

Artículo de revisión

Bat Reservoirs for Rabies Virus and Epidemiology of Rabies in Colombia: a review*Murciélagos reservorios del virus de la Rabia y epidemiología de la Rabia en Colombia: una revisión**Reservatórios de morcegos do vírus da raiva e epidemiologia da raiva na Colômbia: uma revisão*Jimmy Fernando Cifuentes Jiménez¹, Est. MVZ; Rubén Darío Pérez Lopéz², MVZ; Noel Verjan García^{1*}, MVZ, MSc, PhD, [CvLAC](#)**Fecha correspondencia:**

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Comparte

**Abstract**

Rabies is a fatal zoonotic disease caused by a neurotropic RNA virus part of the family *Rhabdoviridae*. The disease is characterized by encephalitic inflammation and is responsible for a number of annual fatal deaths of people and animals in the world. Chiropters such as insectivorous, frugivorous and hematophagous bats are the major transmitters, reservoirs, and vectors of the rabies virus. In Colombia, a total of thirty-five cases of human rabies occurred between 2000 - 2014. Of the thirty-five cases twenty-two were transmitted by bats. The genetic variant V3 (hematophagous bats) were responsible for 24 human deaths, while the genetic variant V4, carried by insectivorous bats, caused three human deaths. The disease is mainly linked to infected cats that may have hunted infected bats and then transmitted the virus to humans. Diagnosis is usually made by detection of the virus from infected animals and techniques such as RT-PCR, which might be used to promote active surveillance in bat populations. Rabies is a preventable disease and vaccination of pets confers protective immunity, however, vaccination coverage of pets in Colombia is still limited. This review article collects epidemiological data of rabies virus genetic variants associated with chiropters species that have been reported to transmit the virus in the world and emphasizes on rabies cases reported in Colombia during the last decades.

Keywords: *Chiroptera, genetic variants, Rhabdoviridae, zoonotic diseases.***Resumen**

La rabia es una enfermedad zoonóticas mortal causada por un virus ARN neurotrópico de la familia *Rhabdoviridae*. La enfermedad se caracteriza por encefalitis y es responsable de múltiples muertes anuales de personas y animales. Quirópteros insectívoros, frugívoros y hematófagos son los principales transmisores, reservorios y vectores del virus. En Colombia, un total de 35 casos de rabia humana fueron reportados entre 2000- 2014, veintidós de ellos fueron transmitidos por murciélagos. La variante genética V3 (murciélagos hematófagos) fue responsable de 24 muertes humanas, mientras que la variante genética V4 (murciélagos insectívoros) causó tres muertes humanas. La enfermedad es transmitida al humano principalmente

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por gatos infectados que pudieron haber cazado murciélagos infectados. El diagnóstico se realiza mediante la detección del virus en animales rabiosos y técnicas como RT-PCR podrían utilizarse para promover la vigilancia activa de las poblaciones de murciélagos. La rabia es una enfermedad prevenible y la vacunación en animales domésticos confiere inmunidad protectora, sin embargo, la cobertura de vacunación en animales domésticos en Colombia es aún limitada. Esta revisión recoge datos epidemiológicos de las variantes genéticas del virus en especies de quirópteros reportadas como transmisores del virus en el mundo y enfatiza en los casos de rabia reportados en Colombia durante las últimas décadas.

Palabras clave: *Chiroptera*, enfermedad zoonótica, *Rhabdoviridae*, variantes genéticas.

Resumo

A raiva é uma enfermidade zoonótica fatal causada pelo vírus neurotrópico do ARN da família *Rhabdoviridae*. A enfermidade é conhecida por encefalite e é responsável das numerosas mortes anuais de animais e pessoas. Quirópteros e insetívoros, frugívoros e sanguessugas são os principais transmissores, reservatórios e vetores do vírus. Na Colômbia o Instituto Nacional de Saúde relatou um total de 35 casos de raiva humana entre 2000- 2014. Vinte dois deles foram transmitidos pelos morcegos. A variante genética V3 (morcegos hematófagos) foi responsável de 24 mortes humanas. Enquanto a variante genética V4 (morcegos insetívoros) provocou três mortes humanas. A enfermidade está ligada principalmente á os gatos infectados que poderiam ter caçado morcegos infectados e, em seguida, transmitida a os seres humanos. Diagnóstico e normalmente feito a través da detecção do vírus em animais rabiosos e técnicas tais como RT- PCR poderiam se utilizar para promover a vigilância ativa das populações de morcegos. A raiva é uma enfermidade evitável em animais domésticos cuja vacinação confere imunidade protetora, no entanto, a cobertura de vacinação em animais de estimação na Colômbia ainda é limitada, precisando de mais cobertura. Este artigo de revisão recolha dados epidemiológicos e variantes genéticas do vírus, e a diversidade de espécies de morcego reportado como transmissores do vírus no mundo. O manuscrito também resume os principais casos de raiva reportados na Colômbia nas últimas décadas e enfatiza a necessidade de reforçar a vigilância ativa para o diagnóstico de raiva em morcegos em todo o país.

Palavras-chave: *Chiroptera*, doença zoonótica, *Rhabdoviridae*, variantes genéticas.

Rabies virus

The rabies virus (RABV) is an RNA virus that belongs to the order *Mononegavirales*¹, genus *Lyssavirus*, and in the family *Rhabdoviridae*². The virus particles are bullet shaped and measure 75 nm in diameter with a length of 200 nm³. The genus *Lyssavirus* includes fourteen species which are classified by their genomic sequence; they are the RABV, Lagos bat virus (LBV), Mokola virus (MOKV), Duvenhage virus (DUVV), European bat lyssavirus type 1 (EBLV1), European bat lyssavirus type 2 (EBLV2), Australian bat lyssavirus (ABLV), Aravan virus (ARAV) and Khujand virus (KHUV), Irkut virus (IRKV), West Caucasian bat virus (WCBV), Bokeloh bat lyssavirus (BBLV), Ikoma Lyssavirus (IKOV) and Shimoni bat virus (SHIBV). Nine of the fourteen species of *lyssavirus* (DUVV, EBLV1, EBLV2, ABLV, ARAV, KHUV, IRKV, WCBV and BBLV) have been isolated from insectivorous bats. Additionally, the members of the genus *Lyssavirus* were subdivided into three phylogroups (I, II and III). Phylogroup I includes RABV, DUVV, EBLV1, EBLV2, ABLV, ARAV, KHUV, IRKV and BBLV^{2,4}. Phylogroup II includes LBV and MOKV and a new *lyssavirus* named "Shimoni bat virus" (SHIBV), which was isolated from a freshly dead

insectivorous bat (*Hipposideros commersoni*) in the coastal region of Kenya in 2009 and was found to belong to the Phylogroup II *lyssaviruses*⁵. Phylogroup III is compounded by the WCBV and one currently unclassified *lyssaviruses* identified as IKOV². The RABV genome consists of an approximately 12 kb nonsegmented, single negative strand RNA molecule⁶, encoding five structural proteins that are: the nucleocapsid (N) protein, a phosphoprotein (P), matrix (M) protein, glycoprotein (G), and an RNA-dependent RNA polymerase (L) in the order 3'-N-P-M-G-L-5'⁷. It is impossible to make a clinical differentiation between the disease caused by any virus species⁸.

Rabies, the disease

Rabies is one of the most important zoonotic diseases that is caused by a highly neurotropic *Rhabdovirus*⁹. The disease is reported in domestic and wild animals worldwide and it's estimated to cause up to 70,000 human deaths per year, mostly in rural areas of Asia and Africa¹⁰. Rabies represents a neuroinvasive disease that is characterized by acute encephalitis with two clinical manifestations; the furious (classical or encephalitic) and the paralytic form. Furious rabies is the most common form of human rabies, accounting for approximately 80 % of cases. With the exception of Antarctica, rabies is endemic to all continents. Classical RABVs have a worldwide distribution except for a few island nations such as Great Britain, Ireland, New Zealand, Hawaii, the continents of Australia and Antarctica, and an increasing number of Western European countries¹¹. The disease affects a broad spectrum of warm blooded animals. All mammals are susceptible to varying degrees, especially members of the order *Carnivora* and *Chiroptera*¹⁰.

Rabies is an invariable fatal disease, particularly when clinical symptoms have developed. Rabid animals commonly show neurological changes such as paraparesis, aggressiveness, hydrophobia and sialorrhea, agitation, mental confusion, and tetraparesis¹². In Latin America, rabies is classified into two epidemiologic forms; urban rabies and sylvatic rabies. The common vampire bat, *Desmodus rotundus*, has emerged as the principal RABV reservoir host along the species natural range from Mexico to South America. *D. rotundus* was correlated with the vampire bat RABV variants which is more prevalent than the dog RABV variants in Peru, however, the prevalence of any RABV variant might be dependent of local or regional RABV vaccination plans⁹.

Pathogenesis of rabies

Paralytic rabies is characterized by flaccid paralysis in the bitten limb, which ascends symmetrically or asymmetrically, whereas the furious rabies manifests hyper-excitability, autonomic dysfunction, hydrophobia, and aerophobia^{13,14}.

RABV may enter the organism by different transmission routes such as animal bites or scratches, and the virus can remain latent close to the inoculation site for long periods of time. The virus replicates slowly within muscle cells until it arrives to the neuromuscular junction¹⁴. The viral glycoprotein is essential for transsynaptic spreading by using cell nicotinic acetylcholine receptors at the neuromuscular junctions¹⁵. The virus migrates along peripheral nerves to the central nervous system via retrograde fast axonal transport, a process that is facilitated by the viral P protein¹⁷, at a rate of 12 to 100 mm/d¹⁶. In the CNS, RABV replicates in neurons and induces necrosis and inflammation¹⁸. Then, the virus spread to other organs of the body, reaching the salivary gland where high viral concentrations of virus could be found in the saliva, peripheral nerves and other organs² (Figure 1).

RABV might be present in different tissues of bats: particularly the buccal cavity, saliva, and brown fat. The distribution of RABV in tissues was evaluated in 26 bat species from Brazil. The species *Artibeus lituratus* (13), *Myotis nigricans* (4), *Eptesicus furinalis* (5), *Eptesicus diminutus* (1), *Lasiurus blossevillii* (1), and *Lasiurus ega* (2) were tested by using hnRT-PCR. The virus was detected in tongue (92 % and 85 %), brown fat (82 % and 77 %), lung (62 % and 77 %), heart (42 % and 77%), stomach (92 % and 64 %), liver (38 % and 67 %), spleen (43 % and 27 %), bladder (73 % and 88 %), kidney (77 % and 38 %), intestine tissues (77 % and 38 %), and feces (38 % and 42 %) from frugivorous and insectivorous bats. It was noted that the virus was higher in stomachs of frugivorous bats than from insectivorous bats¹⁹. A possible reason for those findings could be that infected bats defecate on the fruits that are eaten later by frugivorous bats, which ingest the virus. Contrary, insectivorous bats, due to their eating habits, may have lower viral loads. However, additional studies are necessary to determine the relationship between feeding habits of the non-hematophagous bats and the rabies virus.

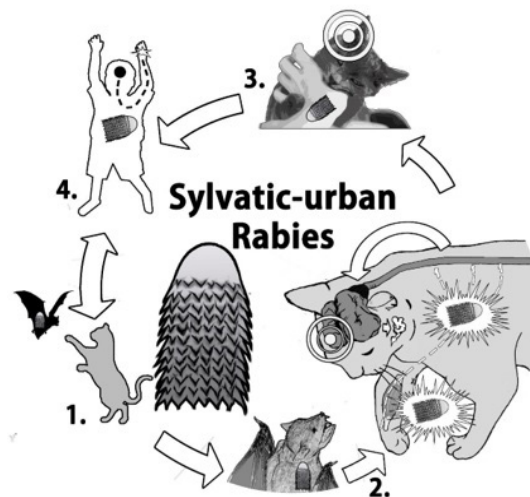


Figure 1. Connection of sylvatic rabies to urban ecosystem. 1. The cat hunts an infected chiropter. 2. The cat is bitten, the virus enters the body through the wound by infiltration of saliva, and the virus replicates in the peripheral muscles close to the wound and migrates to CNS. In the CNS, the virus is distributed to other organs, eventually reaching the salivary glands. 3. The rabid animal shows neurological changes. 4. The animal is predisposed to attack humans or other animals. The cycle repeats and causes the death of the infected individual.

Rabies virus transmission

The RABV is usually transmitted from an infected animal to one that's susceptible¹. The virus is usually present in the saliva of rabid animals, and it enters the body via infiltration of virus-laden saliva into a wound or by the exposure of mucosal surfaces to saliva from an infected animal (bites)²⁰. Additionally, a rare form of transmission may be by the inhalation of aerosols with RABV present. This would most likely occur in caves with dense populations of bats in which the virus is present²¹.

The RABV and the majority of lyssaviruses, are found in natural bat reservoirs. These animals are unique among mammals due to having exceptional sociality and longevity. Given these features and the recognized status of bats as reservoirs for RABVs in the Americas, individual bats may experience repeated exposure to RABV during their lifetime²². Hematophagous, frugivorous, and insectivorous bats can transmit

the RABV²³, and phylogenetic analysis using nucleotide sequences of N or G genes revealed that RABV is grouped into clusters according to the bat species that support the existence of species-specific variants or lineages of the virus ^{19, 24}.

Main species of Bats reservoirs for rabies virus in the world

The class *Mammalia* has 5,416 species and the order *Chiroptera* comprises of the second largest group of mammals in number, with 1,120 species ²³. Bats have a worldwide distribution, absent only in the Polar Regions and some oceanic islands. Most of these animals live in tropical and subtropical regions, but can be found in temperate regions ²⁵. A number of bats reservoirs for RABV has been described, including hematophagous, insectivorous and frugivorous bats (Table 1), although the RABV has been detected mainly in insectivorous bat species (Figure 2).

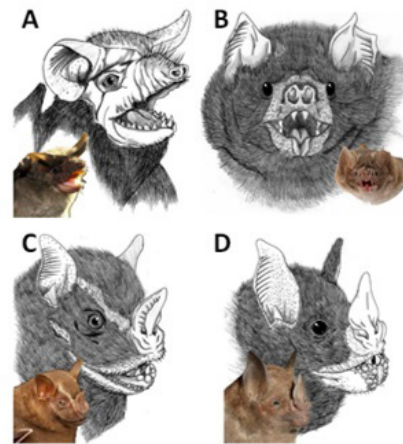


Figure 2. Main bat reservoirs of RABV are represented by a photograph and a schematic diagram. A. *Tadarida brasiliensis* (insectivoro), B. *Desmodus rotundus* (hematophagous), C. *Artibeus lituratus* (frugivorous) and D. *Carollia perspicillata* (frugivorous).

Insectivorous bat species have worldwide distribution and can be found in almost all ecosystems, together with others frugivorous bats species, whereas all the three species of hematophagous bats are only found in Latin America, with the common vampire (*Desmodus rotundus*) being the only well-known RABV reservoir ²⁶.

Table.1 *Chiroptera* species that have been reported positive for the presence of RABV in the world.

N	Species	Location	Ecology	Reference
1	<i>Desmodus rotundus</i>	Brazil/Colombia	hematophagous	12, 27
2	<i>Diphylla ecaudata</i>	Brazil	hematophagous	8, 25
3	<i>Diaemus youngi</i>	Brazil	hematophagous	25
4	<i>Tadarida brasiliensis</i>	Chile	insectivorous	8, 28
5	<i>Lasiurus cinereus</i>	Chile	insectivorous	28
6	<i>Histiotus macrotus</i>	Chile	insectivorous	28
7	<i>Myotis chiloensis</i>	Chile	insectivorous	28
8	<i>Artibeus lituratus</i>	Brazil	frugivorous	24
9	<i>Chrotopterus auritus</i>	Brazil	omnivorous	25
10	<i>Eptesicus fuscus</i>	United state	insectivorous	22
11	<i>Myotis sp</i>	United state	insectivorous	29

N	Species	Location	Ecology	Reference
12	<i>Lasionycteris noctivagans</i>	United states migratory tree-roosting hoary	insectivorous	30
13	<i>Lasiurus cinereus</i>	United states migratory tree-roosting hoary NA	insectivorous	30
14	<i>Eptesicus spp</i>	Brazil	insectivorous	31
15	<i>Nyctinomops macrotis</i>	Brazil	insectivorous	25
16	<i>Molossus spp</i>	Brazil	insectivorous	31
17	<i>Tadarida spp</i>	Brazil	insectivorous	31
18	<i>Histiotus velatus</i>	Brazil	insectivorous	25
19	<i>Lasiurus spp</i>	Brazil	insectivorous	31
20	<i>Molossus ater</i>	Brazil	insectivorous	32
21	<i>Molossus molossus</i>	Brazil	insectivorous	33
22	<i>Eumops auripendulus</i>	Brazil- Ecuador	insectivorous	33
23	<i>Nyctinomops laticaudatus</i>	Brazil- Ecuador	insectivorous	33
24	<i>Eptesicus furinalis</i>	Brazil- Ecuador	insectivorous	33
25	<i>Lasiurus cinereus</i>	United states	insectivorous	34
26	<i>Lasiurus borealis</i>	United states/ Chile	insectivorous	28, 34
27	<i>Lasionycteris noctivagans</i>	United states	insectivorous	35
28	<i>Dasypterus floridanus</i>	Florida/ solitary	insectivorous	35
29	<i>Phyllostomus superciliatum</i>	United states	insectivorous	36
30	<i>Myotis evo</i>	United states	insectivorous	35
31	<i>Antrozous pallidus</i>	Canada, México and Cuba	insectivorous	35
32	<i>Macrotus californicus</i>	Mexico and the United States	insectivorous	35
33	<i>Pipistrellus hesperus</i>	western United States and Mexico	insectivorous	35
34	<i>Myotis keenii</i>	Canada	insectivorous	37
35	<i>Euderma maculata</i>	Canada	insectivorous	37
36	<i>Myotis lucifugus</i>	Canada	insectivorous	37
37	<i>Myotis yumanensis</i>	Canada	insectivorous	37
38	<i>Lasiurus intermedius</i>	Canada	insectivorous	37
39	<i>Myotis evotis</i>	Canada	insectivorous	37
40	<i>Myotis nattereri</i>	German and France	insectivorous	38
41	<i>Myotis dasycneme</i>	Germany	insectivorous	38
42	<i>Myotis daubentonii</i>	Germany	insectivorous	38
43	<i>Eptesicus isabellinus</i>	Germany	insectivorous	38
44	<i>Eptesicus serotinus</i>	Germany- Spain	insectivorous	38, 39
45	<i>Pipistrellus nathusii</i>	Germany	insectivorous	38
46	<i>Miniopterus schreibersii</i>	Southeastern Europe	insectivorous	40
47	<i>Hipposideros commersoni</i>	Kenya	insectivorous	5
48	<i>Rousettus aegyptiacus</i>	Europe Mediterranean	frugivorous	5
49	<i>Pipistrellus pipistrellus</i>	Spain EBLV1, EBLV2	insectivorous	39
50	<i>Artibeus jamaicensis</i>	Kenia	frugivorous	8
51	<i>Eidolon helvum</i>	Nigeria	frugivorous	41
52	<i>Rousettus aegyptiacus</i>	Kenia	frugivorous	41
53	<i>Micropteropus pusillus</i>	Central African Republic	frugivorous	41
54	<i>Epomophorus wahlbergi</i>	South Africa	frugivorous	41
55	<i>Nycteris gambiensis</i>	Guinea	insectivorous	42
56	<i>Artibeus obscurus</i>	Brazil	frugivorous	25
57	<i>Phyllostomus hastatus hastatus</i>	Brazil	insectivorous	25, 43
58	<i>Uroderma bilobatum</i>	Colombia	insectivorous	44
59	<i>Phyllostomus hastatus</i>	Colombia	Frugivoro-insectivorous ⁴⁵	44
60	<i>Eptesicus braziliensis</i>	Colombia	insectivorous	46
61	<i>Carollia perspicillata</i>	Colombia	frugivorous	47
62	<i>Myotis nigricans</i>	Colombia	insectivorous	47
63	<i>Lasiurus ega</i>	Colombia	insectivorous	47
64	<i>Molossus molossus</i>	Colombia	insectivorous	48

Rabies cases and virus variants in Colombia

RABV in Colombia have been grouped into three variants, Colombian genetic variant I viruses (isolated in Arauca and the Central Region of the country), Colombian genetic variant II viruses (isolated in the Caribbean Region) and the third group that consists of viruses isolated from two insectivorous bats (*Eptesicus brasiliensis* and *Molossus molossus*), three domestic dogs and a human. The genetic sequence analysis indicated that the virus isolates belonging to the third group were variants of bat RABV, the first finding that associated bats to rabies in dogs and humans in Colombia ⁴⁹. Rabies in Colombia and other Latin American countries are an important public health and economic problem, and the disease is categorized as urban rabies and sylvatic rabies, which have distinct epidemiological cycles ^{50,51}. Urban rabies is usually transmitted by the domestic dog and this animal is the main transmitter and reservoir of the virus, whereas sylvatic rabies is transmitted principally by bats the main reservoirs of the disease that is transmitted to domestic animals such as cows and horses, however, mongooses and coyotes can share the Variant 1 (urban rabies) with domestic dogs in less proportion ^{49,56}.

Wild RABV variants identified in Colombia are V3 (hematophagous bats), V4 (insectivorous bats), V5 (hematophagous bats) and V8 (skunk) ⁵². A study that analyzed a total of 124 samples obtained from human cases of rabies and 8 from other mammal species within the period 1994–2005, identified eight genetic lineages (GL1- GL8), of RABV. Phylogenetic analyses of the partial nucleoprotein gene sequence determined specific variants within those genotypes. The GL4 comprised of Variant V3 and V8 and a variant not determined (ND), which were associated with hematophagous bats, the GL5 and GL6 consisted of V4 viruses associated with *Tadarida brasiliensis* bats, the GL5 grouped independently. The GL7 and GL8 segregated independently within clades associated with colonial insectivorous and solitary bats, both of these were not determined variants. RABVs isolated from humans grouped within GL2, GL3 and GL4, which corresponded to V1, V3, V8 and ND. Dogs and *Desmodus rotundus* are the two major RABV reservoirs and vectors in Colombia, although insectivorous bats may also be involved ⁵³.

RABV variants V3 and V4 are the most prevalent in Colombia, and this situation seems to be influenced by increased deforestation and urban architecture that provides shelter, causing more frequent interactions between humans and bats ⁴⁴. The variant V4, that is associated with frugivorous and insectivorous bats, was isolated from a dog and a human in the northern of Colombia ⁵⁴.

RABV was isolated for the first time from bats in Colombia by Alarcon in 1968, who reported the isolation of two strains of RABV from insectivorous and frugivorous bats (*Myotis nigricans*, *Lasiurus ega* and *Carollia perspicillata*) captured from areas of the departments of Guajira, Santander and Antioquia. Other bat species such as *Glossophaga longirostris*, *Artibeus lituratus palmarum*, *Platyrrhinus Helleri* (*Vampyrops Helleri*), *Trachops Cirrhosus cirrhosus*, *Peropteryx kappleri*, *Phyllostomus Hastatus*, *Saccopteryx bilineata*, and *Molossus molossus* were negative to the virus ⁴⁷. In the urban zone of Cali, the species *Carollia perspicillata*, *Artibeus lituratus*, *Eptesicus brasiliensis*, *Myotis nigricans*, *Molossus molossus*, and *Tadarida brasiliensis* were also reported as transmitters of RABV in the period December 2000 to June 2002 ⁵⁵.

The National Institute of Health of Colombia has reported a considerable number of cases of sylvatic rabies transmitted by bats since 2000. A total of thirty-five cases of human rabies, twenty-two of those cases were transmitted by bats, eight by cats,

and five by dogs. Regarding the virus variants, two were variant VA (atypical), five-Variant V1 (domestic and wild dogs, mongooses and coyotes), twenty-four-variant V3 (hematophagous bat), three-Variant V4 (insectivorous bat), and one of those cases was variant V8 (skunk) ⁵⁶. Outbreaks of rabies disease have been recorded in high magnitude in Bajo Baudo, Choco, where hematophagous bats represent the major threat for sylvatic rabies transmission ^{51, 57}. Outbreaks of rabies caused by variants V3 have also been recorded in San Luis de Palenque in Casanare and Floridablanca in Santander ⁵⁶, and Santander de Quilichao in Cauca, where a cat was the transmitter to humans, linking sylvatic rabies and the urban ecosystem ²⁷. In Encino and Piedecuesta in Santander, other two human rabies cases were reported to be caused by the variant V3 transmitted by cat and a hematophagous bat, respectively. Cats usually get infected by contact with bats during their predatory behavior ⁵⁰.

Outbreaks of rabies caused by variant V4 were reported in Moniquirá, Boyacá, where the insectivorous bat *Tadarida brasiliensis* was identified as the main reservoir ⁴⁴. The variant V4 was also responsible for an outbreak in Roldanillo, Valle del Cauca in 2012 ⁵⁶, whereas in Barrancabermeja, Santander an outbreak was caused by the atypical variant VA ⁵⁶. An atypical RABV variant from sylvatic origin was also responsible for the disease in a child that had contact with a cat in San Luis, Tolima in 2010 (INS, 2014).

Diagnosis of rabies infection

The diagnosis of the virus is conducted by the use of a number of methodologies that include the detection of rabies antigens in tissues, nucleic acids, amplification of rabies particles or serological tests.

Mouse inoculation test

It consist of an intracerebral inoculation of a clarified supernatant of a 10-20 % (w/v) homogenate of brain material including brainstem (cortex, Ammon's horn, thalamus, medulla oblongata) in an isotonic buffered solution with antibiotics, into groups of 3 to 10 mice that should be observed by 28 days to record mortality and detection of RABV. In the case of newborn mouse, they can be evaluated on days 5, 7, 9 and 11 post-inoculation. Any deaths occurring during the first 4 days are regarded as nonspecific (due to stress or bacterial infection) ⁵⁹.

Fluorescent antibody test (FAT)

It is the most widely used test for rabies diagnosis recommended by WHO and OIE as the gold standard test and it may be used directly on a smear. The test uses purified immunoglobulin previously conjugated with fluorescein isothiocyanate (FITC) that is added onto an acetone-fixed brain tissue smear, preferably made from several parts of the central nervous system. FAT provides a reliable diagnosis in 98–100 % of cases for all RABV strains if a potent conjugate is used. FAT can be applied in fresh or frozen brain tissues sections, with very similar results (99.8% sensitivity and 100 % specificity) when applied to fresh or formalin-fixed tissues ⁶¹, however, it should not be used in decomposed tissue samples ⁶⁰.

Reverse transcription PCR (RT-PCR)

Classical reverse transcription-polymerase chain reaction assay has been reported to be a sensitive and specific tool for routine diagnostic purposes ⁶². It consist of the use of a reverse transcriptase to synthesize a complementary DNA copy from viral RNA and then, conventional PCR is used to amplify a gene fragment from the virus genome ⁶³. The technique can be used to detect RABV in decomposed samples that

often appear due to the warm climate and fluctuations in ambient temperature during sample transport and storage ⁶⁴. Positive diagnostic results from such samples are reliable but negative results may be invalid ⁵⁸.

A comparison of RT-PCR and MIT for detection of RABV in 95 positive samples that were stored for 4–13 years at -20 and -80 °C revealed that only 32 (33,6 %) of the samples were positive with the mouse inoculation test, while RT-PCR detected the viral genome in 62 (65,3 %) samples. Samples that were stored for >10 years gave 59.7 % positivity by RT-PCR and only 22.1 % by MIT ⁶⁵.

Heminested RT-PCR (HnRT-PCR)

It is one of the most sensitive and rapid technique for rabies diagnosis. The method can be applied to both living animals and post mortem collected samples, when the brain samples are in a decomposed state. The PCR products can be used for DNA sequencing for final identification of virus origin by epidemiological analysis ⁶⁶. Our group implemented this technique to analyze brain tissues from a number of bats collected in rural areas of the Tolima region (Figure 2). A total of eleven bats species including *Artibeus jamaicensis*, *Artibeus lituratus*, *Carollia perspicillata*, *Desmodus rotundus*, *Molossus molossus*, *Molossus ater*, *Myotis nigricans*, *Phyllostomus hastatus*, *Platyrrhinus dorsalis*, *Saccopteryx bilineata* and *Saccopteryx leptura* were analyzed by HnRT-PCR and all samples were negative for the presence of RABV (Figure 2).

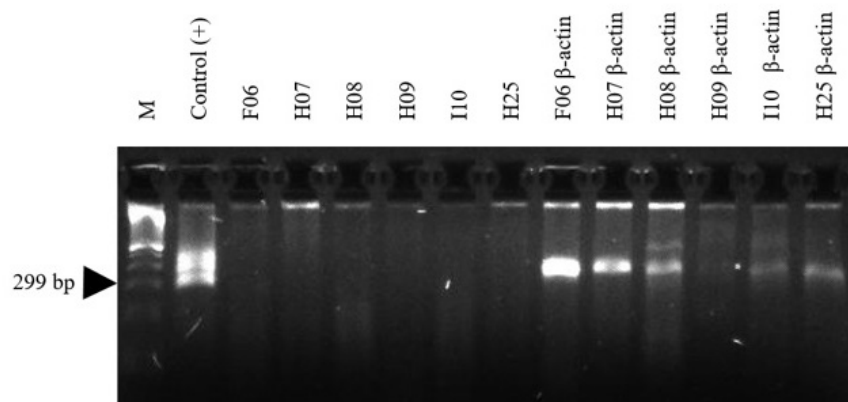


Figure 3. HnRT-PCR amplification of 299 bp of RABV N gene from chiropters brain tissues. Total RNA was extracted from brain tissue of six captured bats in the Tolima region and subjected to RT-PCR analysis and the cDNA construction was validated with the amplification of the housekeeping gene β -actin in each sample. M: 100 bp DNA ladder; Positive control RNA from RABV was kindly provided by Dr. Andres Páez from the National Institute of Health of Colombia (INS). F06, H07, H08, H09, I10 and H25 corresponds to brain bats samples.

Real time PCR

It is an alternative test that have been demonstrated to be more sensitive than HnRT-PCR and can be used to further extend the rabies RNA detection limits in decomposed samples ⁶⁷. This technique allows to assess gene expression analysis, determination of viral load and detection of genetically modified organisms ⁶⁸. The primers are designed in such a way that a fluorescent signal is generated only when the primers are incorporated into an amplification product. Detection of target sequences occurs by monitoring the fluorescence generated by intercalating dyes or fluorophore labelled primers for sequence-specific probes ⁶⁹. This assay has high sensitivity and specificity enabling simultaneous amplification and quantification of

specific nucleic acid sequences that made it exceptional for diagnosis for this infectious agent ^{69,70}. Real time PCR was compared to conventional RT-PCR to analyze saliva samples from 21 suspected patients and found that the sensitivity was superior (75% vs. 37%) to that offered by RT-PCR ⁷¹.

Enzyme linked immunosorbent assay (ELISA)

The ELISA is the most used diagnostic test for rabies, which measures specific immunoglobulins such as IgM. The test consist in the detection of RABV neutralizing antibody in sera samples taken from the suspected animal ⁷². The test reduce time, facilitate handling and avoid the use of biosecurity level 2 or 3 laboratories, do not require live RABV or cell culture and can be automated. The test was developed for domestic carnivores and wildlife and is the only one certified and prescribed by the OIE for rabies detection. However, the ELISA test, although it may have 100 % specificity, the sensitivity was around 78.2 % when 593 samples of domestic carnivores were evaluated ^{73,74}.

Prevention of rabies

Rabies is considered a vaccine-preventable disease and annual vaccination of pets is the recommended strategy to prevent and control rabies ⁷⁶. However, it has been reported that epizootic areas of the rabies virus are self-limiting and *D. rotundus* sacrificial campaigns have minimal or no incidence on the rate of presentation of the disease in these areas ⁷⁷. In the case of an attack by a potentially rabid animal, it must be immediately informed to the competent health authorities to start an appropriate research (study of focus, rabies vaccination of dogs and cats, watching suspected animals, taking and sending samples, and institutional active search) and specific treatment which is contemplated by the INS ⁵². It is recommended to avoid wild animals as pets and not to handle animals suspected of carrying the disease (dogs, cats, cattle) or handle bats that are on the ground or showing abnormal behavior ⁷⁵. Competent authorities in each country must have an active surveillance system for the disease and appropriate vaccination programs in areas with high risk of presentation ¹⁰. Unfortunately, in Colombia the coverage of vaccination of pets is very limited, the percentage of vaccination coverage of municipalities in Colombia is limited only 59,9 % of municipalities have a % coverage ≥ 80 %, 7,96 % of municipalities a % coverage between 70 and 79 %, 6,64 % of the municipalities a % coverage between 60-69 % and 25,5 % of the remaining municipalities a % coverage ≤ 59 % ⁷⁸.

Conclusions

Rabies is a zoonotic disease caused by a neurotropic virus that is present in many species of chiropters all over the world. RABV induces an acute encephalitis, clinically manifested as furious and a paralytic form. The virus is classified in Latin countries as urban rabies and sylvatic rabies that are transmitted by domestic animals and bats respectively (Figure 1). Insectivorous and frugivorous bats act predominantly as reservoirs of rabies and transmitters of the disease in many part of the world. In Colombia, variants V3,V4 and VA (variant hematophagous bat, variant insectivorous bat and variant atypical associated with bats), of the virus, are responsible for a significant number of rabies outbreaks in human ⁵². The disease is mainly linked to felines such as cats that may hunt infected bats, getting infected too, which make a rabid cat that usually contact with humans, transmitting the virus. Thus evaluation of rapid diagnosis techniques such as RT-PCR to detect RABV in bat tissues might be needed to promote active surveillance to bats populations. Rabies is prevented by annual vaccination of pets, however, the coverage of this activity in Colombia is still limited, thus much effort is needed to educate and sensitize the people on this fatal disease.

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Conflicts of interest

The authors declare they have no conflicts of interest with regard to the work presented in this report.

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