Relationship between chlorophyll a fluorescence and yield in banana (*Musa* AAA Simmonds cv. Cavendish)

Relación entre la fluorescencia de la clorofila a y el rendimiento en el banano (*Musa* AAA Simmonds cv. Cavendish)



ABSTRACT

Chlorophyll a fluorescence is used to characterize plant responses to natural environments or stress conditions. This study aimed to identify correlations between the maximum quantum efficiency of PSII (Qy) with climatic and soil variables associated with bunch weight. The experiment design used in the field included an observational and longitudinal study, where 72 production units in the independent vegetative stage were randomly selected, of which 36 units were from a low production area, and 36 were from a high production area. In each unit, the quantum yield (Qy: Fv/Fm) was measured and correlated with climatic and soil variables, the nutrient content in the leaves, and the average weight of the cluster in each sampled batch. The results indicated a correlation between a Qy of -0.703 for temperature and -0.583 for radiation. On the other hand, pH and resistance to soil penetration were directly related to the Qy to a lesser extent, with r^2 values of 12.77 and 24.28%, respectively. Rainfall, wind, gusts, dew point, atmospheric pressure, and texture composition of the soil did not significantly affect Qy variation; when attacks by leaf pests (mites) occurred, significant reductions were also observed. Qy is a variable that responds to the stress generated by factors external to a plant that can significantly affect production, with a reduction of up to 2 kg per cluster and a bias of three to four weeks when a loss of 0.04 points occurs in the Qy value.

Additional keywords: abiotic stress; photosystem II; plant phenology; plant response; Musaceae.

³ Corresponding author. dfferiag@unal.edu.co



¹ Universidad Nacional de Colombia, Sede Medellín, Facultad de Ciencias Agrarias, Medellin (Colombia). ORCID Feria-Gómez, D.F.: 0000-0003-1470-1388; ORCID Córdoba-Gaona, O.J.: 0000-0003-2642-4146

² Asociación de Bananeros de Colombia (AUGURA), Centro de Investigaciones del Banano (Cenibanano), Carepa (Colombia). ORCID Londoño-Puerta, D.A.: 0000-0002-8359-8815



RESUMEN

La fluorescencia de la clorofila a es empleada para caracterizar la respuesta de los vegetales a ambientes naturales o bajo condiciones de estrés. El objetivo de este estudio fue identificar correlaciones entre la máxima eficiencia cuántica de PSII (Qy) con variables climáticas y edáficas asociadas al peso del racimo. Se empleo un diseño experimental de tipo observacional y longitudinal, donde se seleccionaron al azar 72 unidades productivas en etapa vegetativa independiente, 36 en un lote de baja producción y 36 en un lote de alta producción, a cada una de las cuales se le midió el rendimiento cuántico (Qy: Fv/Fm) del fotosistema II, y se correlacionó con variables climáticas y edáficas, con el contenido de nutrientes en las hojas y con el peso promedio del racimo en cada uno de los lotes muestreados. Los resultados indicaron una correlación entre Qy de -0,703 con respecto a la temperatura y -0,583 con la radiación. Por otro lado, el pH y la resistencia a la penetración del suelo se relacionaron de forma directa con Qy en menor proporción con valores de r^2 de 12,77 y 24,28%, respectivamente. La precipitación, el viento y sus rachas, el punto de rocío, la presión atmosférica y la composición textural del suelo no afectaron de forma significativa la variación Qy. Cuando se produjeron ataques de plagas en hojas (ácaros), también se observaron reducciones significativas en Qy. Qy es una variable que responde al estrés generado por factores externos a la planta, estos pueden afectar significativamente la producción, la cual se reduce hasta en 2 kg por racimo, con un sesgo de tres a cuatro semanas, al presentarse la pérdida de 0,04 puntos en el valor de Qy.

Palabras clave adicionales: estrés abiótico, fotosistema II; fenología vegetal; respuesta de la planta; Musaceae.

Received: 18-08-2021 Accepted: 26-10-2021 Published: 23-02-2022

INTRODUCTION

Banana (Musa AAA Simmonds) is one of the more consumed fruits worldwide because of its nutritional contribution. In 2019, this species was planted in 5,158,582 ha, with a production of 116,781,658 t, where countries such as India (26.0%), China (10.2%), Indonesia (6.2%), Brazil (5.8%), and Ecuador (5.6%) are the main producers. Colombia ranks tenth in production with 2,914,419 (2.5%) in 105,609 ha (FAO, 2021). Bananas are of economic and social importance, being the third agricultural export product after flowers and coffee (ICA, 2019). In 2020, Colombia produced a total of 109 million boxes for export from 51,454 ha. The Urabá region of Antioquia has the largest established area with 36,582 ha and a production of 1,299,526 t (AUGURA, 2021). One of the greatest challenges for banana production in this region is implementing agronomic management strategies to reduce the negative effects of climate variability on crop yield (Barrera et al., 2020). As part of this strategy, determining the physiological responses of plants and how they are affected by environmental aspects such as climate, nutrition, and agronomic management is of great importance to recognize the main factors that affect production (Maxwell and Johnson, 2000; Sánchez-Reinoso et al., 2019). One of the more regularly affected components is photosystem II (PSII) because of deficiencies in nutrition, changes in climatic variables, soil conditions, and damage generated by crop management. This leads to a reduction in light absorption and increases the dissipation mechanisms of excess energy by fluorescence (Maxwell and Johnson, 2000).

The description of the photosynthetic process in terms of photochemistry and its dissipation in heat and other forms has been of great value in characterizing the responses of plants. The chlorophyll fluorescence protocol with functional parameters has made it possible to understand the responses of plants to the associated external factors that affect photochemistry and the relationship between assimilated light energy with a reduction in the assimilation of carbon potential crop yield (Adak, 2018). Chlorophyll fluorescence has been used to understand the physiological mechanisms of cold tolerance in banana plants (Zhang et al., 2012); nutritional deficiencies such as iron (Catalina, 2015), calcium (Sanz et al., 2001), and nitrogen (Pérez et al., 2017) have been used to identify mechanisms of resistance to drought stress in bananas (Ravi et al., 2013) and for monitoring and early detection to reduce the spread of diseases in crops (Kim et al., 2019). However, little is known about the participation of chlorophyll fluorescence parameters and their relationship with the productive potential

of bananas in terms of climatic and edaphic supply. Therefore, this study aimed to identify correlations between the maximum quantum efficiency of PSII (Qy = Fv/Fm) with climatic and edaphic variables associated with fruit yield, helping to determine an indicator that explains variations in banana fruit yield.

MATERIALS AND METHODS

Location

The study was conducted in the Ramiro Jaramillo Sossa – AUGURA-CENIBANANO Experimental Field (CE) located in the municipality of Carepa, Antioquia (Colombia) (7°46'48.2" N; 76°40'23.0" W) in 2007, a farm dedicated to the conventional production of bananas for export. The experimental field is at 20 m a.s.l., with an average temperature of 27.0°C, solar brightness of 1,800 h year¹, relative humidity of 87%, and rainfall of 2,933 mm with 28% distributed in the first semester (cycle) and 62% in the second semester of production (Pérez and Torres-Bazurto, 2020). An AgroMET meteorological station (RainWise® Inc.) and 15 min intervals were used to determine the climatic variables: radiation (W m⁻²), temperature (°C), relative humidity (%), dew point (°C), atmospheric pressure (mb), rainfall (mm), wind speed (m s⁻¹) and vapor pressure deficit (VPD).

Experiment design

This work was an observational study. Two experiment areas (plots) of $1,563 \text{ m}^2$ were selected, with two productive levels. The first plot (P12) was planted in 2007, with an average weight of 27.13 kg per bunch, and the second (P6) plot was sown in 2005, with an average bunch weight of 22.91 kg. Each experiment plot (Fig. 1) had 36 productive units (mother-daughter) of bananas that were randomly selected, with a single successional sucker in an F10 + 2 physiological stage (leaf three emitted entirely). The plant material was from the *Musa* AAA group, Cavendish subgroup, Gran Nain clone, sown at a density of 1,780 plants/ ha in plot P6 and 1,685 plants/ha in plot P12.

Chlorophyll a fluorescence

The maximum fluorescence (Fm), basal (Fo), variable (Fv), and maximum quantum efficiency of the PSII (Qy = Fv/Fm) were estimated in the middle third of

the leaf in the sucker in each production unit. The chlorophyll fluorometer OS30p+ (Opti-Sciences) was used on dark-adapted leaves (30 min) with a modulation light intensity at 70% and saturation light intensity at 3,600 μ mols at 60% with the width of 1.0 s. This was repeated until leaf F10 + 10 (independent vegetative stage), always ensuring the leaf sampling was in position three from week 47 of 2020 to week 8 of 2021.

Soil analysis

A soil sample was taken at 0.3 m from the pseudostem in each production unit. The soil texture was determined using the Bouyoucos (1936) method (hydrometer), while the soil pH was estimated using a SevenCompact S220 pH meter with Inlab sensor[®] Expert Pro - ISM (Mettler - Toledo) in 1:1 solution (20 g of soil: 20 mL of distilled water). Additionally, soil penetration resistance was determined in the first 0.8 m depth at each site, using a 6.08 Penetrometer (Eijkelkamp) with a Thetaprobe ML3 soil moisture sensor.

Foliar nutrient content

In the sucker of each productive unit, samples of leaf tissue between 10 and 15 cm wide were taken (perpendicular to the central rib) on leaf number three (3). The samples were washed with distilled water and oven-dried at 45°C for 2 weeks, ground and sieved to 2 mm, and analyzed in the Soil Health Laboratory BioFertilizar S.A.S. to determine the content of foliar nutrients: nitrogen (Kjeldhal method), phosphorus (Phosphomolybdate complex colorimetric method), sulfur (Bactogelatin BaCl₂ turbidimetry method), boron (Azomethin h colorimetric method) and the nutrients calcium, magnesium, potassium, iron, manganese, copper, and zinc (atomic absorption method).

Bunch weight

The average bunch (11 weeks) weight of the plants at harvest in each plot was recorded weekly using an XR3000 (Tru-Test[®]) electronic scale.

Statistical analysis

The experiment design used in the field was an observational and longitudinal study without intervening





in their natural course over time (Manterola and Zavando, 2009). Assumptions of normality and homoscedasticity for Qy (Fv/Fm) were carried out with Kolmogorov-Smirnov and Bartlett tests (P < 0.05), respectively. Since the data did not meet these assumptions, the comparison of significant differences for Qy was done using the Kruskal-Wallis rank-sum test (P < 0.05). Once the comparisons were made between the leaves emitted from the F10, a correlation of the group of leaves of equal efficiency and the climatic variables was carried out, using a two-way ANOVA. For this, the value of the median Qy every 15 min was used as a sample for its correlation with radiation, temperature, VPD, humidity, dew point, atmospheric pressure, wind speed, and streaks through Pearson correlation (P < 0.05) and principal components analysis.

For the correlation with the soil variables (pH, proportion of soil particles, and resistance to penetration), the average Fv/Fm per sampling point was used, and a Pearson raster correlation was calculated in SAGA (2.3.2) using the scatter plot tool. On the other hand, the rainfall was correlated using accumulated values at 7 d before the measurement of Qy. For the correlation with weekly yield (average bunch weight), the bias that presented a higher Pearson correlation coefficient (P<0.05) with the Qy curve from leaf six was evaluated. All analyses were carried out with the statistical software R (v. 4.1.0), using the libraries



agricolae (De Mendiburu, 2021), nortest (Gross and Ligges, 2015), ggplot2 (Wickham, 2016), and GGally (Schloerke, 2021).

RESULTS AND DISCUSSION

Chlorophyll a fluorescence

Qy (Fv/Fm) is subject to the foliar development of bananas. Figure 2 shows the quantum yield of photosystem II (Qy) for each leaf emitted in the independent vegetative phenological stage (F10 \pm 11 leaves). In this stage, the banana plant emits several leaves (± 12) before floral differentiation. The Qy values increased from leaf F10 to F10 + 4, at which point the emitted leaves presented similar values for the quantum efficiency of Photosystem II. This variation was associated with the fact that even though leaf F10 is a functional leaf, it is the first completely open leaf that the sucker emits and begins the stage of independent vegetative growth; from this moment on, the leaves increase in size and functionality. From leaf number 5 (F10 + 4) to number 9 (F10 + 8), Qy remained constant. This behavior shows that, although the photosystems are active in the leaf tissue, leaf development significantly influences efficiency.



Figure 2. Maximum quantum efficiency of PSII (Qy = Fv/Fm) in each banana leaf emitted by Cavendish subgroup, Gran Nain clone. According to the Kruskal-Wallis test, boxes with different lowercase letters indicate statistical differences between leaves ($P \le 0.05$).

In banana cultivation, two stages are identified during the development of plants: a vegetative stage characterized by the emission of leaves and a productive stage easily identifiable by the emission of inflorescence. There is a phase known as the dependent vegetative during the vegetative stage where sucker development depends on the mother plant. Once the sucker emits the F10 leaf (10 cm wide), the plant enters the independent vegetative stage (Lobo *et al.*, 2020).

According to Robinson and Galán (2010), the first leaves produced from the central meristem of a developing banana regrowth (sucker) are scale leaves, followed by narrow sword-shaped leaves and broader leaves with blades that gradually widen to mature. Thus, leaf F10 signals the end of hormonal inhibition of suckers by mother plants and the beginning of the emission of leaves that begin to significantly increase their size (Donato *et al.*, 2015).

In this study, the F10+9 and F10+10 leaves showed severe attack by the phytophagous mite (*Tetranychus* sp.), commonly known as "red spider mite", which generates foliar tanning through its feeding, with a significant decrease in Qy. Similar results were reported by Chacón-Hernández *et al.* (2018), who indicated that prolonged feeding periods by *Tetranychus urticae* result in a significant decrease in the content and fluorescence of chlorophyll in rose crops; and Buffon *et al.* (2020), which verified a decrease in chlorophyll fluorescence parameters in rice species attacked by the mite *Schizotetranychus oryzae* (Acari: Tetranychidae).

Chlorophyll a fluorescence – climate relationship

When the Qy was related to the climatic variables, a significant correlation (P < 0.05) was found for the quantum yield of chlorophyll a, compared to the behavior of temperature (-0.703), VPD (-0.740), and radiation (-0.583); environmental factors that have an immediate influence on the functioning of the Photosystem II and are inversely proportional (Fig. 3).

Adams *et al.* (1989) measured Qy during the day. They found that, as the day progresses, this variable decreases as the result of an increase in temperature and radiation, which reach their maximum peaks in the afternoon. However, Qy values increase again towards the afternoon hours. The contrary was reported by Thomas and Turner (2001), who evaluated this variable during the day at total sun exposure; however, Qy decreased throughout the day. Temperature also affects quantum efficiency; Haldimann and Feller (2004) found that, when exceeding 35° C, this variable begins to reduce significantly, results that coincide with that reported by Havaux (1993) in potatoes. Although this temperature was never reached in this study, the results showed a sensitivity of bananas to temperature variations over 30° C, with a decrease in Qy.

For relative humidity, there was a high and positive correlation with Qy. It should be noted that both temperature and relative humidity are climatic variables that regulate the VPD in the atmosphere directly and inversely proportionally, respectively. VPD has been identified as one of the most determining factors in the functioning of plants and has been considered one of the more important causes of plant mortality, mainly induced by water deficits. In this sense, Grossiord *et al.* (2020) suggested that stomatal conductance decreases with a high VPD, and transpiration increases up to a given VPD threshold in most species, leading to a cascade of subsequent impacts, including reduced photosynthesis, growth, and production.

Accumulated rainfall and Fv/Fm had a low correlation of 0.13, not significant (P=0.45). This result was due to the indirect relationship that rain has with the response to stress in leaves, which also indicates that high rainfall does not translate into an increase in Oy or vice versa; the adequate availability of moisture in the soil regulates the response of the photosynthetic



apparatus. Authors such as Bukhov and Carpentier (2004) indicated that Qy is not sensitive to early or moderate water stress only when it is due to severe drought or severe water deficits and in certain species. Even under greenhouse conditions, the induction of water stress affects the variation significantly in Qy after 15 d (Živčák et al., 2008), while results obtained by Da Silva and Arrabaça (2004) did not find a correlation between Qy and rapid water stress (RWS). However, a water deficit generates dehydration and overheating of cells and tissue, reducing the CO_2 assimilation capacity and, finally, the yields of certain crops (Ravi et al., 2013). Work carried out by Robinson and Bower (1988) showed that growth and yield decreased drastically when soil moisture fell below 66% of the total available soil moisture in banana cultivation.

Chlorophyll a fluorescence - soil relationship

When the soil physical properties and the Qy were correlated, higher coefficients were presented for pH

in plot P6, with a resistance to penetration of 0.3-0.6 m in plot P12 and values of r^2 of 12.77 and 24.28%, respectively (Tab. 1).

Table 1. Correlation coefficients between the soil physical properties and the quantum yield of PSII (Fv/Fm) in banana leaves of Cavendish subgroup, Gran Nain clone.

Physical properties	Correlation coefficients* (r²) for Qy (Fv/Fm)		
	Plot P6	Plot P12	
Soil pH	12.77	2.57	
Clay (%)	1.19	0.27	
Sand (%)	0.66	2.55	
Silt (%)	0.38	3.14	
Penetration resistance 0-0.3 m (MPa)	0.34	0.08	
Penetration resistance 0.3-0.6 m (MPa)	3.79	24.18	
Penetration resistance 0.6-0.8 m (MPa)	5.63	3.29	

* Pearson raster - raster correlation using the scatterplot tool (P<0.05).

Figure 4. Spatial correlation of soil variables and the quantum yield of PSII ($\Omega y = Fv/Fm$). A: plot number 6 and B: plot number 12.

This result indicated a spatial distribution of low Qy zones with a pH lower than 5.4 (Fig. 4A) and resistance below 1.36 Mpa (Fig. 4B).

The maximum photochemical efficiency of PSII (Fv/ Fm) and the effective light quantum yield [Y (II)] in blueberry (*Vaccinium* spp.) leaves decreased as the soil pH increased, going from 0.787 (pH 4.5) to 0.745 (pH 6.0) (Jiang *et al.*, 2017). High pH values influence the absorption, transport, and distribution of nutrients in plants, lead to an imbalance of nutrients and affect plant growth and development. A change in Chlorophyll a fluorescence can indirectly reflect a shift in photosynthetic activity and yield potency. Therefore, it is possible to indirectly measure the photosynthesis capacity of plants through the fluorescence of chlorophyll (Jiang *et al.*, 2017).

Chlorophyll a fluorescence – foliar nutrient content relationship

In both plots, the Fv/Fm presented statistical difference, where the highest value was recorded in P6 (lower production) and the lowest in P12 (higher production) (Fig. 5).

This result may be related to the availability of some elements in the leaves (Tab. 2), where P12 had lower concentrations of nitrogen, manganese, and boron than P6. However, P12 had higher production, with a better balance in the different relationships between the nutritional elements in the leaf tissue. According to Cheng *et al.* (2000), for significant changes in the variation of Qy to occur, the nitrogen contents in

leaves must be below 2%, a situation that occurred for the Qy values registered in both plots of production but was lower in P6 (lower yield).

Figure 5. Maximum quantum efficiency of the PSII (Qy=Fv/Fm) in each plot of banana leaves of Cavendish subgroup, Gran Nain clone. According to the Kruskal-Wallis test, boxes with different lowercase letters indicate statistical differences between leaves ($P \le 0.05$).

Chlorophyll a fluorescence – bunch weight relationship

The weekly average bunch weight curve was evaluated with different biases (0 to 4 weeks) to the Qy

Table 2. Foliar chemical analysis of banana Cavendish subgroup, Gran Nain clone in the study areas (P6 and P12).

Mineral nutrient	Optimal ranges *	Plot			
		6	Category**	12	Category**
N %	2.70-3.60	1.97	В	1.89	В
P %	0.16-0.27	0.12	В	0.15	В
К %	3.20-5.40	1.73	В	1.94	В
Ca %	0.60-1.20	0.32	В	0.43	В
Mg %	0.27-0.60	0.17	В	0.18	В
S %	0.16-0.30	0.11	В	0.14	В
Fe (mg kg ⁻¹⁾	80-360	100	М	115	М
Mn (mg kg⁻¹)	200-1,000	190	В	150	В
Cu (mg kg ⁻¹)	6-30	12	М	16	М
Zn (mg kg ⁻¹)	20-50	23	M	35	M
B (mg kg ⁻¹)	10-25	17	М	11	М

Interpretation: B: low (poor); M: medium (sufficient); H: High. Osorio (2014).

2

Figure 6. Correlation with time-biased production and quantum yield of PSII (Qy=Fv/Fm) in banana leaves of Cavendish subgroup, Gran Nain clone. A: P6, and B: P12.

measurement, where the highest $r^2 = 0.68$ was seen four weeks after measuring the quantum yield efficiency in banana leaves in plot P6, and three weeks with $r^2 = 0.67$ in plot P12, which represents, in addition to a faster sensitivity of plot P12 to stress, a decrease of approximately 1.5 kg in the bunches of P6 and 2.9 kg in P12 after the collapse in Qy occurred in week 5 (Fig. 6).

It is important to note that, although P6 had higher Qy values than P12, it continued to present lower values in the average bunch weight, which indicated that production is sensitive to the variation in PSII efficiency within the same plot but, in terms of other plots, it is not correlated with the eigenvalue of Qy.

CONCLUSION

The variation of the Qy was correlated with environmental variables, such as radiation, temperature, VPD, relative humidity, soil pH, and soil resistance to penetration. The quantum yield of the photosystem II explained reductions in fruit yield that impacted farm productivity as a response of banana plants to any stress.

ACKNOWLEDGMENT

This study was carried out thanks to the support of the Asociación de Bananeros de Colombia (AUGURA), the Colombian Banana Research Center (CENIBANANO), and the Universidad Nacional de Colombia. The authors thank CENIBANANO for their assistance in this experiment. Thanks to the Experimental Field Staff, who provided resources for this study.

Conflict of Interest: The manuscript was prepared and reviewed with the participation of the authors, who declare that there exists no conflict of interest that puts at risk the validity of the results.

BIBLIOGRAPHIC REFERENCES

10

- Adak, M.K. 2018. Analysis of chlorophyll fluorescence: A reliable technique in determination of stress on plants. pp. 63-88. In: Sengar R. and A. Singh (eds). Eco-friendly agro-biological techniques for enhancing crop productivity. Springer, Singapur. Doi: 10.1007/978-981-10-6934-5_4
- Adams III, W.W., M. Díaz, and K. Winter. 1989. Diurnal changes in photochemical efficiency, the reduction state of Q, radiationless energy dissipation, and non-photochemical fluorescence quenching in cacti exposed to natural sunlight in northern Venezuela. Oecologia 80(4), 553-561. Doi: 10.1007/BF00380081
- AUGURA, Asociación de Bananeros de Colombia. 2021. Coyuntura bananera 2020. In: https://augura.com. co/wp-content/uploads/2021/04/Coyuntura-Bananera-2020.pdf; consulted: May, 2021.
- Barrera Violet, J.L., J.R. Cartagena Valenzuela, and O.A. Nanclares Gómez. 2020. Influence of high planting densities and arrangements on yield and fruit development of *Musa* AAA Simmonds. Acta Agron. 69(1), 46-53. Doi: 10.15446/acag.v69n1.79834
- Bouyoucos, G.J. 1936. Directions for making mechanical analyses of soils by the hydrometer method. Soil Sci. 42(3), 225-230. Doi: 10.1097/00010694-193609000-00007
- Buffon, G., E.A.R. Blasi, T.I. Lamb, J.M. Adamski, J. Schwambach, F.K. Ricachenevsky, A. Bertolazi, V. Silveira, M.C.B. Lopes, and R. Sperotto. 2020. Nipponbare and wild rice species as unexpected tolerance and susceptibility sources against *Schizotetranychus oryzae* (Acari: Tetranychidae) mite infestation. Front. Plant Sci. 12, 613568. Doi: 10.3389/fpls.2021.613568
- Bukhov, N.G. and R. Carpentier. 2004. Effects of water stress on the photosynthetic efficiency of plants. pp. 623-635. In: Papageorgiou G.C. and Govindjee (eds.). Chlorophyll a fluorescence. Advances in photosynthesis and respiration. Vol. 19. Springer, Dordrecht, The Netherlands. Doi: 10.1007/978-1-4020-3218-9 24
- Catalina Tomás, A. 2015. Utilización de medidas de fluorescencia de la clorofila para monitorizar el estado

nutricional y estimar el potencial enológico en viñedos afectados por clorosis férrica. PhD thesis. Escuela Técnica Superior de Ingenierías Agrias, Universidad de Valladolid, Palencia, Spain. Doi: 10.35376/10324/16657

- Chacón-Hernández, J.C., I. Camacho-Aguilar, E. Cerna-Chavez, S. Ordaz-Silva, Y.M. Ochoa-Fuentes, and J. Landeros-Flores. 2018. Efectos de Tetranychus urticae y Phytoseiulus persimilis (Acari: Tetranychidae: Phytoseiidae) en la clorofila de plantas de rosal (Rosa sp.). Agrociencia 52(6), 895-909.
- Cheng, L., L.H. Fuchigami, and P.J. Breen. 2000. Light absorption and partitioning in relation to nitrogen content in `Fuji' apple leaves. J. Amer. Soc. Hort. Sci. 125(5), 581-587. Doi: 10.21273/JASHS.125.5.581
- Da Silva, J.M. and M.C. Arrabaça. 2004. Photosynthesis in the water-stressed C4 grass *Setaria sphacelata* is mainly limited by stomata with both rapidly and slowly imposed water deficits. Physiol. Planta. 121(3), 409-420. Doi: 10.1111/j.1399-3054.2004.00328.x
- De Mendiburu, F. 2021. Agricolae: Statistical procedures for agricultural research. R v. 1.3-5. http://cran.r-project. org/package=agricolae; consulted: May, 2021.
- Donato, S.L.R., A.M. Arantes, E.F. Coelho, and M.G.V. Rodrigues. 2015. Considerações ecofisiológicas e estratégias de manejo da bananeira. In: VIII Simposio Brasileiro sobre Bananicultura – SIBANANA, Embrapa Mandioca e Fruticultura, Montes Claros, Brazil.
- FAO. 2021. FAOSTAT Food and agriculture data. In: https://www.fao.org/faostat/en/#home; consulted: May, 2021.
- Gross, J. and U. Ligges. 2015. Package 'nortest': Tests for normality. R v. 1.0-4. In: https://cran.r-project.org/ web/packages/nortest/index.html; consulted: May, 2021.
- Grossiord, C., T.N. Buckley, L.A. Cernusak, K.A. Novick, B. Poulter, R.T.W. Siegwolf, J.S. Sperry, and N.G. Mc-Dowell. 2020. Plant responses to rising vapor pressure deficit. New Phytol. 226(6), 1550-1566. Doi: 10.1111/ nph.16485
- Haldimann, P. and U. Feller. 2004. Inhibition of photosynthesis by high temperature in oak (*Quercus pubescens* L.) leaves grown under natural conditions closely correlates with a reversible heat-dependent reduction of the activation state of ribulose-1,5-bisphosphate carboxylase/oxygenase. Plant Cell Environ. 27(9), 1169-1183. Doi: 10.1111/j.1365-3040.2004.01222.x
- Havaux, M. 1993. Rapid photosynthetic adaptation to heat stress triggered in potato leaves by moderately elevated temperatures. Plant Cell Environ. 16(4), 461-467. Doi: 10.1111/j.1365-3040.1993.tb00893.x
- ICA, Instituto Colombiano Agropecuario. 2019. Delegación China busca aumentar, en más de un millón de toneladas, las importaciones de banano colombiano. In: https://www.ica.gov.co/noticias/ica-china-exportacion-banano; consulted: May, 2021.

- Jiang, Y., Y. Li, Q. Zeng, J. Wei, and H. Yu. 2017. The effect of soil pH on plant growth, leaf chlorophyll fluorescence and mineral element content of two blueberries. Acta Hortic. 1180, 269-276. Doi: 10.17660/ actahortic.2017.1180.36
- Kim, J.H., S.R. Bhandari, S.Y. Chae, M.C. Cho, and J.G. Lee. 2019. Application of maximum quantum yield, a parameter of chlorophyll fluorescence, for early determination of bacterial wilt in tomato seedlings. Hortic. Environ. Biotechnol. 60(6), 821-829. Doi: 10.1007/ s13580-019-00182-0
- Lobo, M.G. and F.J. Fernández Rojas. 2020. Biology and postharvest physiology of banana. pp. 19-44. In: Siddiq, M., J. Ahmed, and M.G. Lobo (eds.). Handbook of banana production, postharvest science, processing technology, and nutrition. Wiley, Hoboken, NJ. 10.1002/9781119528265.ch2
- Manterola D., C. and D. Zavando M. 2009. Cómo interpretar los "Niveles de Evidencia" en los diferentes escenarios clínicos. Rev. Chil. Cir. 61(6), 582-595. Doi: 10.4067/s0718-40262009000600017
- Maxwell, K. and G.N. Johnson. 2000. Chlorophyll fluorescence — a practical guide. J. Exp. Bot. 51(345), 659-668. Doi: 10.1093/jexbot/51.345.659
- Osorio, N.W. 2014. Manejo de los nutrientes en suelos del trópico. 2nd ed. Universidad Nacional de Colombia. Editorial LA Vieco, Medellin, Colombia.
- Pérez Asseff, J.M., E.J. Peña Salamanca, and C. Torres González. 2017. Efecto del nitrógeno y la irradianza en la eficiencia fotosintética del anamú *Petiveria alliacea* (Phytolaccaceae). Rev. Acad. Colomb. Cienc. Fis. Nat. 31(118), 49-55.
- Pérez, W.A. and J. Torres-Bazurto. 2020. Carbon-nitrogen ratio in soils with fertilizer applications and nutrient absorption in banana (*Musa* spp.) cv. Williams. Agron. Colomb. 38(2), 253-260. Doi: 10.15446/agron.colomb. v38n2.78075
- Ravi, I., S. Uma, M.M. Vaganan, and M.M. Mustaffa. 2013. Phenotyping bananas for drought resistance. Front. Physio. 4, 9. Doi: 10.3389/fphys.2013.00009

- Robinson, J.C. and J.P. Bower. 1988. Transpiration from banana leaves in the subtropics in response to diurnal and seasonal factors and high evaporative demand. Sci. Hortic. 37(1-2), 129-143. Doi: 10.1016/0304-4238(88)90156-2
- Robinson, J.C. and V. Galán S. (eds.). 2010. Bananas and plantains. Serie Crop Production Science in Horticulture Vol. 19. 2nd ed. Cabi, Wallingford, UK. Doi: 10.1079/9781845936587.0000
- Sánchez-Reinoso, A.D., Y. Jiménez-Pulido, J.P. Martínez-Pérez, C.S. Pinilla, and G. Fischer. 2019. Chlorophyll fluorescence and other physiological parameters as indicators of waterlogging and shadow stress in lulo (Solanum quitoense var. septentrionale) seedlings. Rev. Colomb. Cienc. Hortíc. 13(3), 325-335. Doi: 10.17584/ rcch. 2019v13i3.100171
- Sanz Miguel, M.A., A. Blanco Braña, E. Monje, and J. Val Falcón. 2001. Caracterización de la deficiencia de calcio en plantas de tomate utilizando parámetros fisiológicos. Revista ITEA 97V(1), 26-38.
- Schloerke, B., Di Cook, J. Larmarange, F. Briatte, M. Marbach, E. Thoen, A. Elberg, O. Toomet, J. Crowley, H. Hofmann, and H. Wickham. 2021. GGally: Extension to 'ggplot2'. R v. 2.1.1. In: https://CRAN.R-project. org/package=GGally; consulted: May, 2021.
- Thomas, D.S. and D.W. Turner. 2001. Banana (*Musa* sp.) leaf gas exchange and chlorophyll fluorescence in response to soil drought, shading and lamina folding. Sci. Hortic. 90(1-2), 93-108. Doi: 10.1016/ s0304-4238(00)00260-0
- Wickham, H. 2016. ggplot2: Elegant graphics for data analysis. Springer-Verlag, New York. Doi: 10.1007/978-3-319-24277-4
- Zhang, J.Z., Q. Zhang, Y.J. Chen, L.L. Sun, L.Y. Song, and C.L. Peng. 2012. Improved tolerance toward low temperature in banana (*Musa* AAA Group Cavendish Williams). S. Afr. J. Bot. 78, 290-294. Doi: 10.1016/j. sajb.2011.08.002
- Živčák, M., M. Brestič, K. Olšovská, and P. Slamka. 2008. Performance index as a sensitive indicator of water stress in *Triticum aestivum* L. Plant Soil Environ. 54(4), 133-139. Doi: 10.17221/392-pse