Charest, 2007; Hildebrandt et al., 2007; Schneider, Labory, et al., 2013).

Due to the importance of these alternatives in the recovery or stabilization of areas degraded by mining activities, this study aimed to evaluate the effect of mining on the physicochemical composition of affected soils and the abundance and diversity of vegetation and AMF in the municipality of Santa Isabel, province of Tolima (Colombia).

Materials and methods

Study area

The study was conducted on areas around gold mining locations in Santa Isabel, Tolima, Colombia, where gold has been extracted since 1990 through underground exploration and waste is deposited in terraces. The gold deposit exploration process involves the removal of natural vegetation and soil. The removed soil is fragmented and extracted with xanthates and dithiophosphates to release metallic elements, causing acid rain in the mine. Then, leaching with sodium cyanide occurs, recovering gold by adding zinc dust.

Three areas were evaluated at 1,945 meters above sea level. The first area comprises mining waste deposit terraces (SI-M) at 04°43' N and 75°04' W. The second is an intermediate area with mining waste and initial natural plant successions (SI-SV) located at 04°43' N and 75°04' W. The third is an adjacent area to the mining waste zone with native vegetation and no mining deposits (SI-N), classified as an Andean humid-montane low forest at 04°43' N and 75°05' W. It has an average temperature of 16 ° C, with an average annual rainfall of 1,900 mm (Segura-Madrigal et al., 2020).

The vegetation associated with the study area was evaluated in the SI-SV and SI-N areas but not in the SI-M area, as it did not have any vegetation. The characterization of physicochemical factors and AMF evaluation were only conducted for the SI-M and SI-N areas.

Plant characterization

Plants were characterized using the rapid area plot (RAP) methodology, proposed by Isa-Jaum (2004), with rectangular transects of 4 x 50 m for SI-N. In SI-SV, it was modified to 4 x 14m due to the low vegetation of the area. The vegetation was assessed by census and measuring the species present, selecting those with a diameter at breast height (dbh) \geq 2.5 cm on the left side and all the plant species on the right side. The plant material collected was first pressed on newspaper and then preserved with 70 % alcohol, registering ephemeral characteristics (Ariza et al., 2009). Samples were determined by comparison with JSTOR images of type specimens (Global Plants, 2017), virtual herbaria in COL (Universidad Nacional de Colombia [UNAL], 2016), KEW (The Herbarium Catalog, Royal Botanical Gardens, 2017) and MO (Tropicos, 2017) and by confrontation with material deposited in the Universidad del Tolima TOLI Herbarium.

Sampling and soil analysis

The soil of each area was collected at four sample points, separated by 10 m in the form of an "x," using a Pressler borer at a depth between 0 - 20 cm. Each point was composed of five subsamples homogenized and collected in plastic bags until analysis (Instituto Geográfico Agustín Codazzí [IGAC], 2014).

Physicochemical parameters

Physicochemical analyses (i.e., pH, organic matter, phosphorous, sulfur, Al+H, exchangeable bases, cation exchange capacity (CEC), minor elements, and electrical conductivity (EC)) of the areas under study (SI-M and SI-N) were carried out in the Agroanalysis Soil Laboratory. The total content of heavy metals (arsenic, cadmium, chromium, cobalt, lead, zinc, copper, vanadium, mercury, nickel, and selenium) was established by x-ray fluorescence in the ALPHA 1 SAS Laboratory. Furthermore, available heavy metals (arsenic, cadmium, chromium, chromium, chromium, mercury, and lead) were measured in AGROSAVIA analytical chemistry laboratory.

AMF spore isolation, quantification, and multiplication

The quantitative analysis of the AMF was made from 10 g of soil in each study area and point. Spores were obtained from 10 g of soil using the wet sieving method and sucrose

gradient centrifugation. Evaluations were conducted from soil obtained directly during the collection and using trap crops. The latter consisted of making four replicates per sampling point from 150 g of soil, homogenizing it in soil and sterile sand 3:1 ratio. Then, it was deposited in pots where 20 seeds of *Brachiaria decumbens* were planted. After one month, only three plants were selected and maintained per pot. Plants were irrigated at field capacity and fertilized once a month with a solution of humic extracts with a low phosphorus content. After three months, plants were subjected to water stress and cutting off their leaves, and subsequently, after fifteen days, soil spores were extracted using the methodology published by Gerdemann and Nicolson (1963) with two centrifugations at 4,000 and 3,000 rpm for 5 and 10 minutes respectively (Sieverding, 1983; Usuga et al., 2008).

Morphologic identification of AMF

Spores were grouped according to their morphology. Each morphotype was fixed in polyvinyl alcohol-lactic acid-glycerol (PVLG) and PVLG + Melzer's reagent, taking photographic records in a compound microscope (Primo Star with a 0.5x adapter and an Axiocam coupled camera Erc 5s with a 5-megapixel Zen program). Finally, a comparison was made with the morphological descriptions reported by Blaszkowsky (2003), Schüßler (2013), Oehl et al. (2011), and International Culture Collection of (Vesicular) Arbuscular Mycorrhizal Fungi (INVAM) (University of Kansas, 2017) to complete their identification.

Diversity indices and statistical analysis

AMF and vegetation diversity and abundance were evaluated using adequate numbers proposed by Hill (1973) and the Shannon-Wiener Diversity Index and Simpson's Dominance Index.

Physicochemical soil properties and AMF abundance were analyzed using the Shapiro-Wilk normality test (data of total spore abundance were log (x + 1) transformed prior to analysis to meet requirements for normality and homogeneity of variance). These results were then used to perform an analysis of variance (ANOVA) using the Tukey or Kruskal Wallis comparison methods, depending on whether the data were parametric or non-parametric. The multivariate analysis of principal components and the canonical correlation test between physicochemical soil variables and the diversity of AMF species were used. Tests were carried out using the statistical program CANOCO 4.5 for Windows and Past.

Results and discussion

Floristic abundance and richness

The study registered 605 individuals belonging to 29 families, 46 genera, and 54 species (Table 1). The most abundant plant families in the study areas were Poaceae with 231 individuals (four species), i.e., 38 % of the total percentage of families, Asteraceae with 95 (eight species), Melastomataceae with 38 (eight species), and Solanaceae with 15 (four species). These comprised 24 % of the total percentage of families found in the study, and the remaining 38 % corresponds to the remaining 24 families found. Moreover, diversity (Shannon) and dominance (Simpson) indices show that the floristic composition of the zones analyzed was homogeneous. Nevertheless, the conserved area showed a higher species diversity than the intermediate area.

	Study areas						
T 7 • 11	SI-M		SI-SV	SI-N		SI-M	SI-N
Variables	AME	Vecetation	Vecetation	AME	Vecetetien	TC	TC
	ANT	vegetation	vegetation	ANT	vegetation	Alv	
Species	5	0	10	6	45	7	10
Individuals	19	0	138	420	467	282	618
Simpson_1-D	0.6039	NE	0.6187	0.3251	0.8127	0.2909	0.57
Shannon_H (H')	1.191	NE	1.34	0.6172	2.48	0.6143	1.09
Diversity in order Q1	3.29	NE	3.81	1.85	11.94	1.84	2.99
Diversity in order Q2	1.65	NE	1.62	3.07	1.23	3.43	0.91

Table 1. Diversity indices and Hill numbers. SI-M (mining deposit areas); SI-SV (area with natural plant succession); SI-N (area without mining intervention); TC (trap crops).

Note. NE: Not evaluated

Source: Elaborated by the authors

The SI-SV area shows a diversity equal to a theoretical community with 3.8 genera. On the other hand, a diversity equal to a community with 11.9 effective genera was found in the conserved

area. We can conclude that the conserved area is 8.12 times more diverse when expressing this equivalence.

Our study recognizes the typical floristic richness of the Colombian Andean region in zones surrounding degraded areas due to gold mining activities. Fifty-four plant species were reported together for the SI-N and the SI-SV areas. The abundance and diversity of the floristic composition highlight the Melastomataceae, Asteraceae, and Solanaceae families as the most abundant in the SI-N zone, agreeing with the species diversity and dominance registered for the premontane Andean range (Alvear et al., 2010). The genus Miconia (Melastomataceae) was the most abundant and diverse in these families. It is, however, congruent since this genus is considered the largest in the number of woody plants with flowers, with around 1,050 species described and concentrated throughout the Andes (Goldenberg et al., 2013; Roa-García & Torres-Gonzáles, 2021).

The most predominant species found in our study was the oak tree *Quercus humboldtii* Bonpl. [Fagaceae] (Lozano & Torres, 1974) because it represents 6 % of the total percentage of species found in the SI-N area with 34 individuals. It can be related to its wide distribution (from 1,000 to 3,600 meters above sea level); however, it is more restricted to the high and humid parts of the Andes (Alvear et al., 2010; Corporación Autónoma Regional del Tolima [CORTOLIMA], 2009; Nieto & Joker, 2000; Segura-Madrigal et al., 2020). It is possible that when observing species with high associative and dominant tendencies as *Q. humboldtii*, the floristic composition of the zones analyzed was homogeneous since this has been previously reported In forests with similar altitude and precipitation conditions (Alvear et al., 2010; CORTOLIMA, 2009; Segura-Madrigal et al., 2020).

Besides characterizing the vegetation typical of the area, the floristic composition of an area with mining sludge and native soil (SI-SV) is recognized, noting a primary vegetation type very common in recovery areas, with the presence of the families Asteraceae and Poaceae (Ramirez & Rangel-Ch., 2019). These have a high capacity to settle in open places with degrees of anthropic intervention, such as mining sites and livestock pasture areas, among others. Thanks to their anemochory dispersion strategy, they travel long distances and colonize uncovered or bare soils (Arias & Barrera, 2007; Ranjeet et al., 2010). Within these families, the predominant genera were *Baccharis* (Asteraceae), *Paspalum*, and *Panicum* (Poaceae).

The use of pioneer plants with detoxification capacity and stabilization of contaminated environments, enhanced with the association of AMF, is a strategy used globally (Audet & Charest, 2007; Hildebrandt et al., 2007; Schneider, Labory, et al., 2013). In this study, *Baccharis* stands out as a potential species in the recovery of degraded areas. It is pioneer and abundant, a condition that favors the appearance of propagules of other native plant species (Sarmiento, 2008), establishes in previous stages, and indicates an advance in the plant succession process because it strengthens the first millimeters of the soil profile, which is critical in erosion control (Acuerdo municipal No. 006, 2012; Rangel et al., 2011; Sarmiento, 2008). Besides, *Baccharis* sp. can grow in soils contaminated with heavy metals, together with its potential in the phytoextraction of lead (Pb) and zinc (Zn) (from 2,120 to 3,060 mg/kg of Pb and from 1,090 to 1,490 mg/kg of Zn) making it an important genus in management plans for degraded areas due to mining activity (Afonso et al., 2021; Bech et al., 2012).

Physicochemical soil properties

The soil in the SI-M and SI-N areas was classified as a sandy loam. The parameters that showed significant differences (p < 0.05) were bulk density, Na, Ca, Mg, Al+H, Zn, Cu, Fe, and Mn. Regarding the heavy metals As, Cd, Cr, Co, Cu, Ni, Pb, Vn, and Zn, the similarity analysis showed that most showed significant differences between the study areas, except for Ni and Cr (Table 2). The principal component analysis (PCA) showed that 69.5 % of the data was explained by PC1 and 24.3 % by PC2, totaling 93.4 % of the total variability from the physicochemical attributes. The variables that mainly contributed to the segregation of components were electrical conductivity (mS/cm), pH, percentage of organic matter, and Fe, S, P, and Mn content. Regarding the PCA performed on heavy metals, PC1 represented 83.9 % of the total variability and only 10.3 % for PC2. Hence, PC1 and PC2 contributed 94.2 % of the variability, whereas all heavy metals contributed directly to the total variance.

The canonical correlation analysis indicated that AMF is related to the SI-N area, and the heavy metals are associated with the SI-M area. Furthermore, the number of spores found had a positive relationship with organic matter and the minor elements Fe, Mn, Cu, and S. The opposite case was found for heavy metals that have a negative correlation with the number of spores, especially zinc, lead, vanadium, and chromium; moreover, the latter also showed high values in the SI-N area (Figure 1).



Figure 1. Canonical correspondence analysis of the physicochemical variables associated with the study areas (SI-N, SI-M) and AMF abundance.

Characteristics	SI-M	SI-N
Texture % sand	66.75 ± 10.28 (a)	78 ± 3.11 (a)
Texture % silt	27.75 ± 9.67 (a)	17.5 ± 1.29 (a)
Texture % clay	5.5 ± 2.31 (a)	4.5 ± 2.0 (a)
Bulk density (g cc ⁻¹)	1.1575 ± 0.10 (a)	0.8975 ± 0.02 (b)
Electric conductivity (mS cm ⁻¹)	424.975 ± 284.72 (a)	181.175 ± 27.55 (a)
рН (Н2О)	6.995 ± 0.70 (a)	5.075 ± 0.12 (b)
Organic matter (%)	1.875 ± 2.0 (a)	3.6175 ± 0.53 (a)
S (mg kg ⁻¹)	43.85 ± 45.07 (a)	14.95 ± 2.65 (a)
P (mg kg ⁻¹)	3.505 ± 2.12 (a)	4.36 ± 5.45 (a)
Na (cmol kg ⁻¹)	0.405 ± 0.04 (a)	0.1725 ± 0.06 (b)
K (cmol kg ⁻¹)	0.2475 ± 0.06 (a)	0.185 ± 0.02 (a)
Ca (cmol kg ⁻¹)	3.8675 ± 0.27 (a)	2.965 ± 0.31 (b)
Mg (cmol kg ⁻¹)	0.97 ± 0.06 (a)	0.7825 ± 0.07 (b)
Al+H (cmol kg ⁻¹)	0 (a)	0.63 ± 0.20 (b)
Effective cation exchange capacity (ECEC)	3.15 ± 0.0 (a)	3.15 ± 0.0 (a)
Cu (mg kg ⁻¹)	0.5775 ± 0.07 (a)	0.225 ± 0.08 (b)
Zn (mg kg ⁻¹)	1.25 ± 0.17 (a)	0.68 ± 0.17 (b)
Fe (mg kg ⁻¹)	99.2425 ± 15.70 (a)	63.33 ± 19.01 (b)
Mn (mg kg ⁻¹)	64.2675 ± 21.94 (a)	24.7375 ± 10.08 (b)
B (mg kg ⁻¹)	0.36 ± 0.07 (a)	0.4725 ± 0.18 (a)
Total arsenic (mg kg ⁻¹)	50 ± 51.15 (a)	6.25 ± 1.50 (b)
Total cadmium (mg kg ⁻¹) g	150 ± 237.88 (a)	7 ± 0.0 (b)
Total chromium (mg kg ⁻¹)	51 ± 31.95 (a)	8 ± 8.45 (a)
Total lead (mg kg ⁻¹)	538 ± 777.22 (a)	9.5 ± 3.32 (b)
Total cobalt (mg kg ⁻¹)	46.5 ± 72.34 (a)	1.5 ± 1.0 (b)
Total zinc (mg kg ⁻¹)	1940.25 ± 2790.39 (a)	89 ± 12.73 (b)
Total copper (mg kg ⁻¹)	61.5 ± 56.65 (a)	5.75 ± 2.50 (b)
Total vanadium (mg kg ⁻¹)	91.75 ± 17.97 (a)	26.5 ± 14.08 (b)
Total nickel (mg kg ⁻¹)	28.25 ± 21.69 (a)	10 ± 2.16 (a)
Available arsenic (mg kg ⁻¹)	0.37	NE
Available cadmium (mg kg ⁻¹)	10.64	NE
Available chromium (mg kg ⁻¹)	1.97	NE
Available lead (mg kg ⁻¹)	66.98	NE
Available mercury (mg kg ⁻¹)	< 0.10	NE

Table 2. Physicochemical characteristics of the area with mining deposits (SI-M) and the area without mining intervention (SI-N)

Note. (a) and (b): Average n = 4. Parameters with the same letters show no significant differences. Data are means \pm the standard deviation with a confidence interval of 95 %. NE: Not evaluated. Total heavy metal values were measured using X-ray fluorescence, and available heavy metal values using inductively coupled plasma emission spectrophotometry. Source: Elaborated by the authors.

Heavy metals and vegetation were influential factors in the diversity and abundance of AMF. An example is an SI-M area, with an abrupt decrease in plant species and high total concentrations of As, Ni, Zn, Cd, V, and Cr. It reduces fungi diversity and abundance due to the inability to develop their obligatory symbiotic association, and decreases spore viability (Schneider, Labory, et al., 2013). Two of the heavy metals with the most negative influence on richness indices in mycorrhizal communities are Cr, which is only related to the genus *Gigaspora* in tree samples found in soils contaminated by this metal (Khan, 2001), and V, which reduces mycorrhizal communities can settle in soils contaminated by this metal, which does not affect symbiosis (Rodríguez-Rodríguez et al., 2021; Vidal, 2012).

The increase of spores in the SI-N area may be related to high organic matter (OM) values since it has a positive correlation with genera such as *Acaulospora*, *Entrophospora*, and *Scutellospora* (Perez & Vertel, 2010; Serralde & Ramírez, 2004; Willis et al., 2016). This relationship can be associated with the positive influence that OM has on CEC, soil structure, fertility, water retention capacity, porosity, diversity, and microbial activity. In turn, these microorganisms counteract soil compaction and increase OM levels, as reflected in the SI-N area, in which low bulk density levels and a high abundance of microorganisms are found (Ramírez et al., 2019; Rodríguez-Rodríguez et al., 2021; Rojas, 2013; Salamanca & Sadeghian, 2005).

The values obtained from the minor elements Fe, Mn, Cu, and S also show a possible effect on the composition of the AMF communities in Andean soils. Influence is reported especially for *Glomus* species and those belonging to Diversisporales order in dry seasons, increasing abundance (Ramírez et al., 2019); in this study, the species *D. eburnea* showed the highest correlation to Fe and Mn.

AMF richness and abundance

The AMF count from direct soil samples showed that the spores in the SI-N area are 22 times more abundant (420/19) than those in the SI-M area. Furthermore, it was 2.1 times (618/282) more abundant when the count was obtained from trap crops. These increased the AMF richness for the SI-M (282 spores and seven species) and the SI-N areas (618 spores and ten species). Thus, from soil samples and trap crops, eight different morphospecies were registered in SI-M and ten in SI-N, corresponding to five families, seven genera, and 11 species (Table 3).

Glomeraceae showed the highest number of morphospecies with four, followed by Diversisporaceae with three. Acaulosporaceae and Archaeosporaceae had one morphospecie. Diversity indices demonstrate that the SI-N area is the richest in AMF species, with ten species and a higher dominance than the SI-M area, which exhibited three species with similar abundance (Table 1). Nonetheless, the PCA performed on the species matrix pointed out that the first axis or component explains 98.5 % of the total variance. Further, the species with the highest dispersion are *Glomus macrocarpum*, *Glomus microcarpum*, and *Diversispora spurca*.

	Soil samples		Trap	crops
	SI-M	SI-N	SI-M	SI-N
Archaeosporales				
Archaeosporaceae				
Archaeospora myriocarpa	0	3	0	3
Diversisporales				
Acaulosporaceae				
Acaulospora cavernata	0	0	1	4
Diversisporaceae				
Diversispora eburnea	2	6	2	14
Diversispora spurca	0	1	0	1
Diversispora sp.	1	0	0	4
Entrophosporaceae				
Entrophospora infrequens	0	0	8	5
Claroideoglomus etunicatum	0	3	0	19
Glomerales				
Glomeraceae				
Funneliformis badium	0	0	1	0
Glomus macrocarpum	4	69	235	238
Glomus microcarpum	11	338	33	322
Glomus tenebrosum	1	0	2	8
Total:	19	420	282	618

Table 3. Diversity and abundance of AMF. SI-M (mining deposit areas); SI-SV (area with natural plant succession); SI-N (area without mining intervention).

Source: Elaborated by the authors

Additionally, the species with the highest frequency of occurrence in soil samples from both areas was *G. microcarpum* (SI-M: 52.4 %, SI-N: 65.9 %). In trap crops, this same species was the most frequent for SI-N (47.4 %); however, *G. macrocarpum* was the one with the highest frequency of occurrence in the SI-M area (76.5 %), and *G. microcarpum* in the SI-N area (47.4 %).

Regarding AMF abundance and diversity, a significant increase was observed in trap crops compared with direct counting of soil samples, and as expected, a higher number of spores and species in the native area (SI-N) compared to the impacted area (SI-M). This evidence shows the mining waste's apparent effect on the AMF spores.

Glomus was the most representative due to its abundance in Andean areas degraded by gold mining and wide distribution and tolerance to adverse conditions, ratifying its adaptability to this environment. This finding agrees with reports of its high prevalence in soils contaminated with arsenic and other heavy metals (Pulungan & Nasution, 2021; Rodríguez-Rodríguez et al., 2021; Schneider, Stürmer, et al., 2013), high drought levels (León, 2015), acid pH (Serralde & Ramírez,

2004), low OM content, among others (Cavalcanti et al., 2014; Rodríguez-Rodríguez et al., 2021; Schneider, Stürmer, et al., 2013; Wei et al., 2015).

Different authors also point out that species of Glomeraceae are generalists due to their high sporulation capacity and their ability to quickly adapt to degraded soils (Cavalcanti et al., 2014; Guo et al., 2020; Pulungan & Nasution, 2021; Ramos et al., 2014), besides being found in several regions of Colombia, including Tolima (Aranguen, 2015; Monroy et al., 2013; Peña et al., 2006). *Glomus tenebrosum* stands out for its presence in trap crops in both areas, finding only one spore directly in the soil. It is possibly due to its sensitivity to adverse mine tailing conditions since this species has been reported mainly in trap crops in mining degraded areas (Selvam & Mahadevan, 2002). In Colombia, Salamanca and Silva (1998) found this species directly in the soil of the municipality of San José del Guaviare but without any degree of alteration.

Entrophospora infrequens and *Acaulospora cavernata* were only found in the trap crops of the two areas evaluated. *Entrophospora infrequens, G. macrocarpum*, and *G. microcarpum* have been observed directly in degraded areas due to limestone mining (Texeira-Rios et al., 2013). In Colombia, it has been found in Llanos Orientales (eastern plains), both in citrus crops and in two varieties of corn (Monroy et al., 2013; Serralde & Ramírez, 2004). *A. cavernata* has been recorded worldwide in *Thuja occidentalis* agricultural and forest parcels (Bárcenas-Ortega et al., 2011; Ramos et al., 2014; Souza, 2015). However, it had not been associated with areas degraded by mining activities.

Our study is a starting point in consolidating management and recovery plans for mining areas. It allows knowing, for the first time, the vegetation and the AMF community associated with this type of area, which will be essential in subsequent phytoremediation projects. Considering the importance of AMF for the balance of host plants, the possibility of selecting species found in an affected area that can be used in management programs to recover degraded zones or similar areas is opened. Natural ecosystems, such as SI-N, are sources of various populations of microorganisms, including AMF. They can improve the quality of restoration by reintroducing key organisms from the microbiome of native plants enhancing plant diversity, accelerating succession, and increasing plant establishment that is frequently lost in recovery (Koziol, 2018).

For this reason, it is vital to know the native microbiota of plants in natural ecosystems or that altered by anthropic activities through diversity studies, which have brought benefits such as the discovery of species remarkably adapted to various environmental conditions (Aguilera et al., 2014; Estrada et al., 2011; Teixeira-Rios et al., 2013).

The preceding is added to the fact that mining in the last 15 years has doubled in Colombia, thus increasing the degraded land (Ospina-Correa et al., 2021). Therefore, mining waste management is required in affected areas. For example, the province of Cesar employs biochar and AMF to recover soil affected by carbon mining, improving soil properties and stimulating growth of seedlings (Quiroz-Mojica et al., 2021).

Conclusions

This study found that the SI-M area is contaminated with heavy metals such as As, Cd, Co, Pb, Zn, Cu, and Vn. Further, the physicochemical parameters mainly altered are pH, Na, Ca, Mg, Al+H, Zn, Fe, and Mn compared to the SI-N area. The floristic composition of the zones analyzed was homogeneous. The highest diversity of species was found in the SI-N area, noting a reduction of the vegetal cover due to mining tailings in the SI-SV area. Bulk density, alkaline pH, and a decrease in OM conditions, together with the concentration of the heavy metals Cr and Vn, considerably affected the abundance of AMF in the SI-M area. Moreover, our study highlights plant species of Baccharis (Asteraceae), *Agrostis* (Poaceae), and *Miconia* (Melastomataceae), together with the AMF *Glomus microcarpum* and *G. macrocarpum* for showing potential in phytoremediation processes.

Acknowledgments

The authors thank the Plant Genetics and Biotechnology (GEBIUT) research group, School of Sciences, Universidad del Tolima, for their collaboration and countless orientation during this study.

Contribution of the authors

Herik Johan Guzmán Lasso: Elaboration of a project for access to economic resources, design of methodologies, registration of information in the field, construction of databases, analysis of information and preparation of the manuscript; Yessica Lorena Perdomo Useche: Elaboration of a project for access to economic resources, design of methodologies, registration of information in the field, construction of databases, analysis of information and preparation of the manuscript; Hilda Rocio Mosquera Mosquera: Taxonomic identification of specimens, design of methodologies, analysis of information and preparation of the manuscript; Urley Adrian Perez Moncada: Taxonomic identification of specimens, analysis of information and preparation of the manuscript; Maryeimy Varón López: Elaboration of a project for access to economic resources, design of methodologies, registration of information in the field, activities supervisión, analysis of information and preparation of the manuscript.

Ethical implications

No ethical implications.

Conflict of interest

All authors declare that they have no conflicts of interest in this study.

Financing

University of Tolima- research office.

References

Acuerdo municipal No. 006. (2012). Por medio del cual el Concejo municipal de Santa Isabel Tolima adopta el Plan de Desarrollo Municipal "Santa Isabel merece lo mejor 2012-2015" y se dictan otras disposiciones.

https://santaisabeltolima.micolombiadigital.gov.co/sites/santaisabeltolima/content/files/000463/23142_decreto-no-053-de-2022-categorizacion-municipio_santa_isabel_tolima-1.pdf

- Afonso, T. F., Demarco, C. F., Pieniz, S., Silveira, M., de Oliveira Camargo, F., & Andreazza, R. (2021). Analysis of *Baccharis dracunculifolia* and *Baccharis trimera* for Phytoremediation of Heavy Metals in Copper Mining Tailings Area in Southern Brazil. *Applied Biochemistry and Biotechnology*, 194, 694-708. <u>https://doi.org/10.1007/s12010-021-03652-3</u>
- Aguilera, P., Cornejo, P., Borie, F., Barea, J., von Baer, E., & Oehl, F. (2014). Diversity of arbuscular mycorrhizal fungi associated with *Triticum aestivum* L. plants growing in an Andosol with high aluminum level. *Agriculture, Ecosystem and Environment Journal*, 186, 178-184. <u>https://doi.org/10.1016/j.agee.2014.01.029</u>
- Alvear, M., Betancur, J., & Franco-Rosselli, P. (2010). Diversidad florística y estructura de remanentes de bosque Andino en la zona de amortiguación del Parque Nacional Natural los nevados, Cordillera central Colombiana. *Caldasia*, 32, 2357-3759. <u>https://revistas.unal.edu.co/index.php/cal/article/view/36193</u>
- Aranguen, R. (2015). Caracterización y obtención de cultivos puros de esporas nativas del género Glomus sp. asociadas a rizósfera de aguacate [Bachelor's thesis, Universidad del Tolima]. http://repository.ut.edu.co/browse?type=author
- Arias, M., & Barrera, J. (2007). Caracterización florística y estructural de la vegetación vascular en áreas con diferente condición de abandono en la Cantera Soratama, localidad de Usaquén, Bogota. Universitas Scientiarum, 12, 25-45. <u>https://www.redalvc.org/articulo.oa?id=49912204</u>
- Ariza, W., Toro, J., & Lores, A. (2009). Análisis florístico y estructural de los bosques premontanos en el municipio de Amalfi (Antioquia, Colombia). *Colombia Forestal*, 12(1), 1-16. <u>http://www.scielo.org.co/pdf/cofo/v12n1/v12n1a07.pdf</u>
- Audet, P., & Charest, C. (2007). Dynamics of arbuscular mycorrhizal symbiosis in heavy metal phytoremediation: meta-analytical and conceptual perspectives. *Environmental Pollution*, 147(3), 609-614. <u>https://doi.org/10.1016/j.envpol.2006.10.006</u>
- Bárcenas-Ortega, A., Varela-Fregoso, L., Stürme, S. L., & Chávez-Bárcenas, A. T. (2011). Catálogo de hongos micorrizógenos arbusculares de huertos de aguacate de Michoacán, Mexico. http://www.avocadosource.com/wac7/Section 02/BarcenasAE2011.pdf

- Bech, J., Duran, P., Roca, N., Poma, W., Sánchez, I., Barceló, J., & Poschenrieder, C. (2012). Shoot accumulation of several trace elements in native plant species from contaminated soils in the Peruvian Andes. *Journal of Geochemical Exploration*, 113, 106-111. <u>https://doi.org/10.1016/j.gexplo.2011.04.007</u>
- Blaszkowsky, J. (2003). Arbuscular mycorrhizal fungi (Glomeromycota), endogone and complexipes species deposited in the Department of Plant Pathology, Interart Studio, Szczecin, Poland. <u>http://www.zor.zut.edu.pl/Glomeromycota/Taxonomy.html</u>
- Brundrett, M. C., & Tedersoo, L. (2018). Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytologist*, *4*, 1-8. <u>https://doi.org/10.1111/nph.14976</u>
- Casas, R. (2012). El suelo de cultivo y las condiciones climáticas. Editorial Paraninfo.
- Cavalcanti, A., Vanusa, M., Catanho, M., Barreto, M., Ribeiro, M., & Costa L. (2014). Caracterização morfológica e molecular de fungos micorrízicos arbusculares 58 isolados de áreas de mineração de gesso, Araripina, PE, Brasil. *Hoehnea*, 41(3), 393-400. <u>https://doi.org/10.1590/S2236-89062014000300006</u>
- Corporación Autónoma Regional del Tolima [CORTOLIMA]. (2019). Caracterización básica de la subzona hidrográfica del río Totare. <u>https://www.cortolima.gov.co/planes-y-programas/gestion-integral-del-recurso-hidrico/3414-pomca-rio-totare</u>
- Estrada, B., Palenzuela, J., Barea, J., Ruiz, J., da Silva, G., & Oehl, F. (2011). *Diversispora clara* (Glomeromycetes) a new species from saline dunes in the Natural Park Cabo de Gata (Spain). *Mycotaxon*, 118, 73-81. <u>https://doi.org/10.5248/118.73</u>
- Genre, A., Chabaud, M., Faccio, A., Barker, D., & Bonfante, P. (2008): Prepenetration apparatus assembly precedes and predicts the colonization patterns of arbuscular mycorrhizal fungi within the root cortex of both *Medicago truncatula* and *Daucus carota*. *The Plant Cell*, 20, 1407-1420. <u>https://doi.org/10.1105/tpc.108.059014</u>
- Gerdemann, J. W., & Nicolson T. H. (1963). Spores of mycorrhizal endogone species extracted from soil by wet sieving and decanting. *British Mycological Society*, *2*, 235-244. https://doi.org/10.1016/S0007-1536(63)80079-0
- Gill, S. S. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology Biochemistry*, *12*, 909-930. https://doi.org/10.1016/j.plaphy.2010.08.016
- Global Plants. (2017). Database of digitized plant specimens. https://plants.jstor.org/
- Goldenberg, R., Almeda, F., Caddah, M., Martins, A., Meirelles, J., Michelangeli, F., & Weiss, M. (2013). Nomenclator botanicus for the neotropical genus *Miconia* (Melastomataceae: Miconieae). *Phytotaxa*, 106, 1-171. <u>https://doi.org/10.11646/phytotaxa.106.1.1</u>
- Grümberg, B., Conforto, C., Rovea, A., Boxler, M., March, G., & Luna, C. (2010). La glomalina y su relación con la productividad del cultivo de maíz. XXII Congreso Argentino de la Ciencia del Suelo AACS, 2; 23-25. https://fertilizar.org.ar/wp-content/uploads/2010/09/23.pdf

Guo, Y., Chen, J., Tsolmon, B., He, A., Guo, J., Yang, J., & Bao, Y. (2020). Effects of subsidence

and transplanted trees on soil arbuscular mycorrhizal fungal diversity in a coal mining area of the Loess Plateau. *Global Ecology and Conservation, 24.* https://doi.org/10.1016/j.gecco.2020.e01308

- Hakeem, K. R., Sabir, M., Öztürk, M., Mermut, A., Jan, A., Ali, A., & Rizwanul, Q. (2015). *Soil Remediation and Plants.* Elsevier Inc. <u>https://doi.org/10.1016/C2013-0-13078-4</u>
- Hernández-Acosta, E., Banuelos, J., & Trejo-Aguilar, D. (2021). Revisión: Distribución y efecto de los hongos micorrízicos en el agroecosistema de café. Revista de Biología Tropical, 69(2), 445-461. <u>https://doi.org/10.15517/rbt.v69i2.42256</u>
- Hildebrandt, U., Regvar, M., & Bothe, H. (2007). Arbuscular mycorrhiza and heavy metal tolerance. *Phytochemistry*, *68*, 139-146. <u>https://doi.org/10.1016/j.phytochem.2006.09.023</u>
- Hill, M. (1973). Diversity and evenness: a unifying notation and its consequences. *Ecology*, 54, 427-432. <u>https://doi.org/10.2307/1934352</u>
- Instituto geográfico Agustín Codazzi [IGAC]. (2014). Metodología levantamiento de suelos. https://www.igac.gov.co/es/contenido/tramites-y-servicios/analisis-de-las-propiedadesfisicas-quimicas-biologicas-o-mineralogicas-de-los-suelos
- Isa-Jaum. (2004). Propuesta metodológica de parcelas normalizadas para los inventarios de vegetación. Equipo de investigación Convenio isajaum. <u>http://www.corantioquia.gov.co/ciadoc/FLORA/AIRNR_CN_6425_2005_Tomo1.pd</u> <u>f</u>
- Khan, A. (2001). Relationships between chromium biomagnification ratio, accumulation factor, and mycorrhizae in plants growing on tannery effluentpolluted. *Soil*, *26*, 417-423. https://doi.org/10.1016/S0160-4120(01)00022-8
- Kottek, M., Grieser, J., Beck, C., Rudolf, B., & Rubel, F. (2006). World map of the Köppen-Geiger climate classification updated. *Meteorologische Zeitschrift*, 15, 259-263. <u>https://doi.org/10.1127/0941-2948/2006/0130</u>
- Koziol, L., Schultz, P., House, G., Bauer, J., Middleton, E., & Bever, J. (2018). The plant microbiome and native plant restoration: The example of native mycorrhizal fungi. *BioScience*, 68, 996-1006. <u>https://doi.org/10.1093/biosci/biy125</u>
- León, D. (2015). Comunidades nativas de hongos formadores de micorrizas arbusculares asociadas a yuca silvestre en la amazonia colombiana en época seca y lluviosa bajo dos tipos de paisaje (denudación y llanura aluvial) [Master's thesis, Pontificia Universidad Javeriana]. https://repository.javeriana.edu.co/handle/10554/17009
- Lozano, G., & Torres, J. (1974). Aspectos generales sobre la distribución, sistemática, fitosociológica y clasificación ecológica de los bosques de roble (*Quercus*) en Colombia. *Ecología Tropical*, 1(2), 46-79.
- Marrugo-Negrete, J., Marrugo-Madrid, S., Pinedo-Hernandez, J., Durango-Hernandez, J., & Diez, S. (2016). Screening of native plant species for phytoremediation potential at a Hgcontaminated mining site. *Science Total Environment*, 542, 809-816.

https://doi.org/10.1016/j.scitotenv.2015.10.117

- Medina, M., Orozco, F., & Márquez, M. (2009). Diversidad de Hongos Micorrizógenos Arbusculares de una crono-secuencia de suelos aluviales degradados por actividad minera en el bajo cauca Antioqueño, Colombia. *Revista Facultad Nacional de Agronomía Medellín,* 62(1), 4749-4759. <u>http://www.redalyc.org/articulo.oa?id=179915377002</u>
- Monroy, H. J., Salamanca, C., Cano, C., Moreno, L., & Orduz, J. (2013). Influencia de las coberturas en cultivos de cítricos sobre los hongos formadores de micorrizas arbusculares en Oxisoles del piedemonte llanero colombiano. *Corpoica Ciencia y Tecnología Agropecuaria*, 14(1), 53-65. <u>https://doi.org/10.21930/rcta.vol14_num1_art:343</u>
- Nieto, V., & Joker, D. (2000). Quercus humbodltii. Danida Forest Seed Centre.
- Oehl, F., Sieverding, E., Palenzuela, J., Ineichen, K., & Alves da Silva, G. (2011). Advances in Glomeromycota taxonomy and classification. *IMA Fungus*, *2*, 191-199. https://doi.org/10.5598/imafungus.2011.02.02.10
- Ospina-Correa, J., Osorio-Cachaya, J., Henao-Arroyave, A., Palacio-Acevedo, D., & Giraldo-Builes, J. (2021). Retos y oportunidades para la industria minera como potencial impulsor del desarrollo en Colombia. *TecnoLógicas*, 24(50), 239-256. <u>https://doi.org/10.22430/22565337.1683</u>
- Peña, C., Cardona, G., Mazorra, A., Arguellez, J. H., & Arcos, A. (2006). *Micorrizas arbusculares de la Amazonia colombiana Catálogo ilustrado*. <u>http://sinchi.org.co/</u>
- Perez, C., & Vertel, M. (2010). Evaluación de la colonización de micorrizas arbusculares en pasto Bothriochloa pertusa. Revista MVZ Cordoba, 15(3), 2165-2174. https://doi.org/10.21897/rmvz.303
- Pulungan, A., & Nasution, Y. (2021). Biodiversity Arbuscular Mycorrhizal Fungi in the former Gold Mine Area in North Sumatra. *Journal of Physics: Conference Series, 1819.* <u>https://doi.org/10.1088/1742-6596/1819/1/012045</u>
- Quiroga, J. A., Roa, H. Y., Melo, O., & Fernández, F. (2019). Estructura de fragmentos de bosque seco tropical en el sur del departamento del Tolima, Colombia. *Boletín científico centro de museo de historia natural, 23(1), 31-51.* <u>https://doi.org/10.17151/bccm.2019.23.1.2</u>
- Quiroz-Mojica, L., Daza-Mendoza, M., Díaz-Muegue, L., Melo-Rios, A., & Peñuela-Mesa, G. (2021). Efecto de biochar, micorrizas arbusculares y *Guazuma ulmifolia*, en la rehabilitación de suelos mineros. *Terra Latinoamericana*, 39. <u>https://doi.org/10.28940/terra.v39i0.709</u>
- Ramirez, G., & Rangel-Ch, O. (2019). Sucesión vegetal en áreas de minería a cielo abierto en el bosque pluvial tropical del departamento del Chocó, Colombia. Revista académica colombiana ciencias exactas fisicas naturales, 43, 673-688. <u>http://dx.doi.org/10.18257/raccefyn.896</u>
- Ramírez, M., & Rodríguez, A. (2012). Mecanismos de defensa y respuestas de las plantas en la interacción micorrícica: una revisión. *Revista Colombiana de Biotecnología*, 14, 271-284. https://revistas.unal.edu.co/index.php/biotecnologia/article/view/32888
- Ramírez, M., Pére, U., Serralde, D., Peñaranda, A., Roveda, G., & Rodriguez A. (2019).

Diversidad de comunidades de hongos formadores de micorrizas arbusculares asociados a los cultivos de uchuva (*Physalis peruviana* L.). *Agronomía Colombiana 37(3)*, 239-254. https://doi.org/10.15446/agron.colomb.v37n2

- Ramos, P., Saggin, O., Barbosa, H., Stürmer, S., Siqueira, J., & Carbone, M. (2014). Fungos micorrízicos arbusculares em campos de murundus após a conversão para sistemas agrícolas no cerrado. *Revista Brasileira de Ciência do Solo, 38*, 1703-1711. https://doi.org/10.1590/S0100-06832014000600005
- Rangel, J., Lowy, P., & Aguilar, M. (2011). Colombia diversidad biótica II. Tipos de vegetación en Colombia. Universidad Nacional de Colombia.
- Ranjeet, K., Venita, J., & Shambhu, P. (2010). Impact of degradation on biodiversity status and management of an alpine meadow within Govind Wildlife Sanctuary and National Park, Uttarkashi, India. *International Journal of Biodiversity Science, Ecosystem Services & Management*, 6, 146-156. <u>https://doi.org/10.1080/21513732.2011.568972</u>
- Restrepo, K., Montoya, M., Henao, P., Gutiérrez, L., & Molina, L. (2019). Caracterización de hongos micorrízicos arbusculares de suelos ganaderos del trópico alto y trópico bajo en Antioquia, Colombia. *Idesia (Arica), 37(1),* 35-44. <u>https://dx.doi.org/10.4067/S0718-34292019005000301</u>
- Rios, T., Gomes, R., Costa, L., Oehl, F., & Pereira, C. (2013). Arbuscular mycorrhizal fungi in a semi-arid, limestone mining-impacted area of Brazil. *Acta Botanica Brasilica*, 27(4), 688-693. <u>https://doi.org/10.1590/S0102-33062013000400006</u>
- Roa-García, C. E., & Torres-González, A. M. (2021). Caracterización florística y estructural como línea de base para la restauración ecológica de bosques en la microcuenca del rio Barbas. Revista de la Academia Colombiana de Ciencias Exactas, físicas y Naturales, 45(174). https://go.gale.com/ps/i.do?id=GALE%7CA659502224&sid=googleScholar&v=2.1&i t=r&linkaccess=abs&issn=03703908&p=IFME&sw=w&userGroupName=anon%7Eb 727cbe4
- Rodríguez-Rodríguez, R. M., Kemmelmeier, K., Fátima Pedroso, D., Araújo, F., Valentim dos Santos, J., Gastauer, M., Fróis, C., Ramos, S., Siqueira, J., & Carbone, M. (2021). Native arbuscular mycorrhizal fungi respond to rehabilitation in iron ore mining areas from the Eastern Brazilian Amazon. *Pedobiologia, 89.* https://doi.org/10.1016/j.pedobi.2021.150768
- Rojas, J. (2013). Comparación de métodos de determinación en ensayo de rotaciones en siembra direct. INTA Centro Regional Chaco Formosa.
- Royal Botanical Gardens. (1853). Kew herbarium catalogue. http://apps.kew.org/herbcat/navigator.do
- Salamanca, A., & Sadeghian S. (2005). La densidad aparente y su relación con otras propiedades en suelos de la zona cafetera colombiana. *Cenicafé*, *56*(4), 381-397. <u>https://www.cenicafe.org/es/publications/arc056%2804%29381-397.pdf</u>
- Salamanca, C. R., & Silva, M. (1998). Las micorrizas como alternativa para el manejo sostenible de los agroecosistemas tropicales. Corpoica.

- Sandoval-Pineda, J. F., Pérez-Moncada, U. A., Rodriguez, A., & Torres-Rojas, E. (2020). Alta presencia de cadmio resulta en baja diversidad de hongos formadores de micorrizas arbusculares asociados a cacao (*Theobroma cacao L.*). Acta Biologica Colombiana, 25(3), 333-344. <u>http://dx.doi.org/10.15446/abc.v25n3.78746</u>
- Sarmiento, Y. (2008). Restauración en explotaciones de minas caliza. Luna Azul, 27, 75-84. http://www.scielo.org.co/pdf/luaz/n27/n27a05.pdf
- Schneider, J., Labory, C., Rangel, W., Alves, E., Guilherme, L., & Guimarães, L. R. (2013). Anatomy and ultrastructure alterations of *Leucaena leucocephala* (Lam.) inoculated with mycorrhizal fungi in response to arsenic-contaminated soil. *Journal of Hazardous Materials* J1, 262, 1245-1258. <u>https://doi.org/10.1016/j.jhazmat.2012.05.091</u>
- Schneider, J., Stürmer, S., Guilherme, L., Moreira, S., & Soares, C. (2013b). Arbuscular mycorrhizal fungi in arsenic-contaminated areas in Brazil. *Journal of Hazardous Materials J1*, 262, 1105-1115. <u>https://doi.org/10.1016/j.jhazmat.2012.09.063</u>
- Schüßler. A. (2013). PHYLOGENY. http://www.amf-phylogeny.com
- Segura-Madrigal, M. A., Andrade, C., & Sierra-Ramírez, E. (2020). Diversidad florística y captura de carbono en robledales y pasturas con árboles en Santa Isabel, Tolima, Colombia. *Revista de Biología Tropical, 68(2),* 383-393. <u>https://doi.org/10.15517/rbt.v68i2.37579</u>
- Selvam, A., & Mahadevan, A. (2002). Distribution of mycorrhizas in an abandoned fly ash pond and mined sites of Neyveli Lignite 64 Corporation, Tamil Nadu, India. Basic and Applied Ecology, 284(40), 277-284. <u>https://doi.org/10.1078/1439-1791-00107</u>
- Serralde, A., & Ramírez, M. (2004). Análisis de poblaciones de micorrizas en maiz (Zea mays) cultivado en suelos ácidos bajo diferentes tratamientos agronómicos. Ciencia y Tecnología Agropecuaria, 5(1), 31-40. https://doi.org/10.21930/rcta.vol5_num1_art:22
- Sieverding, E. (1983). Manual de métodos para la investigación de la micorriza vesículo arbúscular en el laboratorio. Centro Internacional de Agricultura Tropical (CIAT).
- Souza, T. (2015). Handbook of arbuscular mycorrhizal fungi. Springer. <u>https://doi.org/10.1007/978-3-319-24850-9</u>
- Takarina, N., Sinaga, I., & Rifqoh, T. (2021). Riparian plant diversity in relation to artisanal mining sites in Cikidang River, Banten, Indonesia. *Biodiversitas Journal of Biological Diversity*, 22(1). <u>https://doi.org/10.13057/biodiv/d220149</u>
- Teixeira-Rios, T., Gomes de Souza, R., Costa Maia, L., Oehl, F., & Pereira, C. (2013). Arbuscular mycorrhizal fungi in a semi-arid, limestone mining-impacted area of Brazil. Acta Botanica Brasilica, 27, 688-693. <u>https://doi.org/10.1590/S0102-33062013000400006</u>

Tropicos.	(2017).	Missouri	Botanical	Garden.		
http://www.tropicos.org/NameSearch.aspx?projectid=7						
Universidad	Nacional	de	Colombia	[UNAL.		
(2016).	Coled	cciones	de	plantas.		
http://www.biovirtual.unal.edu.co/es/colecciones/search/plants/						

- University of Kansas. (2017). International Culture Collection of (Vesicular) Arbuscular Mycorrhizal Fungi (INVAM). <u>http://invam.wvu.edu/</u>
- Usuga, C., Castañeda, D., & Franco, A. (2008). Multiplicación de hongos micorriza arbuscular (h.m.a.) y efecto de lamicorrización en plantas micropropagadas de banano (*Musa AAA* cv. *Gran Enano*) (Musaceae). *Revista Facultad Nacional de Agron*omía, *61*(1), 4279-4290. <u>http://www.scielo.org.co/pdf/rfnam/v61n1/a05v61n1.pdf</u>
- Vidal, J. (2012). Respuesta de la simbiosis tripartita rhizobium-leguminosa-micorriza arbuscular ante vanadio y níquel [Master's thesis, Institución de enseñanza e investigación en ciencias agrícolas]. https://1library.co/document/lq5wl77q-respuesta-simbiosis-tripartita-rhizobiumleguminosa-micorriza-arbuscular-vanadio.html
- Wei, Y., Chen, Z., Wu, F., Hou, H., Li, J., Shangguan, Y., Zhang, J., Li, F., & Zeng, Q. (2015). Molecular diversity of arbuscular mycorrhizal fungi at a large-scale antimony mining area in southern China. *Journal Environment Science*, 29, 18-26. <u>https://doi.org/10.1016/j.jes.2014.10.002</u>
- Willis, A., Sparks, T., Felinov, B., & Harris, P. (2016). Organic amendment increases arbuscular mycorrhizal fungal diversity in primary coastal dunes. *European Journal of Ecology*, 2, 1-8. <u>https://doi.org/10.1515/eje-2016-0011</u>