## Artículo de Revisión

## Reactive species of oxygen, oxidative stress and its relationship with tissular destruction in periodontitis

Especies reactivas de oxígeno, estrés oxidativo y su relación con la destrucción tisular en

## periodontitis

Espécies reativas de oxigênio, estresse oxidativo e sua relação com destruição tecidual na periodontite

Lorena Meñaca-Guerrero<sup>1 ™ ORCID</sup>, Amileth Suarez-Causado<sup>2</sup>™ ORCID, Antonio José Díaz-Caballero<sup>3</sup>™ ORCID

1. Biochemistry Master's student, GITOUC research group, University of Cartagena, School of Medicine, Cartagena- Bolívar, Colombia. 2. PhD in biochemistry and molecular biology, director of Prometeus & Biomedicina Aplicada a las Ciencias Clínicas research group, University of Cartagena, School of Medicine, Cartagena- Bolívar, Colombia.

3. PhD in biomedical sciences, director of GITOUC research group, University of Cartagena, School of Dentistry, Cartagena- Bolívar, Colombia.

**Fecha correspondencia:** Recibido: noviembre de 2019. Aceptado: junio de 2020.

#### Forma de citar:

Meñaca-Guerrero L, Suarez-Causado A, Díaz-Caballero AJ. Reactive species of oxygen, oxidative stress and its relationship with tissular destruction in periodontitis. Rev. CES Odont 2020; 33(2): 112-127.

Open access © Derecho de autor Licencia creative commons Ética de publicaciones Revisión por pares Gestión por Open Journal System DOI: http://dx.doi.org/10.21615/ cesodon.33.2.10 ISSN 0120-971X e-ISSN 2215-9185

## Abstract

During periodontitis inflammatory mediators and reactive oxygen species (ROS) are released, when they increase they produce oxidative stress. This review article describes the role played by ROS and oxidative stress in the development and evolution of inflammation and tissue injury during periodontitis. For this, a review of the literature was carried out in databases such as PubMed, ScienceDirect, Wiley Online Library, Springer, Plos one, Nature, Sage journals, Hindawi and Taylor & Francis Online, showing the following results: ROS produce direct damage and indirect to periodontal tissues. Direct damages include lipid peroxidation, protein and DNA oxidation. Indirect damage involves the regulation of signaling pathways of the nuclear transcription factor kappa B (NF- $\kappa$ B), the c-Jun N-terminal kinase pathway (JNK), the pathways of inflammasome and autophagy causing tissue destruction and creation of a pro-inflammatory state in periodontitis.

Keywords: Reactive oxygen species; Oxidative stress; Periodontitis

## Resumen

Durante la periodontitis se liberan mediadores inflamatorios y especies reactivas de oxígeno (ROS), cuando se incrementan producen estrés oxidativo. Este artículo de revisión describe el papel que desempeñan las ROS y el estrés oxidativo en el desarrollo y evolución de la inflamación y lesión tisular durante la periodontitis. Para ello, se realizó una revisión de la literatura en bases de datos como PubMed, ScienceDirect, Wiley Online Library, Springer, Plos one, Nature, Sage journals, Hindawi y Taylor & Francis Online, mostrando los siguientes resultados: las ROS producen daño directo e indirecto a los tejidos periodontales. Los daños directos incluyen peroxidación de lípidos, oxidación de proteínas y del ADN. Los daños

indirectos involucran la regulación de las vías de señalización del factor de transcripción nuclear kappa B (NF-κB), la vía de la quinasa c-Jun N-terminal (JNK), las vías del inflamasoma y autofagia provocando la destrucción tisular y creación de un estado proinflamatorio en la periodontitis.

Palabras clave: Especies reactivas de oxígeno; Estrés oxidativo; Periodontitis.

#### Resumo

Durante a periodontite, são liberados mediadores inflamatórios e espécies reativas de oxigênio (EROs), no momento em que eles incrementam produzem estresse oxidativo. Este artigo de revisão descreve o papel que desempenham as EROS e o estresse oxidativo no desenvolvimento e na evolução da inflamação e lesão tecidual durante a periodontite. Por isso, uma revisão da literatura foi realizada em bancos de dados como PubMed, ScienceDirect, Wiley Online Library, Springer, Plos one, Nature, Sage, Hindawi e Taylor & Francis Online, mostrando os seguintes resultados: as EROS produzem dano direto e indireto para os tecidos periodontais. O dano direto inclui peroxidação lipídica, oxidação de proteínas e DNA. O dano indireto involucra a regulação das vias de sinalização do fator de transcrição nuclear kappa B (NF-κB), da via c-Jun N-terminal kinase (JNK), das vias inflamassoma e autofagia, causando destruição tecidual e criação de um estado pró-inflamatório na periodontite.

Palavras-chave: Espécies reativas de oxigênio; Estresse oxidativo; Periodontite.

## Introduction

The periodontium is a specialized connective tissue that is constituted by the gum, the periodontal ligament, the cement and the alveolar bone. Its main function is to provide support and support to the tooth in its alveolus (1) The teeth are in contact with the junctional epithelium, a non-keratinized stratified squamous epithelium that allows the migration of polymorphonuclear neutrophils (PMN) as a defense mechanism (2).

The periodontal tissue is affected by periodontal diseases and conditions that includes various pathologies including periodontitis whose classification and definition were updated by the American Academy of Periodontics (AAP) and the European Federation of Periodontics (EFP) in 2017 (3). Periodontitis is a chronic inflammatory disease of high prevalence, affecting 11.2% of adults in the world, which represents the sixth most common human disease (4). In Colombia, the majority of the population (61.8%) between 18 and 79 years old evidences periodontitis in its different degrees of severity (5).

Periodontitis has been linked to obesity, type 2 diabetes mellitus (6), type 1 diabetes mellitus (7), cardiovascular disease and coronary heart disease (8).

There is evidence of a positive association between periodontitis and oxidative stress (9). During periodontitis, ROS are released from PMN in response to invading microorganisms, these ROS are responsible for oxidative stress and much of the tissue damage during infection (10). This association between oxidative stress and pathophysiology and tissue destruction in periodontitis has been subject to observational, experimental and systematic reviews, but these are not enough and the results are inconclusive (11) and the signaling pathways are not well established, so more research in this area is necessary to a comprehensive understanding of

periodontitis (12). A better understanding of this association can give us a more established view of the pathogenesis of periodontitis, and therefore improve therapeutic strategies.

The objective of this review is to describe the role played by ROS and oxidative stress in the development and evolution of inflammation and tissue injury during periodontitis. In this review, the following terms are defined and explained: periodontitis, free radicals, ROS, oxidative stress, antioxidant systems and the mechanism of action of reactive oxygen species and oxidative stress in tissue destruction in periodontitis, including signaling pathways which regulates ROS to produce tissue injury in the periodontium

## **Materials and methods**

#### Type of study: Review article

**Topic:** ROS, oxidative stress and its relationship with the pathophysiology of periodontitis.

**Inclusion criteria:** research published during published from January 1, 2007 to December 31, 2018 full text, clinical trials, cross-sectional studies, longitudinal studies, observational studies, cross-sectional studies, cases and controls, review articles, systematic reviews, epidemiological reports, articles related to the current classification of periodontal diseases and conditions, articles on the global burden of periodontal diseases, book chapters and *in vitro* and *in vivo* studies. An idiomatic restriction was applied, including only works published in English and in international journals, except the national epidemiological report (ENSAB IV).

**Exclusion criteria:** letters to the director, newspapers, conferences, news, comments, case reports, symposia, compendiums, pilot studies, expert opinions and illustrated essays.

#### **Search strategies**

Initially a preliminary search was conducted to estimate the amount of information published on the subject under study, identify the terms that will be used in the search and the most appropriate databases. A systematic search was then carried out in the following databases: PubMed, ScienceDirect, Wiley Online Library, Springer, Plos one, Nature, Sage journals, Hindawi and Taylor & Francis Online. Descriptors of such as "ROS", "oxidative stress", "antioxidant", "chronic periodontitis" and "periodontal disease" "periodontitis" were used and related to each other and with free terms. This search was executed during June 1, 2017 to December 31, 2018.

The authors of this document made critical readings of the articles for their selection, extraction and data analysis. The selected articles were classified and sorted according to their type and according to the descriptors previously mentioned. In total, 68 scientific publications were selected.

#### Results

Of the 68 selected publications that met the inclusion criteria, 32 are review articles, 11 are case-control investigations, 7 are in vitro studies, 3 are clinical trials, 2 are systematic reviews, 3 are cross-sectional studies, 2 are chapters of books, 2 are articles developed in the context of the 2017 world workshop on the current classification of periodontal and peri-implant diseases and conditions, 1 article on the impact of the global burden of periodontal diseases developed in the context of the Milan World Exposition 2015 "Feeding the planet, energy for life", 1 longitudinal study, 1 observational study, 2 experimental study with *in vitro* and *in vivo* models and 1 epidemiological report.

#### Periodontitis

Periodontitis is an inflammatory disease that causes loss of the periodontal junction in which the host and the microbiota are associated. During this disease, host derived proteinases are activated, which allow the loss of marginal fibers of the periodontal ligament, the apical migration of the binding epithelium and the apical propagation of the bacterial biofilm in the tooth root (13). This disease produces destruction of the soft and hard tissues that support the tooth (14), mobility of the teeth and ultimately the loss of the same (15). The diagnosis and classification of this pathology is carried out by traditional clinical measurements such as depth of the periodontal pocket, bleeding on probing (BOP), clinical attachment loss (CAL) and radiographic bone loss (16, 17).

The primary etiological factor of periodontitis are pathogenic microorganisms, while the immune response affects the progression of the disease (18), which arises from excessive inflammatory responses resulting from complex interactions between the host and the subgingival dysbiotic microbiota that involve elements both innate and adaptive immunity (19). Periodontopathogenic bacteria include: *Aggregatibacter actinomycetemcomitans (A. actinomycetemcomitans), Porphyromonas gingivalis (P. gingivalis), Tannerella forsythia (T. forsythia), Treponema denticola (T. denticola) and Prevotella intermedia (P. intermedia)* (20), as part of aggressive bacteria classified in colors by Socransky (21).

Periodontitis is characterized among many other aspects, by the production of prostaglandins E2 (PGE2), proinflammatory cytokines such as interleukin 1 (IL1), interleukin 6 (IL6), tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) and ROS. These factors promote tissue destruction by inhibiting collagen synthesis, activating matrix metalloproteinases (MMPs), blocking the activity of tissue inhibitors of matrix metalloproteinases (TIMP), and at the same time stimulating bone resorption of alveolar bone (22).

This pathology is associated with hyper-reactive neutrophils with an increased production of ROS in response to the stimulation of Fc-gamma receptors (Fc $\gamma$ R) (23), toll-like receptors (TLR) and cytokine receptors (24), resulting in periodontal tissue injury and activation of macrophages, neutrophils and fibroblasts to generate more ROS (22).

#### Free radicals and reactive oxygen species

Free radicals are atoms or groups of atoms that have an unpaired electron in their outermost orbit, which gives it high instability, reactivity and an enormous capacity to combine nonspecifically with biomolecules, producing different radical and non-radical species, which are generally harmful to humans, since they can alter the configuration of molecular structures, leading to damage or molecular and tissue

instability (25). When the free radical subtracts the electron it needs, a redox reaction takes place, in which the free radical is reduced and the stable molecule that has lost the electron is oxidized, becoming a free radical and thus initiating a chain reaction (26). As a product of metabolism, different types of free radicals are generated, such as: ROS and reactive nitrogen species (RNS) (25).

ROS are broadly defined as chemically reactive oxygen-containing molecules (27). They can be classified into free radicals and non-radical oxygen species involved in the production of oxygen radicals. Free radicals include the superoxide anion  $(0_2^{-})$ , hydroxyl (OH<sup>-</sup>), hydroperoxyl (HOO<sup>-</sup>), peroxyl (ROO<sup>-</sup>). Singlet oxygen  $({}^{1}O_2)$ , hypochlorous acid (HOCl) and hydrogen peroxide  $(H_2O_2)$  are non-radical oxygen species (26). El OH<sup>-</sup>,  $O_2^{-}$  and  $H_2O_2$  are of greater physiological importance (28).

#### Sources and formation of reactive oxygen species

The cells of the body are exposed to oxidants from endogenous and exogenous sources. Exogenous sources include heat, trauma, ionizing radiation, UV radiation, ozone, smoking, infection and metabolism of a broad spectrum of drugs and xenobiotic. Endogenous sources are mainly byproducts of metabolism by functional generation by host defense cells (phagocytes) and cells of connective tissues (22, 29).

The main endogenous source of ROS is the electron transport chain (ETC). From the mitochondrial respiratory complexes I and III in the ETC some electrons escape and react with oxygen to generate  $O_2^{-}(30)$ , which is transferred to the mitochondrial matrix through the inner mitochondrial membrane. Subsequently, by action of superoxide dismutase 2 (SOD2),  $O_2^{-}$  becomes  $H_2O_2$ .  $O_2^{-}$  is also produced in the cytosol by enzymatic reactions involving NADPH oxidases (NOX), xanthine oxidase, arachidonic acid, among others (27).  $H_2O_2$  is a much more stable molecule  $O_2^{-}$ , capable of diffusing through biological membranes. In the presence of metal cations, such as iron or copper,  $H_2O_2$  is converted to OH<sup>-</sup> through the Fenton reaction (31, 32). The OH<sup>-</sup> is extremely unstable and reactive; with high oxidizing power, it oxidizes lipids, proteins and DNA indiscriminately, resulting in damage or genomic instability (26).

#### **Oxidative stress**

When the production of ROS and RNS increases in such a way that the antioxidant response cannot restore the system, oxidative stress occurs (10). The term oxidative stress was defined in 1985 by Helmut Sies as a disturbance in the prooxidant-antioxidant balance in favor of the former, giving rise to potential damage (33, 34). Nowadays, oxidative stress is defined as a persistent imbalance between the production of highly reactive molecular species (oxidants) and antioxidants in favor of oxidants, which leads to an interruption of redox signaling and control and/or molecular damage (35). Oxidative stress can be divided into four ranges: basal oxidative stress, low intensity, intermediate and intense. Probably, low intensity oxidative stress is detected by the Nrf2/Keap system, which is activated by minimal amounts of ROS (36). The nuclear transcription factor NF-E2 related to factor 2 (Nrf2) translocates to the nucleus where it interacts with antioxidant response elements (ARE) in promoters of target genes that encode antioxidant defense enzymes (37). NF- $\kappa$ B, activating protein 1 (AP-1) and MAP kinases are involved in intermediate intensity oxidative stress through the expression of antioxidant enzymes and certain genes of inflammation and reprogramming of general cellular functions. Apoptosis or necrosis occurs in response to intense oxidative stress (36).

#### **Antioxidant systems**

Antioxidants are substances that are present at low concentrations and retard or significantly prevent the oxidation of an oxidizable substrate (38). Antioxidants can be classified according to their mode of action in preventive antioxidants and antioxidants that break chains (Scavenging) (34). Antioxidants have also been classified as enzymatic and non-enzymatic (28, 30, 39, 40). Table 1 summarizes some enzymatic antioxidants and Table 2 summarizes some non-enzymatic antioxidants.

Name	Function
Superoxide dismutase (SOD)(36)	$0_2^{-+} 0_2^{-+} 2H^* \rightarrow 0_2^{-} + H_2^{-} 0_2^{-}$
Catalase (CAT)(41)	$2H_2O_2 \rightarrow 2H_2O + O_2$
Glutathione peroxidase (GPx)(41)	$H_2O_2 + 2GSH \rightarrow H_2O + GSSG$
Glutathione reductase (GR)(41)	GSSG+NADPH+H <sup>+</sup> 2GSH+NADP <sup>+</sup>
Peroxiredoxin (Prx)(39)	It catalyzes the reduction of peroxides. The peroxides oxidize the cysteine from the catalytic site which then reacts with another cysteine residue to form a disulfide that is subsequently reduced by an electron donor
Thioredoxin (Trx)(39)	It reduces the oxidized proteins by forming disulfide bonds, transferring electrons from their reactive cysteines. The dithiol residues of Trx are reduced by NADPH electrons using thioredoxin reductase.

Table 1. Enzymatic Antioxidants

\*GSH (reduced glutathione), GSSG (glutathione disulfide), H2O2 (hydrogen peroxide), NADP+ (nicotinamide-adenine-dinucleotide-phosphate), NADPH (reduced nicotinamide-adenine-dinucleotide-phosphate), O2- (superoxide anion)

Name	Function
Vitamin C (42)	It works synergistically with vitamin E to quench free radicals. Regenerates the reduced form of vitamin E
Vitamin E(43)	It maintains the integrity of the cell membrane against lipid peroxidation by eliminating peroxyl radicals.
Carotenoids (42, 44)	eta-carotene is efficient in the elimination of singlet oxygen. It protects cell membranes and lipoproteins against the damage of peroxyl radicals.
Glutathione (45)	Direct purification of ROS and the restitution of other antioxidants such as vitamin E and ascorbic acid to their reduced forms. Works cyclically with GPx / GR and NADPH

**Table 1.** Non-enzymatic antioxidants

\*GPx (glutathione peroxidase), GR (glutathione reductase), NADPH (reduced nicotinamide-adenine-dinucleotide-phosphate), ROS (reactive oxygen species).

# Mechanism of action of reactive oxygen species and oxidative stress in tissue destruction in periodontitis

• Tissue destruction in periodontitis is considered to be the result of an excessive inflammatory response to the subgingival biofilm (12), since cellular components such as lipopolysaccharides (LPS) and the DNA of periodontopathogenic bacteria cause the activation of signaling pathways related to inflammation in gingival fibroblasts and the production of inflammatory cytokines such as interleukin-8 and TNF- $\alpha$ . Similarly, hyperreactive recruitment and activation of PMNs that produce ROS in response of the host to pathogens occurs (22). In this sense, neutrophils produce  $O_2^-$  by respiratory burst with the participation of the NOX enzyme. In turn  $O_2^-$  can be released from the phagosomal environment to the extracellular to become ROS such as OH<sup>-</sup>,  $1O_2$ , HOCl and  $H_2O_2$  (45).

The release of ROS plays a critical role in the destruction of tissue in periodontitis (12), these in excess cannot be balanced by the antioxidant defense system, which leads to oxidative stress and tissue damage in the periodontium (45). Oxidative stress can cause damage directly to the periodontal tissue and indirectly to it by activating cell signaling pathways related to inflammation, apoptosis and other factors (46). Direct tissue damage caused by ROS includes cytotoxic effects such as lipid and phospholipid peroxidation, oxidative damage to proteins and DNA, interfering with cell growth and cell cycle progression, inducing apoptosis of gingival fibroblasts and causing matrix degradation extracellular (MEC) through MMP induction and glycosaminogly-can breakdown (12), cell membrane destruction, protein denaturation and enzymatic inactivation, mitochondrial lesions and more ROS production (46). Protein damage involves folding or unfolding proteins, protein fragmentation and polymerization reactions, protein radical formation, formation of protein carbonylation products. DNA damage causes chain breaks, base mutations, guanine to 8-hydroxyguanine conversion, deletions, insertions and sequence amplification (22, 29).

Tissue destruction can be assessed by measuring biomarkers malondialdehyde (MDA), 8-hydroxysoxyguanosine (8-OHdG), carbonyl proteins (PC), total antioxidant capacity (TAOC), SOD, among others (45). Patients with periodontitis have higher levels of MDA (47), 8-OHdG (48), PC (49) and lower levels of SOD (50) and TAOC than healthy controls (51).

Likewise, this damage to periodontal tissue leads to the overproduction of lipid peroxides, inflammatory mediators and oxidized proteins, which further activate macrophages, neutrophils and fibroblasts to generate more ROS and create a vicious circle (22).

Indirect tissue damage caused by ROS occurs through the following mechanisms:

- ROS can promote osteoclastogenesis (12).
- Activation of NF-κB, initiating a signaling cascade that regulates inflammatory and immune responses.
- Activation of JNK, producing cellular apoptosis.
- ROS are associated with inflammasome activation resulting in piroptosis.

• ROS play a critical role in autophagy. However, there is still no direct evidence to show that the activation or inactivation of autophagy is triggered by the regulation of redox signaling in periodontitis (46).

ROS can indirectly promote osteoclastogenesis because they act as an intracellular signaling molecule during it. Alveolar bone destruction and resorption requires osteoclasts, in turn the generation of osteoclasts requires that NF- $\kappa$ B activating receptor ligand (RANKL) bind to its receptor (RANK) in the marrow macrophages which leads to differentiating them into osteoclasts. Under physiological conditions, RANKL is expressed mainly in mesenchymal cells of the osteoblast lineage, but in periodontitis it is abundantly produced by activated lymphocytes (12).

ROS activate or depress the NF- $\kappa$ B signaling pathway. Some studies have shown that ROS, in particular H<sub>2</sub>O<sub>2</sub>, can activate IKK (IkB kinase complex) in some cell types. In contrast, other investigations have shown that H<sub>2</sub>O<sub>2</sub> in some cells can inactivate IKK, in this sense the inhibitory effect can be mediated by the oxidation of IKK $\beta$  in cysteine 179 by ROS. It is presumed that NIK, the kinase involved in the non-canonical pathway, is activated by ROS through the inhibition of phosphatases. In addition, the direct oxidation of p50 by ROS inhibits its ability to bind to DNA (52). H<sub>2</sub>O<sub>2</sub> regulates the activation of NF- $\kappa$ B and does so in part through the alternative phosphorylation of I $\kappa$ B $\alpha$  in Tyr42 (53). The activation of NF- $\kappa$ B increases the concentration of MMP (22), promotes the expression of proinflammatory cytokines, these factors lead to inflammatory responses and osteoclastic differentiation and thereby the destruction of periodontal tissue (54).

Oxidative stress can also activate JNK signaling during periodontitis (46). It has been shown that JNK activation induced by ROS occurs through the regulatory kinase of the signal of apoptosis 1 (ASK-1) (55). ASK1 produces the subsequent activation (phosphorylation) of JNK, phosphorylated JNK activates apoptotic signaling either through overexpression of proapoptotic genes by transactivation of specific transcription factors or by directly modulating the activities of pro and antiapoptotic mitochondrial proteins by phosphorylation events (56). In the mitochondria JNK amplifies ROS production generated largely by Complex I (55). JNK can stimulate the release of cytochrome C from the inner mitochondrial membrane through Bid-Bax, promoting the formation of apoptosomes. In another mechanism, JNK can promote the release of Smac/Diablo (Smac) by inactivating the inhibitory complex TRAF2/IAP1 to initiate the activation of caspases (56). One study showed that the activation of JNK by ROS disrupted the periodontal junction epithelium through dissociation of E-cadherin (57).

ROS are involved in the pathogenesis of periodontitis through the activation of the NLRP3 inflammasome pathway (58). In this sense an increase in ROS concentrations activates the protein that interacts with thioredoxin (TXNIP) and this binds with the protein with pyrin 3 domain of the NLR family (NLRP3) activating the NLRP3 inflammasome (59, 60), which eventually upon activation of caspase-1 to cleave pro-IL-1 $\beta$  and pro-IL-18 in its active forms of secretion (IL-1 $\beta$  and IL-18) and induce pyroptosis (61-63). Elevated levels of IL-1 $\beta$  and IL-18 may contribute to periodontal destruction and inflammation (64).

ROS can activate or suppress the autophagy process, through the following potential mechanisms:

- Phosphorylation of the Bcl-2 apoptosis regulator by JNK in a ROS-dependent manner that leads to the dissociation of Beclin 1 (BECN1) and therefore induces autophagy.
- Initiation of the PI3K/AKT pathway, which leads to the activation of the rapamycin target protein in mammalian cells (mTOR), inhibiting autophagy.
- ROS excesses produce activation of the AMP-activated protein kinase (AMPK), which phosphorylates the tuberin protein (TSC2) and activates the TSC1/TSC2 complex, thus inhibiting the target protein of rapamycin complex1 (TORC1) and stimulating serine/threonine protein kinase (ULK), which is an important initiator of autophagy.
- Activation of the Atg12-Atg5 complex, which promotes the elongation of autophagy (46).

Together, the factors mediated by ROS that cause periodontal destruction directly or indirectly contribute to the development and progression of periodontitis (12, 22). Figure 1 summarizes the mechanisms of action of ROS, oxidative stress in tissue destruction in periodontitis.

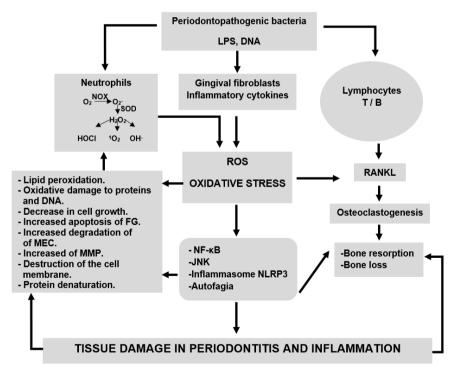


Figure 1. Mechanisms of action of ROS, oxidative stress in tissue destruction in periodontitis

The periodontopathogenic microbiota can activate immune system cells such as neutrophils, T and B lymphocytes and gingival fibroblasts. Activated neutrophils produce the superoxide anion ( $O_2$ ), using the enzyme NADPH-oxidase in the process of respiratory explosion. The  $O_2$  can be converted by superoxide dismutase or by spontaneous dismutation into hydrogen peroxide ( $H_1O_2$ ), which can be converted into other ROS including hypochlorous acid (HOCI), hydroxyl radical (OH) and singlet oxygen ( $O_2$ ). An increase in ROS produces oxidative stress, which together can cause direct and indirect damage to periodontal tissue. Direct damage includes lipid peroxidation, oxidative damage to proteins and DNA, decrease in cell growth, increased apoptosis of gingival fibroblasts (FG), increased degradation of the extracellular matrix (MEC), increased matrix metalloproteinases (MMP), destruction of the cell membrane and protein denaturation direct damage encludes stimulation of osteoclastogenesis, activation of the pathways signaling nuclear transcription factor kappa B (NF-KB), the N-terminal c-Jun kinase pathway (JNK), activation of Inflammasome NLRP3, activation or inhibition of autophagy. Activated T and B lymphocytes produce RANKL that generates osteoclastogenesis, promoting bone resorption and bone loss. In turn, bacterial cellular components such as DNA and lipopolysaccharides (LPS) stimulate the activation of signaling pathways related to inflammation in gingival fibroblasts and the production of inflammatory cytokines. All these factors play an essential role in the destruction of periodontal tissue and create a vice circle of inflammation that produces even more ROS.

## Discussion

The association between oxidative stress and periodontitis has been the subject of review-type investigations, this article has tried to describe and give a possible explanation of this association through the mechanisms of action that allow ROS and oxidative stress to originate and establish periodontal destruction, reflecting that these factors are fundamental in the pathogenesis of periodontitis. Similarly, Wang et al. report that periodontitis is associated with a hyperactivity of peripheral blood neutrophils that are the main source of ROS and oxidative stress is implicated in the pathogenesis of periodontitis (45). In the same way, Dahiya et al., Liu et al. establish that oxidative stress is fundamental in the damage to periodontal tissue that results from interactions between the host and the microbiota due to increased ROS, antioxidant deficiency or by activating signaling pathways sensitive to redox changes and creating a proinflammatory state (22, 46). This differs with Tóthová et al. who expressed that despite decades of research the role that oxidative stress plays in periodontitis is unclear (11).

Previously, it was mentioned that ROS cause lipid peroxidation, oxidative damage to proteins and DNA, so research has been carried out that evaluates the different biomarkers of oxidative stress in periodontitis. A widely studied biomarker is MDA, which is a product of lipid peroxidation (14). Some research has studied MDA levels in patients with periodontitis. Trivedi et al. conducted a study in which spectrophotometrically analyzed MDA levels in individuals with type 2 diabetes mellitus with and without periodontitis and periodontally healthy patients with and without type 2 diabetes mellitus, in this study it was shown that MDA levels in both groups of periodontitis were higher than in periodontally healthy groups, concluding that the body's antioxidant mechanisms are partially collapsed due to excessive free radical production during periodontitis (47), which supports the results of Almerich-Silla et al. in which significantly higher levels of MDA are reflected in patients with periodontitis than in healthy controls and gingivitis, demonstrating significant changes in oxidative stress by measuring different markers of oxidative stress that increased with the deterioration of the periodontal state (16). High levels of serum and salivary MDA, without changes in antioxidant status, can cause systemic and local complications in patients with periodontitis (65). On the other hand Baltacioğlu et al. They reported that there is no significant difference in serum MDA levels, but in salivary of this same biomarker in patients with periodontitis and healthy controls (66).

A biomarker of oxidative damage of DNA that has been studied in periodontitis is 8-OHdG. The results published by Villa - Correa et al. showed higher salivary levels of 8-OHdG in individuals with periodontitis than in healthy controls, this shows that the increase in levels in this biomarker may be a prognostic indicator of periodontal destruction induced by oxidative stress (48). These results are consistent with those presented by Zamora-Pérez et al. with higher levels of salivary 8-OHdG in periodontitis, which indicates a direct relationship with DNA damage (67).

Oxidative damage to proteins can cause their denaturation and therefore the loss of their function (46), one way to estimate this damage is by measuring PC, which is higher in patients with periodontitis, This suggests that there is an increase in oxidative stress during periodontitis (49).

Antioxidants have also been evaluated in patients with periodontitis. Trivedi et al. compared salivary levels of SOD in subjects with periodontitis and healthy, revealing lower measurements of this antioxidant enzyme in the periodontitis group than in

the control group, which evidences that oxidative stress plays a fundamental role in the pathogenesis of periodontitis (68). Similarly Canakci et al. support less SOD activity in periodontitis (50).

## Conclusion

Oxidative stress (excess ROS) plays a critical role in the development and evolution of tissue injury and periodontal inflammation, which leads to tissue destruction and creation of a proinflammatory state in periodontitis. This is because ROS are capable of stimulating the production of proinflammatory molecules, regulate apoptosis, activate immune cells that produce more ROS, creating a vice circle of inflammation and enhancing tissue injury. In addition, ROS stimulates osteoclastogenesis, producing bone resorption, which would lead to tooth mobility and long-term tooth loss. However, it cannot be inferred whether oxidative stress is a factor prior to the development of periodontitis or if it is only triggered as a result of periodontal inflammation. Therefore, investigations of oxidative damage in this pathology and the development of biomarkers of oxidative stress could be useful for the monitoring and evaluation of periodontal treatment.

On the other hand, observational and experimental research on these topics is required to be more homogeneous in terms of the type of sample and the methods used to estimate and measure the different oxidative stress biomarkers.

#### **Complications and difficulties**

There are several publications on oxidative rhinitis and periodontitis, but these present a great deal of variability in the protocols and methods for determining periodontal and oxidative stress parameters. In addition to the above, the sample size is reduced and there are few publications of systematic reviews on the relationship between periodontitis and oxidative stress.

#### Acknowledgment

Acknowledgment to the School of Medicine and School of Dentistry of the University of Cartagena.

#### **Declaration of conflict of interests**

The authors of this manuscript declare they have no conflicts of interest.

#### References

- Andrei M, Dinischiotu A, Didilescu AC, Ionita D, Demetrescu I. Periodontal materials and cell biology for guided tissue and bone regeneration. Ann Anat. 2018;216:164-169.
- Ekuni D, Firth JD, Putnins EE. Expression of Reactive Oxygen Species in Junctional and Pocket Epithelium. In: Ekuni D, Battino M, Tomofuji T, Putnins EE, editors. Studies on Periodontal Disease. New York, NY: Springer New York; 2014. p. 53-75.
- Caton JG, Armitage G, Berglundh T, Chapple ILC, Jepsen S, Kornman KS, et al. A new classification scheme for periodontal and peri-implant diseases and conditions – Introduction and key changes from the 1999 classification. Journal of Clinical Periodontology. 2018;45(S20):S1-S8.

- 4. Tonetti MS, Jepsen S, Jin L, Otomo-Corgel J. Impact of the global burden of periodontal diseases on health, nutrition and wellbeing of mankind: A call for global action. Journal of Clinical Periodontology. 2017;44(5):456-462.
- Colombia. Ministerio de Salud y Protección Social. IV Estudio Nacional de Salud Bucal. ENSAB IV: Para saber cómo estamos y saber qué hacemos. [Available from: <u>https://www.minsalud.gov.co/sites/rid/Lists/BibliotecaDigital/RIDE/VS/</u> <u>PP/ENSAB-IV-Situacion-Bucal-Actual.pdf</u>.
- 6. Le Sage F, Meilhac O, Gonthier MP. Porphyromonas gingivalis lipopolysaccharide induces pro-inflammatory adipokine secretion and oxidative stress by regulating Toll-like receptor-mediated signaling pathways and redox enzymes in adipocytes. Mol Cell Endocrinol. 2017;446:102-110.
- 7. Preshaw PM, Alba AL, Herrera D, Jepsen S, Konstantinidis A, Makrilakis K, et al. Periodontitis and diabetes: a two-way relationship. Diabetologia. 2012; 55(1):21-31.
- 8. Muñoz-Torres FJ, Mukamal KJ, Pai JK, Willett W, Joshipura KJ. Relationship between tooth loss and peripheral arterial disease among women. J Clin Periodontol. 2017;44(10):989-995.
- 9. D'Aiuto F, Nibali L, Parkar M, Patel K, Suvan J, Donos N. Oxidative stress, systemic inflammation, and severe periodontitis. J Dent Res. 2010;89(11):1241-1246.
- 10. Acquier AB, De Couto Pita AK, Busch L, Sánchez GA. Parameters of oxidative stress in saliva from patients with aggressive and chronic periodontitis. Redox Rep. 2017;22(3):119-126.
- 11. Tóthová L, Celec P. Oxidative Stress and Antioxidants in the Diagnosis and Therapy of Periodontitis. Front Physiol. 2017;8:1055.
- 12. Kanzaki H, Wada S, Narimiya T, Yamaguchi Y, Katsumata Y, Itohiya K, et al. Pathways that Regulate ROS Scavenging Enzymes, and Their Role in Defense Against Tissue Destruction in Periodontitis. Front Physiol. 2017;8:351.
- 13. Tonetti MS, Greenwell H, Kornman KS. Staging and grading of periodontitis: Framework and proposal of a new classification and case definition. Journal of Periodontology. 2018;89(S1):S159-S72.
- Ambati M, Rani KR, Reddy PV, Suryaprasanna J, Dasari R, Gireddy H. Evaluation of oxidative stress in chronic periodontitis patients following systemic antioxidant supplementation: A clinical and biochemical study. J Nat Sci Biol Med. 2017;8(1):99-103.
- 15. Simpson TC, Weldon JC, Worthington HV, Needleman I, Wild SH, Moles DR, et al. Treatment of periodontal disease for glycaemic control in people with diabetes mellitus. Cochrane Database Syst Rev. 2015;2015(11):Cd004714.
- Almerich-Silla JM, Montiel-Company JM, Pastor S, Serrano F, Puig-Silla M, Dasí F. Oxidative Stress Parameters in Saliva and Its Association with Periodontal Disease and Types of Bacteria. Dis Markers. 2015;2015:653537.

- 17. Srivastava N, Nayak PA, Rana S. Point of Care- A Novel Approach to Periodontal Diagnosis-A Review. J Clin Diagn Res. 2017;11(8):Ze01-ze6.
- Cekici A, Kantarci A, Hasturk H, Van Dyke TE. Inflammatory and immune pathways in the pathogenesis of periodontal disease. Periodontology 2000. 2014; 64(1):57-80.
- 19. Hajishengallis G, Korostoff JM. Revisiting the Page & Schroeder model: the good, the bad and the unknowns in the periodontal host response 40 years later. Periodontol 2000. 2017;75(1):116-151.
- 20. Bozoglan A, Ertugrul AS, Taspinar M, Yuzbasioglu B. Determining the relationship between atherosclerosis and periodontopathogenic microorganisms in chronic periodontitis patients. Acta Odontol Scand. 2017;75(4):233-242.
- 21. Torrungruang K, Jitpakdeebordin S, Charatkulangkun O, Gleebbua Y. Porphyromonas gingivalis, Aggregatibacter actinomycetemcomitans, and Treponema denticola / Prevotella intermedia Co-Infection Are Associated with Severe Periodontitis in a Thai Population. PLoS One. 2015;10(8):e0136646.
- 22. Dahiya P, Kamal R, Gupta R, Bhardwaj R, Chaudhary K, Kaur S. Reactive oxygen species in periodontitis. J Indian Soc Periodontol. 2013;17(4):411-116.
- 23. Ling MR, Chapple ILC, Matthews JB. Neutrophil superoxide release and plasma C-reactive protein levels pre- and post-periodontal therapy. Journal of Clinical Periodontology. 2016;43(8):652-658.
- 24. White PC, Chicca IJ, Cooper PR, Milward MR, Chapple ILC. Neutrophil Extracellular Traps in Periodontitis: A Web of Intrigue. Journal of Dental Research. 2016;95(1):26-34.
- 25. Losada-Barreiro S, Bravo-Díaz C. Free radicals and polyphenols: The redox chemistry of neurodegenerative diseases. Eur J Med Chem. 2017;133:379-402.
- 26. Phaniendra A, Jestadi DB, Periyasamy L. Free radicals: properties, sources, targets, and their implication in various diseases. Indian J Clin Biochem. 2015;30(1):11-26.
- 27. Zou Z, Chang H, Li H, Wang S. Induction of reactive oxygen species: an emerging approach for cancer therapy. Apoptosis. 2017;22(11):1321-1335.
- 28. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. World Allergy Organ J. 2012;5(1):9-19.
- 29. Chapple ILC, Matthews JB. The role of reactive oxygen and antioxidant species in periodontal tissue destruction. Periodontology 2000. 2007;43(1):160-232.
- 30. Ye ZW, Zhang J, Townsend DM, Tew KD. Oxidative stress, redox regulation and diseases of cellular differentiation. Biochim Biophys Acta. 2015;1850(8): 1607-1621.

- 31. Reczek CR, Chandel NS. ROS-dependent signal transduction. Curr Opin Cell Biol. 2015;33:8-13.
- 32. Lee M-C-i. Reactive Oxygen Species and Antioxidant Systems in Periodontal Disease. In: Ekuni D, Battino M, Tomofuji T, Putnins EE, editors. Studies on Periodontal Disease. New York, NY: Springer New York; 2014. p. 3-14.
- 33. Sies H. Oxidative stress: a concept in redox biology and medicine. Redox Biol. 2015;4:180-183.
- 34. Trivedi S, Lal N. Antioxidant enzymes in periodontitis. J Oral Biol Craniofac Res. 2017;7(1):54-57.
- 35. Chapple IL, Milward MR, Ling-Mountford N, Weston P, Carter K, Askey K, et al. Adjunctive daily supplementation with encapsulated fruit, vegetable and berry juice powder concentrates and clinical periodontal outcomes: a double-blind RCT. J Clin Periodontol. 2012;39(1):62-72.
- 36. Lushchak VI. Free radicals, reactive oxygen species, oxidative stress and its classification. Chem Biol Interact. 2014;224:164-175.
- 37. Ma Q. Role of nrf2 in oxidative stress and toxicity. Annu Rev Pharmacol Toxicol. 2013;53:401-426.
- 38. Pendyala G, Thomas B, Kumari S. The challenge of antioxidants to free radicals in periodontitis. J Indian Soc Periodontol. 2008;12(3):79-83.
- 39. Kehrer JP, Klotz LO. Free radicals and related reactive species as mediators of tissue injury and disease: implications for Health. Crit Rev Toxicol. 2015;45(9): 765-798.
- 40. Muniz FW, Nogueira SB, Mendes FL, Rösing CK, Moreira MM, de Andrade GM, et al. The impact of antioxidant agents complimentary to periodontal therapy on oxidative stress and periodontal outcomes: A systematic review. Arch Oral Biol. 2015;60(9):1203-1214.
- 41. Arana C, Moreno-Fernández AM, Gómez-Moreno G, Morales-Portillo C, Serrano-Olmedo I, de la Cuesta Mayor MC, et al. Increased salivary oxidative stress parameters in patients with type 2 diabetes: Relation with periodontal disease. Endocrinol Diabetes Nutr. 2017;64(5):258-264.
- 42. Kumar S, Sharma S, Vasudeva N. Review on antioxidants and evaluation procedures. Chin J Integr Med. 2017.
- 43. Rizvi S, Raza ST, Ahmed F, Ahmad A, Abbas S, Mahdi F. The role of vitamin e in human health and some diseases. Sultan Qaboos Univ Med J. 2014;14(2):e157-165.
- 44. Kaulmann A, Bohn T. Carotenoids, inflammation, and oxidative stress--implications of cellular signaling pathways and relation to chronic disease prevention. Nutr Res. 2014;34(11):907-929.

- 45. Wang Y, Andrukhov O, Rausch-Fan X. Oxidative Stress and Antioxidant System in Periodontitis. Front Physiol. 2017;8:910.
- 46. Liu C, Mo L, Niu Y, Li X, Zhou X, Xu X. The Role of Reactive Oxygen Species and Autophagy in Periodontitis and Their Potential Linkage. Front Physiol. 2017;8:439.
- 47. Trivedi S, Lal N, Mahdi AA, Mittal M, Singh B, Pandey S. Evaluation of Antioxidant Enzymes Activity and Malondialdehyde Levels in Patients With Chronic Periodontitis and Diabetes Mellitus. Journal of Periodontology. 2014;85(5):713-720.
- Villa-Correa YA, Isaza-Guzmán DM, Tobón-Arroyave SI. Prognostic Value of 8-Hydroxy-2'-Deoxyguanosine and Human Neutrophil Elastase/α1-Proteinase Inhibitor Complex as Salivary Biomarkers of Oxidative Stress in Chronic Periodontitis. Journal of Periodontology. 2015;86(11):1260-1267.
- 49. Pradeep AR, Ramchandraprasad MV, Bajaj P, Rao NS, Agarwal E. Protein carbonyl: An oxidative stress marker in gingival crevicular fluid in healthy, gingivitis, and chronic periodontitis subjects. Contemp Clin Dent. 2013;4(1):27-31.
- 50. Canakci CF, Cicek Y, Yildirim A, Sezer U, Canakci V. Increased levels of 8-hydroxydeoxyguanosine and malondialdehyde and its relationship with antioxidant enzymes in saliva of periodontitis patients. Eur J Dent. 2009;3(2):100-106.
- 51. Abou Sulaiman AE, Shehadeh RM. Assessment of total antioxidant capacity and the use of vitamin C in the treatment of non-smokers with chronic periodontitis. J Periodontol. 2010;81(11):1547-1554.
- 52. Morgan MJ, Liu Z-g. Crosstalk of reactive oxygen species and NF-κB signaling. Cell Research. 2011;21(1):103-115.
- 53. Lin Y, Bai L, Chen W, Xu S. The NF-kappaB activation pathways, emerging molecular targets for cancer prevention and therapy. Expert Opin Ther Targets. 2010;14(1):45-55.
- Huang X, Chen H, Xie Y, Cao Z, Lin X, Wang Y. FoxO1 Overexpression Ameliorates TNF-α-Induced Oxidative Damage and Promotes Osteogenesis of Human Periodontal Ligament Stem Cells via Antioxidant Defense Activation. Stem Cells Int. 2019;2019:2120453.
- 55. Chambers JW, LoGrasso PV. Mitochondrial c-Jun N-terminal kinase (JNK) signaling initiates physiological changes resulting in amplification of reactive oxygen species generation. J Biol Chem. 2011;286(18):16052-16062.
- 56. Dhanasekaran DN, Reddy EP. JNK signaling in apoptosis. Oncogene. 2008;27(48):6245-6251.
- 57. Lee G, Kim HJ, Kim HM. RhoA-JNK Regulates the E-Cadherin Junctions of Human Gingival Epithelial Cells. J Dent Res. 2016;95(3):284-291.
- Kurita-Ochiai T, Jia R, Cai Y, Yamaguchi Y, Yamamoto M. Periodontal Disease-Induced Atherosclerosis and Oxidative Stress. Antioxidants (Basel). 2015;4(3): 577-590.

- 59. Abais JM, Xia M, Zhang Y, Boini KM, Li PL. Redox regulation of NLRP3 inflammasomes: ROS as trigger or effector? Antioxid Redox Signal. 2015;22(13):1111-1129.
- 60. Zhou R, Tardivel A, Thorens B, Choi I, Tschopp J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. Nat Immunol. 2010; 11(2):136-140.
- 61. Lei-Leston AC, Murphy AG, Maloy KJ. Epithelial Cell Inflammasomes in Intestinal Immunity and Inflammation. Front Immunol. 2017;8:1168.
- 62. Westbom C, Thompson JK, Leggett A, MacPherson M, Beuschel S, Pass H, et al. Inflammasome Modulation by Chemotherapeutics in Malignant Mesothelioma. PLoS One. 2015;10(12):e0145404.
- 63. Sakhon OS, Victor KA, Choy A, Tsuchiya T, Eulgem T, Pedra JH. NSD1 mitigates caspase-1 activation by listeriolysin O in macrophages. PLoS One. 2013;8(9):e75911.
- 64. Bostanci N, Emingil G, Saygan B, Turkoglu O, Atilla G, Curtis MA, et al. Expression and regulation of the NALP3 inflammasome complex in periodontal diseases. Clin Exp Immunol. 2009;157(3):415-422.
- 65. Ahmadi-Motamayel F, Goodarzi MT, Jamshidi Z, Kebriaei R. Evaluation of Salivary and Serum Antioxidant and Oxidative Stress Statuses in Patients with Chronic Periodontitis: A Case-Control Study. Front Physiol. 2017;8:189.
- 66. Baltacıoğlu E, Yuva P, Aydın G, Alver A, Kahraman C, Karabulut E, et al. Lipid Peroxidation Levels and Total Oxidant/Antioxidant Status in Serum and Saliva From Patients With Chronic and Aggressive Periodontitis. Oxidative Stress Index: A New Biomarker for Periodontal Disease? Journal of Periodontology. 2014;85(10):1432-1441.
- 67. Zamora-Perez AL, Ortiz-García YM, Lazalde-Ramos BP, Guerrero-Velázquez C, Gómez-Meda BC, Ramírez-Aguilar MÁ, et al. Increased micronuclei and nuclear abnormalities in buccal mucosa and oxidative damage in saliva from patients with chronic and aggressive periodontal diseases. Journal of Periodontal Research. 2015;50(1):28-36.
- 68. Trivedi S, Lal N, Mahdi AA, Singh B, Pandey S. Association of salivary lipid peroxidation levels, antioxidant enzymes, and chronic periodontitis. Int J Periodontics Restorative Dent. 2015;35(2):e14-19.