Nasal and vaginal colonization of methicillin-resistant
Staphylococcus aureus in pregnant women in Cartagena, Colombia

Colonización nasal y vaginal por Staphylococcus aureus resistente a meticilina en mujeres embarazadas en Cartagena, Colombia

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Summary

Introduction: The host niche for Staphylococcus aureus (SA) are the anterior nares; however, vaginal colonization rates between 14% and 17.1% in pregnant women have been recently reported, raising interest about the potential risk in postpartum women and in neonates from colonized mothers.

Objectives: To determine the prevalence of nasal and vaginal colonization of SA and the antibiotic susceptibility of the isolates in pregnant women attending a maternity hospital in Cartagena, Colombia.

Methods: Nasal and vaginal swabs were obtained from participants and subjected to microbiological and molecular assays. A post discharge follow-up was performed for up to four weeks.

Results: From 100 pregnant women enrolled in the study, 34 were colonized with SA; 29 only in the nares, three only in the vagina, and two at both sites. Colonization of pregnant women with SA was more common in the nares than in the vagina or at both sites [29/34 (85.3%) vs 3/34 (8.8%) and 2/34 (5.9%); p<0.05]. We obtained 36 SA isolates, nine (25%) of which were methicillin-resistant Staphylococcus aureus (MRSA), one was from the vagina; thus, the overall MRSA colonization rate among pregnant women was 9%. Molecular analysis showed that Panton-Valentine leukocidin (PVL) genes were carried by the vaginal MRSA, seven of the nasal MRSA, and two of the Methicillin-sensitive Staphylococcus aureus (MSSA) isolates. Two MRSA isolates carried SCCmec type I and seven carried SCCmec type IV.

Conclusions: Nasal colonization rate for SA in the study population was similar to previous reports. However, the frequency of nasal colonization of MRSA was higher while vaginal colonization of SA was lower than previously reported in other studies for similar populations. The MRSA isolates obtained showed a community profile.

Keywords: Staphylococcus aureus; Methicillin-resistance; Maternal colonization; Virulence factor; Pregnancy.

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Resumen

Introducción: Staphylococcus aureus (SA) ha adoptado como nicho habitual en las narinas anteriores; sin embargo, recientemente se han reportado tasas de colonización del tracto genital de mujeres embarazadas entre 14% y 17.1%, lo que aumenta el interés respecto al riesgo asociado para el neonato y la madre en el posparto.

Objetivo: Determinar la prevalencia de colonización nasal y vaginal por SA y los patrones de susceptibilidad a antibióticos de los aislamientos en una comunidad obstétrica de Cartagena, Colombia.

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Métodos: Se obtuvieron hisopados nasales y vaginales de las participantes y se sometieron a ensayos microbiológicos y moleculares. Se realizó seguimiento a la madre y al neonato durante cuatro semanas.

Resultados: De 100 embarazadas participantes, 34 estuvieron colonizadas con SA; 29 solo en fosas nasales, tres solo vaginal y dos en ambos sitios. La colonización fue más común en fosas nasales que en vagina o en ambos sitios [29/34 (85.3%) vs 3/34 (8.8%) y 2/34 (5.9%); p < 0.05]. Se obtuvieron 36 aislamientos de SA de los cuales 9 fueron MRSA (25%), proviniendo uno de vagina; la tasa de colonización total por MRSA en embarazadas fue 9%. Los genes para la leucocidina de Panton-Valentine (PVL) se detectaron en la cepa MRSA vaginal, siete MRSA nasales, y dos MSSA. Dos aislamientos MRSA portaban el elemento SCCmec tipo I y siete el tipo IV. No se detectó resistencia a otros antibióticos en los aislamientos MRSA; tres aislamientos susceptibles a meticilina (MSSA) fueron resistentes a eritromicina.

Conclusiones: Aunque la colonización nasal por SA en la población estudiada estuvo dentro del rango reportado previamente, la colonización nasal por MRSA fue mayor, mientras que la colonización vaginal fue más baja que las informadas previamente en otros estudios para poblaciones similares. Los aislamientos MRSA obtenidos presentaron un perfil comunitario.

Palabras clave: Staphylococcus aureus; Resistencia a meticilina; Factores de virulencia; Embarazo; Colonización.

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Staphylococcus aureus (SA) is an important human pathogen responsible for nosocomial and community-acquired infections, which also behaves as commensal flora in healthy individuals – mainly colonizing the anterior nares. Carriers of SA, in particular methicillin-resistant Staphylococcus aureus (MRSA), have a higher risk for developing clinical infections, being the infections caused by MRSA strains the most important at the clinical level because they are more difficult to treat. In recent decades, MRSA strains, especially those associated to the community [community-associated methicillin-resistant Staphylococcus aureus (CA-MRSA)] have become an important issue in public health, given that they are a cause of increased morbidity and mortality. These CA-MRSA strains cause severe skin and soft tissue infections, necrotizing pneumonia, and sepsis in otherwise healthy children, teens, and, more recently, in neonates. The increasing number of infections in neonates caused by CA-MRSA emphasizes the need to identify the possible environmentally or maternally derived sources of infection.

For pregnant women, SA also poses a health risk because it is the main cause of infection of the surgical site, causing between 25% and 50% of infections of the post cesarean surgical site, representing a major cause of morbidity and a cause of puerperal mastitis. Early epidemiological studies showed that 5% of women were colonized with SA in their genital tract and postpartum women had the highest colonization rates; furthermore, vaginal-rectal carriage of SA has been found associated with development of postpartum fever. Although risk factors associated to colonization with MRSA strains during pregnancy have not been fully characterized, associations with race, parity, type of birth, and colonization with group B streptococci have been suggested. It is known that incidence of CA-MRSA infections varies among different communities and populations, and apparently, pregnant women are more susceptible and have risk factors that predispose them to developing these infections. Nevertheless, there is a scarcity of epidemiologic reports about MRSA infections present in pregnant and puerperal women.

More recently, several studies, mainly from the United States, have reported vaginal colonization rates for SA in pregnant women from 14% to 17.1%, and although the risk of vertical transmission has been suggested or even demonstrated, the association of such colonization with infant outcome has not been clearly established.

Thus, there is a need to increase knowledge in this area of research. A literature review of the subject reveals few published studies in this area, which are mainly reviews of clinical cases, making clear that...
more epidemiology-based studies are needed, given the increasing importance of CA-MRSA strains in serious neonatal infections and in women in puerperal stage.

Antibiotic resistance of MRSA strains is due to the acquisition of mecA gene, which codes for PBP-2a, a penicillin-binding protein with low binding affinity for beta-lactam antibiotics. The MecA gene is located in a mobile genetic element, the chromosomal staphylococcal cassette (SCCmec) of which seven major types have been identified, from I to VII. SCCmec types IV, V and VII are the most frequently found in CA-MRSA isolates, which also frequently carry the genes for Panton-Valentine leukocidin (PVL).

Up to 96% of MRSA infections in pregnant women mainly affect the skin and soft tissue, although several cases of pneumonia in the puerperal period have also been attributed to MRSA strains. Because PVL-carrying MRSA strains have been implicated in the development of serious skin and soft tissue infections and in necrotizing pneumonia, in the present study besides determining the SA nasal and vaginal colonization rates and antibiotic profiles of the isolates, we evaluated the presence of PVL genes and SCCmec types of MRSA isolates by multiplex PCR.

**Methods**

**Design, study population, and inclusion criteria.** This was a pilot study performed by the Genetics and Molecular Biology Research Group at Universidad de Cartagena. The study site, the Clínica de Maternidad Rafael Calvo (CMRC), in Cartagena, Colombia, is a university-affiliated maternity clinic attending pregnant women from the urban and rural areas around Cartagena, and whose obstetric service delivers between 8,000 and 8,500 infants per year. Following a research protocol approved by the Ethics Committee of Universidad de Cartagena and the Institutional Review Board of CMRC, women with at least 35 weeks of gestational age, who attended the outpatient or emergency departments during the months of January through June 2009 were enrolled in the study after signing an informed consent. Women with ruptured membranes and those under antibiotic treatment or with clinical evidence suggesting a current staphylococcal infection were excluded from the study.

By following approaches previously described by Pinter et al. and Beigi et al. nasal and vaginal swabs were obtained from the pregnant women enrolled (n=100) with vaginal samples taken from the outer third portion of the vagina. Although the objective of the study was to determine SA nasal and vaginal colonization rates and antibiotic susceptibility, we tried to ascertain any possible associations of colonization with complications related to staphylococcal infection either in the mothers or their newborns implementing a post discharge weekly follow-up by telephone for up to four weeks. Mothers were contacted by telephone at least once a week during the baby’s first month of life. They were asked of the general health of their infants, as well as of their own health. Specific questions were asked to determine whether any fevers, rashes, or other skin lesions had occurred in the infant. Additionally, participants were asked to file a questionnaire about social demographic data and medical records.

**Laboratory methods.** Nasal and vaginal specimens were transported to the microbiology laboratory of the School of Medicine at Universidad de Cartagena and processed within 8 to 18 hours, according to protocols described by Bettin et al. Antibiotic susceptibility of isolates confirmed as SA was performed by the disc diffusion method following recommendations of the Clinical and Laboratory Standards Institute (CLSI). The antibiotics evaluated were: rifampin (5 µg), clindamycin (2 µg), erythromycin (15 µg), gentamicin (10 µg), vancomycin (30 µg), cefoxitin (30 µg), and oxacillin (1 µg). The SA strain ATCC 33591 was used as control. The D-zone test for inducible clindamycin resistance was performed for each isolate according to CLSI method. Isolates were classified as MRSA if they demonstrated resistance to cefoxitin and MSSA if they were suscep-
tible. MRSA were further confirmed by PCR amplification of *mecA* gene. Both MSSA and MRSA isolates were stored at -35°C for subsequent molecular studies.

**Molecular analysis**

*Genomic DNA extraction.* Genomic DNA from each isolate was obtained according to the protocol described by Millar *et al.* with some modifications. Briefly, each SA isolate was sub-cultured in nutrient agar for 24 h at 37°C. Around five colonies were resuspended in 1 ml of Tris 0.5 M, centrifuged at 13,000 rpm x 5 min. Supernatant was discarded and the pellet resuspended in 500 µl buffer TE (10 mM Tris; 1 mM EDTA, pH: 8.0) and boiled at 100°C for 30 min, and then incubated at -35°C for 20 min, thawed at 65°C and finally centrifuged at 13,000 rpm for 15 min. Supernatant containing bacterial DNA was collected in a clean tube and stored at -20°C for subsequent PCR assays.

*Multiplex PCR assays.* Multiplex polymerase chain reaction assays for assessments of presence of the *lukF-PV* (encoding part of the PVL toxin), *mecA*, and *nuc* genes were performed for all SA isolates by using previously described primers. SA strains ATCC 33591 (*mecA*+; *nuc*+; PVL-) and ATCC 25923 (*mecA*; *nuc*+; PVL+) were used as amplification controls and pure water as negative controls. All SA isolates were subjected to multiplex PCR by using a set of three primer pairs previously reported: *MecA1F-MecA2R* that amplifies a 147-bp fragment of *mecA* gene; *Nuc1F-Nuc2R* that amplifies a 300-bp fragment of *nuc* gene specific for SA, and *LukPV1F-LukPV2R* that amplifies a 437-bp fragment of PVL gene. Additionally, all confirmed MRSA isolates were subjected to SCC*mec* typing by using a multiplex PCR assay, according to the protocol described by Zhang *et al.* by using SA strains NCTC10442 for SCC*mec* type I, N315 for SCC*mec* type II, and JCSC4744 for SCC*mec* type IV. PCR products were visualized in a 2% agarose gel stained with ethidium bromide under UV transillumination.

**Statistical analysis.** Statistical analysis was performed with SPSS version 13.0 for Windows. Based on microbiological results, the study population was classified as carrier and non-carrier and subclassified according to the colonization site (vagina/nares/both) and the frequency of colonization was determined for each of the two anatomical sites studied. Univariate analysis was applied to determine association of colonization to potential risk factors by using the Fisher exact test using a *p* value ≤0.05 for statistical significance.

### Table 1
**General demographic characteristics of the study population (n=100)**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Age (years)</strong></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>23.3</td>
</tr>
<tr>
<td>SD</td>
<td>± 5</td>
</tr>
<tr>
<td><strong>Gestational age (weeks)</strong></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>37.7</td>
</tr>
<tr>
<td>SD</td>
<td>± 2</td>
</tr>
<tr>
<td><strong>Geographical origin</strong></td>
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</tr>
<tr>
<td>Urban</td>
<td>74</td>
</tr>
<tr>
<td>Rural</td>
<td>26</td>
</tr>
<tr>
<td><strong>Marital status</strong></td>
<td></td>
</tr>
<tr>
<td>Single</td>
<td>13</td>
</tr>
<tr>
<td>Married</td>
<td>10</td>
</tr>
<tr>
<td>Unmarried partner</td>
<td>77</td>
</tr>
<tr>
<td><strong>Schooling</strong></td>
<td></td>
</tr>
<tr>
<td>Primary</td>
<td>42</td>
</tr>
<tr>
<td>Secondary</td>
<td>22</td>
</tr>
<tr>
<td>Technical</td>
<td>21</td>
</tr>
<tr>
<td>University</td>
<td>4</td>
</tr>
<tr>
<td>None</td>
<td>11</td>
</tr>
<tr>
<td><strong>Parity</strong></td>
<td></td>
</tr>
<tr>
<td>Nullipara</td>
<td>35</td>
</tr>
<tr>
<td>Multipara</td>
<td>65</td>
</tr>
</tbody>
</table>

SD: Standard deviation
Results and discussion

Table 1 shows demographic data from our study population and Table 2 shows the general results of SA isolates from this study. One hundred pregnant women were enrolled in this pilot study during a 6-month period and 34 of them were colonized with SA. Twenty nine of them were colonized only in the nares, three were colonized only in the vagina, and two harbored *S. aureus* at both sites. Colonization of pregnant women with SA was more common in the nares than in the vagina or at both sites \(29/34 (85.3\%) vs 3/34 (8.8\%)\) and \(2/34 (5.9\%); p<0.05\). From the total of participants, we obtained 36 SA isolates, nine of which (25\%) were MRSA, one was from the vagina; thus, the overall MRSA colonization rate among pregnant women was 9\%.

The nasal colonization rate for SA found in our pilot study (29\%) was similar to rates previously reported for the general population\(^2\,\,^2\,\,^2\). However, the nasal colonization by MRSA (8\% of the overall study population, 27.6\% of the nasal SA isolates) was higher than the frequencies previously described both at international and national levels\(^2\,\,^2\,\,^2\). Albeit, at local level there is a report of similar nasal colonization rates for MRSA in school-age children\(^2\,\,^3\); in other local reports, MRSA nasal isolates were not detected among elderly individuals residing in a nursing-home\(^1\,\,^7\). To our knowledge, the colonization for SA in pregnant women had not been evaluated previously in Colombia.

We found in this pilot study 3\% vaginal and 2\% nasal and vaginal colonization rates for SA, comparable to rates reported in similar studies in geographically different populations\(^9\,\,^1\,\,^2\,\,^3\,\,^9\,\,^1\,\,^3\), and a 29\% nasal colonization rate, which is in the range described in other similar studies\(^9\,\,^1\,\,^3\). For MRSA, nasal colonization rates in this group of pregnant women were much higher (8\%) than the rates reported for the general population\(^1\,\,^5\,\,^2\,\,^4\) and for pregnant women in different geographical areas\(^1\,\,^3\). For example, a pilot study carried out in Cleveland (USA) reported that 22\% of the study population were nasal carriers of SA\(^9\). The authors of that study referenced a former report published in 1978 where the nasal colonization of SA in pregnant women was 4\%. In a study similar to ours, Pinter *et al*\(^1\,\,^3\), reported that from 304 pregnant women, 34 (11.2\%) of them were colonized in the nares, seven of whom (2.3\%) were colonized with MRSA\(^1\,\,^3\).

Most recent data on SA colonization in obstetrics (both MSSA and MRSA) are derived mainly from the USA\(^5\,\,^9\,\,^1\,\,^1\,\,^3\) or Europe\(^1\,\,^2\), and until now they provide little evidence that either universal or targeted screening is beneficial for mothers or babies\(^5\,\,^1\,\,^1\,\,^3\,\,^2\,\,^4\,\,^2\,\,^5\). Higher colonization rates with SA are observed when the samples are from rectal-vaginal samples\(^1\,\,^1\,\,^2\,\,^1\,\,^1\) compared to vaginal\(^1\,\,^2\,\,^1\,\,^3\), which may reflect the fact that SA colonizes frequently the gastrointestinal tract. In the study published by Andrews *et al*\(^1\,\,^1\) genital tract colonization by MRSA among pregnant women was evaluated and correlated with infant outcome. From 5732 pregnant women participating in the study, 833 were colonized by SA (14.5\%), 202 of whom were MRSA positive (3.5\% overall MRSA colonization). The authors reported that no cases of early-onset invasive neonatal infection by MRSA occurred among infants in their study.

In the pilot study presented here, from 100 pregnant women screened, only five vaginal isolates were obtained, one of which was MRSA. Social demographic characteristics of our study population

<table>
<thead>
<tr>
<th>Site</th>
<th>SA neg</th>
<th>SA pos (n=36)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>MSSA(^1)</td>
<td>MRSA(^2)</td>
</tr>
<tr>
<td>Nasal</td>
<td>69</td>
<td>23</td>
</tr>
<tr>
<td>Vaginal</td>
<td>95</td>
<td>2</td>
</tr>
<tr>
<td>Both(^3)</td>
<td>98</td>
<td>2</td>
</tr>
</tbody>
</table>

1. Methicillin-resistant *S. aureus*
2. Methicillin-sensitive *S. aureus*
3. Both refers to cases in which *S. aureus* isolates were obtained from vagina and nares from the same patient
are summarized in Table 1. Statistical analysis of this data did not show significant associations with carrier state. This may reflect the fact that our sample population was small, although similar studies with larger populations have also failed to identify risk factors for SA colonization in pregnant women and the implications for maternal and neonatal outcomes. In our study, follow up could only be completed for 52 mother-infant pairs, and we did not observe SA-related complications in mothers or in newborns; thus, we could not identify factors associated to a higher risk of colonization or development of complications either in mothers or infants. To date, rectal vaginal colonization of SA has been associated to race, parity, route of birth, and colonization by *Streptococcus agalactiae*. In fact, Chen et al., reported that patients colonized by *S. agalactiae* were also more likely to be colonized with SA, but those colonized by MRSA were not colonized with *S. agalactiae*.

It is of interest to compare our current results for maternal colonization with SA and MRSA with those of Pinter et al., who reported a maternal colonization rate of 14.1% for SA and of 3% for MRSA, much lower than that reported in the current study. It should be noted, for comparison, that in the study by Pinter et al., all participants delivering their babies via vaginal route were screened on the day of delivery at both the nares and the vagina in contrast with the current study in which pregnant women were screened at both the nares and vagina between 35 and 42 weeks of gestational age, 17% of whom were in labor at the time of sampling. Whether differences in the colonization rates are related to differences in geographical areas or in the methodology used remains uncertain.

Although asymptomatic nasal, vaginal, and rectal colonization with MRSA has been reported to occur in some pregnancy-related clinical cases in which SA colonization has been identified as risk factor for serious systemic infection after delivery, epidemiological studies show that universal screening and decolonization efforts in pregnant women are currently non-cost effective. Similarly, there is no current evidence that vertical transmission of SA constitutes a significant risk for neonatal complications, although vertical transmission of SA have been documented in some studies.

In our pilot study, antibiotic susceptibility of the SA isolates found that all MRSA isolates had the resistance profile associated to the community, with exclusive resistance to methicillin. Also, three MSSA isolates had erythromycin resistance; one of them had the erythromycin inducible clindamycin resistance phenotype, detected by the D-test. We did not find resistance to other antibiotics in MRSA or MSSA isolates.

Table 3 summarizes the characteristics of MRSA isolates. SCC mec typing by multiplex PCR assay (Figure 1) showed that among the MRSA isolates from this study, two had SCC mec type I and seven had SCC mec type IV, data consistent with community-associated MRSA, given that it has been previously reported that MRSA isolates commonly found in community settings frequently bear SCC mec type IV (and V).

Although controversial, Panton-Valentine leukocidin has been widely regarded as a major virulence factor for MRSA. The presence of PVL1 in MRSA has been associated with more severe infections, including bacteraemia, osteomyelitis, and necrotising fasciitis. However, the role of PVL1 in MRSA infections remains unclear.

Table 3: Characteristics of methicillin-resistant *Staphylococcus aureus* isolates (n=9)

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Number of isolates</th>
</tr>
</thead>
<tbody>
<tr>
<td>Origin</td>
<td></td>
</tr>
<tr>
<td>Nasal</td>
<td>8</td>
</tr>
<tr>
<td>Vaginal</td>
<td>1</td>
</tr>
<tr>
<td>PVL1 carrying</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>7</td>
</tr>
<tr>
<td>Negative</td>
<td>2</td>
</tr>
<tr>
<td>SCCmec² types</td>
<td></td>
</tr>
<tr>
<td>Type I</td>
<td>2</td>
</tr>
<tr>
<td>Type IV</td>
<td>7</td>
</tr>
</tbody>
</table>

1 Panton-Valentine Leukocidin  
2 Staphylococcal chromosomal cassette mec.
determinant associated to epidemic clones of CA-MRSA in the world. In the current study, we determined the presence of PVL genes among SA isolates, both MSSA and MRSA. Among MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA isolates, six nasal isolates and the vaginal isolate were PVL positive (Figure 2). In contrast, among the MSSA isolates, only two were PVL positive; thus, PVL genes were more frequently found in MRSA
isolates compared to MSSA [7/9 (77.7%) vs 2/27 (7.4%); p<0.01].

Conclusions

Although this was a small pilot study and it may not be generalized to the entire pregnant population, the colonization rates found for MRSA were high, as were the frequency of PVL-positive MRSA isolates, emphasizing the need for further epidemiologic studies addressed to evaluate the impact of this colonization in puerperal and neonatal morbidity in our geographic region and to identify subpopulations of pregnant women that may benefit from SA screening, given that morbidity and associated costs for MRSA infections in this population are increasingly reported6,8.

We could not find statistical significance among socio demographic factors, colonization status, and development of complications, which could be attributed to the low number of participants able to complete the follow up. Thus, additional studies with more epidemiologic power are needed to ascertain the role of SA colonization in pregnant women from our region.

Antibiotic susceptibility profiles, SCCmec typing, and presence of PVL genes of MRSA isolates points out to their community origin.

Conflict of interest. The authors declare having no conflict of interest related to this study.

Acknowledgements

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