

Selectivity of insecticides for adult workers of *Apis mellifera* (Hymenoptera: Apidae)

Selectividad de insecticidas sobre obreras adultas de *Apis mellifera* (Hymenoptera: Apidae)

DANIELLE THOMAZONI¹, MIGUEL F. SORIA², CÁSSIO KODAMA², VLADSON CARBONARI¹,
RONI P. FORTUNATO², PAULO E. DEGRANDE¹ and V. A. Jr., VALTER¹

Abstract: Cotton is a crop that is attractive to a wide variety of organisms, including pests, natural enemies of the pests and arthropod pollinators. Consequently, integrated pest management is necessary for this crop as chemical control is often required to avoid production losses. Among pollinators found on cotton, *Apis mellifera* is one of the most important, as it can increase cotton production 20 – 30%. On the other hand, selectivity is an important characteristic of insecticides, as it makes it possible to lessen impacts on pollinators and other non-target organisms. We examined the toxicity of various insecticides for honey bees foraging on cotton in a greenhouse, including Turbine® 500 WG (150 g ha⁻¹) (flonicamid), Actara® 250 WG (200 g ha⁻¹) (thiamethoxam), Cartap® 500 SP (1500 g ha⁻¹) (cartap), Talstar® 100 CE (1000 mL ha⁻¹) (bifenthrin) and Match® 50 CE (1000 mL ha⁻¹) (lufenuron). Applications of Talstar®, Actara® and Cartap® were quite toxic. Talstar® was the most deadly, while Match CE® and Turbine® were only slightly toxic for adult honey bees. The Match CE® treatment mortality was similar to that of the control; thus, we classified it as innocuous for *A. mellifera* adults.

Key words: Pollinator. Non-target effects. Environmental impact.

Resumen: El algodón es un cultivo atractivo a una amplia variedad de organismos incluyendo plagas, enemigos naturales de las plagas y los polinizadores. Por lo tanto, el manejo integrado de plagas es necesario para este cultivo ya que el control químico se requiere a menudo para evitar pérdidas de producción. Entre los polinizadores encontrados en algodón, *Apis mellifera* es uno de los más importantes pues puede aumentar la producción de 20 – 30% del algodón. Por otro lado, la selectividad es una característica importante de insecticidas, pues permite disminuir impactos en polinizadores y otros organismos no-blanco. Se examinó la toxicidad de varios insecticidas sobre abejas de *Apis mellifera* forrajeando en el algodón en condiciones de invernadero, incluyendo Turbine® 500 WG (150 g ha⁻¹) (flonicamid), Actara® 250 WG (200 g ha⁻¹) (thiametoxam), Cartap® 500 SP (1500 g ha⁻¹) (cartap), Talstar® 100 CE (1000 ml ha⁻¹) (bifenthrin) y Match® 50 CE (1000 ml ha⁻¹) (lufenuron). Las aplicaciones de Talstar®, Actara® y Cartap® fueron absolutamente tóxicas. Talstar® fue el más mortal, mientras que el Match® CE y Turbine® fueron solamente levemente tóxicos para los adultos de la abeja. El tratamiento Match® CE presentó mortalidad similar a la del control; entonces, lo clasificamos como inofensivo para los adultos de *A. mellifera*.

Palabras clave: Polinizadores. Efecto no blanco. Impacto ambiental.

Introduction

Although cotton (*Gossypium hirsutum* L.) is attacked by agricultural pests that damage roots, stems, leaves, flower buds, flowers, fruits and bolls, it is also host to various pollinators and natural enemies such as predators, parasitoids and pathogens, which are indispensable components of dynamic agroecosystems. Damage inflicted by agricultural pests can drastically reduce cotton productivity, directly affecting seed and fiber characteristics, and reducing their commercial value (Degrande and Gomes 1990). Consequently, pest control techniques, such as cultural, chemical and biological tactics that are part of Integrated Pest Management (IPM) of cotton, have proven to be extremely important for the success of this crop. Their use has increased significantly as it is considered a safe methodology that provides a satisfactory cost/benefit ratio (Degrande 2000; Gallo *et al.* 2002).

Among control methods, chemicals are the most widely employed due to their ease of application and effectiveness

(Santos 1999). However, careless use of this form of control results in adverse effects on non-target organisms, including humans, domestic animals, pollinators, natural enemies and other wild organisms, in addition to contaminating soil and water. These side effects need to be eliminated or minimized in order to avoid problems such as pest reinfestations, secondary pest outbreaks, and evolution of pest resistance (Metcalf 1982; Nakano 1986).

Among the beneficial pollinator insects that visit cotton, the Africanized honey bee, *Apis mellifera* L., 1752 (Hymenoptera: Apidae), is prominent. In addition to its ecological importance as a pollinator, this insect has its own economic importance as a producer of honey, beeswax and royal jelly. Some studies have demonstrated an increase in cotton fruit and seed production on the order of 20 – 30% when honey bees pollinate this crop, which has also been found for other crops, such as sunflower and onion seed (Fermiano 1981; Lorenzon and Martinho 1992; Moreti *et al.* 1996; Gallo *et al.* 2002). Laboratory tests of the effect of insecticides ap-

¹ Universidade Federal da Grande Dourados, Programa de Pós-graduação em Entomologia e Conservação da Biodiversidade, Faculdade de Ciências Biológicas e Ambientais, Departamento de Ciências Biológicas/Unidade II. Rodovia Dourados-Itahum, Km 12, Caixa Postal: 241, Cep: 79804-970, Dourados/MS, Brazil.

² Universidade Federal da Grande Dourados, Programa de Pós-graduação em Produção Vegetal, Faculdade de Ciências Agrárias, Departamento de Ciências Agrárias/Unidade II. Rodovia Dourados-Itahum, Km 12. Cidade Universitária. Aeroporto. Caixa Postal: 533, CEP: 79.804-970, Dourados, MS, Brasil. paulodegrande@ufgd.edu.br. Autor para correspondência.

plied to soybean plants on honey bees have demonstrated that pyrethroids are the most dangerous (100% mortality within two hours after spraying), while carbamates and organophosphates elicited 50% mortality under the same conditions (Rigotti 2005).

If insecticides are used in an appropriate manner, they can control target organisms without negatively affecting natural enemy and pollinator populations; otherwise, these predators and pollinators will be less able to provide their beneficial services (Melksham *et al.* 1988; Davis 1989). One way to achieve this objective would be to use selectively toxic insecticides (Carvalho *et al.* 2002; Degrande *et al.* 2002; Magalhães *et al.* 2002; Godoy *et al.* 2004). Considering the importance of *A. mellifera* as a pollinating agent in agroecosystems, we evaluated the effect of the insecticides Turbine® 500 WG (flonicamid), Actara® 250 WG (thiamethoxam), Cartap® 500 SP (cartap), Talstar® 100 CE (bifenthrin) and Match 50 CE® (lufenuron), which are used on cotton, on the mortality of honey bee workers.

Material and Methods

The experiment was conducted in a greenhouse, with potted cotton plants at the Faculdade de Ciências Agrárias (FCA) of the Universidade Federal da Grande Dourados (UFGD), in Dourados (22°14'S, 54°44'W, altitude 452 m), Mato Grosso do Sul state, Brazil, from November 2006 to January 2007. Each pot was planted with six seeds of cotton cultivar Fibermax 933, later reduced to four plants per pot. The soil used for the plants' cultivation was an Oxisol collected from the subsoil (> 40 cm deep). During flowering, approximately 50 – 55 days after germination, the plants were sprayed with the insecticide treatments at a rate of 400 liters of spray liquid per hectare. The sprayer was washed with water, water + detergent and 92.8% ethanol between treatments, to remove residues of the different insecticides.

The experimental design consisted of randomized blocks, with six treatments and four replicates per treatment. Each

plot consisted of a pot with four plants and 30 *A. mellifera* adult workers, which were confined in gauze cages 98.5×41 cm. The treatments were: 1) Turbine® 500 WG (150 g/ha) (flonicamid), 2) Actara® 250 WG (200 g/ha) (thiamethoxam), 3) Cartap® 500 SP (1500 g/ha⁻¹) (cartap), 4) Talstar® 100 CE (1000 mL/ha⁻¹) (bifenthrin), 5) Match 50 CE® (1000 mL/ha) (lufenuron) and 6) a control sprayed with water; in every case a manual sprayer at a constant pressure was used. Spraying began at 9:00 h, at 29°C, 68% relative humidity and wind speed below 2.5 m s⁻¹, at a rate of 112 L/ha⁻¹, these conditions are those usually adopted in the field for insecticide sprays. The spray was allowed to dry on the plants for three hours before adding the 30 bees per pot. Four replicates were made for each insecticide treatment. Each group of bees was provided with PET bottle caps containing water and with a mixture of honey plus sucrose composed of 50% honey or water and 50% refined sugar (Wiese 1995).

The *A. mellifera* workers were estimated to be about 5-6 days old, based on their coloration and hair density on the thorax, plus their in-colony activities (Free 1980). The bees were collected from two colonies located in apiaries near the university (FCA/UFGD), using a manual aspirator, and were maintained in a B.O.D incubator (28±1°C and 12 hours photophase) for 24 hours; they were fed with the sucrose and honey mixture plus water until they were placed in the gauze cages in the greenhouse. Evaluations were made every 30 minutes for six hours, counting the number of bees that had died. For each evaluation the data were tabulated, expressed as percentage mortality and transformed with square root of (x+0.5) for the statistical analysis. The means were compared by the Tukey's test (P < 0.05).

Results and Discussion

Among the six insecticide treatments, Talstar® 100 CE (1000 mL ha⁻¹) was the least selective, causing 100% mortality of *A. mellifera* workers in the shortest time (Figs. 1 and 2). This

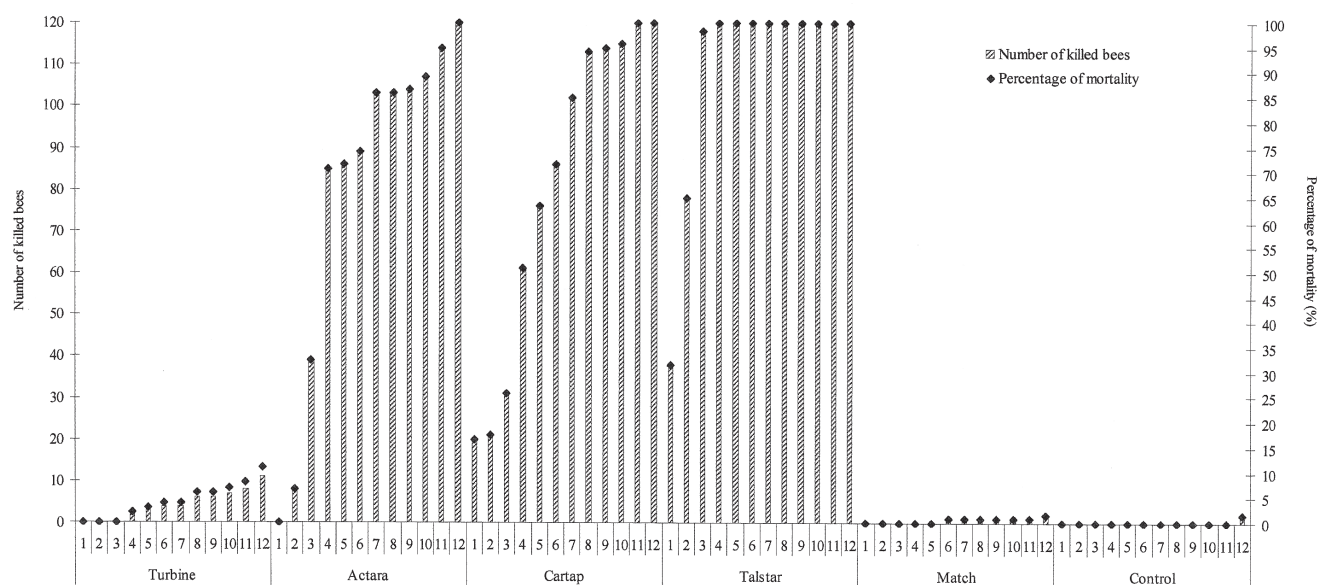


Figure 1. Frequency of accumulated mortality of *Apis mellifera* adult workers over six hours after spraying insecticides on cotton plants in a greenhouse in Dourados, MS, Brazil. 2007. Time after spraying 1 = 30 min., 2 = 60 min., 3 = 90 min., 4 = 120 min., 5 = 150 min., 6 = 180 min., 7 = 210 min., 8 = 240 min., 9 = 270 min., 10 = 300 min., 11 = 330 min., 12 = 360 min.

product is similar to the pyrethroid bifenthrin, which acts on the modulation of Na⁺ channels in the central and peripheral nervous systems of insects. It has a potent “knock-down” effect, and after contact with this insecticide, the workers displayed uncoordinated movements and trembling, manifesting the typical effects of pyrethroid insecticides, also observed by Carvalho (2006). Rigotti (2005) also found that pyrethroid-based insecticides used commercially on soybeans caused 100% mortality of *A. mellifera* adults, within two hours of spraying in the lab.

Treatments with the insecticides Actara® 250 WG (200 g ha⁻¹) (thiamethoxam) and Cartap® 500 SP (1500 g ha⁻¹) (cartap), which affect the insect nervous system, were also not selective, although they killed the bees at a slower rate than did bifenthrin. Total mortality (100%) of bee workers occurred after 360 minutes with Cartap® and after 330 minutes with Actara® (Fig. 1). Carvalho *et al.* (2004 a,b) and Carvalho (2006) reported similar effects on *A. mellifera* workers, independent of the mode of application whether direct spraying or incorporation in the diet; thiamethoxam was highly toxic to bees, requiring a mean of 22.5 hours to kill 100% of the workers. High toxicity of thiamethoxam was also found by Iwasa *et al.* (2004), who reported an LD₅₀ of 0.03 µg/bee. Ac-

cording to Karnatak and Neetu (2005), thiamethoxam is one of the most toxic insecticides for *A. mellifera*.

Similar results were reported by Thompson (2003) and Rhodes and Scott (2006), who classified thiamethoxam as highly toxic. They also indicated that this insecticide can elicit physiological and behavioral effects in the bees, when applied at sublethal doses. These include alterations in the embryonic, larval and pupal phases, as well as changes in flight activity, negatively affecting food collection and consequently colony development.

Treatments with the insecticides Turbine® (150 g ha⁻¹) (flonicamid) and Match 50 CE® (1000 mL ha⁻¹) (lufenuron) were less toxic for *A. mellifera* workers. Match 50 CE® did not significantly affect the bees and could be considered innocuous for them (Figs. 1 and 2). Similar results were found with the predator wasp *Polistes canadensis canadensis* (L., 1758) (Santana-Reis *et al.* 2002) in laboratory tests with lufenuron. This is a chitin-synthesis regulator, acting on ecdysis during the insect larval stages.

Based on our greenhouse evaluations, the insecticide Turbine® was considered selective or relatively non-toxic for bees, as was Match CE®, eliciting only 9.16% mortality (Figs. 1 and 2). Carvalho (2006) also tested lufenuron directly on bees in the lab and found low toxicity, similar to that found for the control, reaching 21% after 72 hours of confinement, and similar to what was reported by Atkins *et al.* (1981), Tew (1996) and Hunt (2000).

Conclusions

Spraying with Talstar®, Actara® and Cartap® was lethal for *A. mellifera*; Talstar® was the most acutely toxic. Match CE® and Turbine® were less toxic; mortality due to Match CE® was similar to that obtained with the control spray. We classified Match CE® as innocuous for adult honey bees under these conditions. This research contributes information that can help conserve *A. mellifera* populations in the vicinity of cotton crops, contributing to the productivity of the cotton crop and to the quality of bee products in neighboring aparies.

Literature cited

- ATKINS, E. L., KELLUM, D.; ATKINS, K. W. 1981. Reducing pesticides hazardous to honeybees. Mortality prediction techniques and integrated management strategies. Berkeley: University of California. 20 p.
- CARVALHO, G. A.; CARVALHO, C. F.; SOUZA, B.; ULHÔA, J. L. R. 2002. Seletividade de inseticidas a *Chrysoperla externa* (Hagen) (Neuroptera: Chrysopidae). Neotropical Entomology 31 (4): 615-621.
- CARVALHO, S. M.; CARVALHO, G. A.; CARVALHO, C. F.; BRIGHENTI, D. M. 2004a. Toxicidade de produtos fitossanitários utilizados em citros sobre operárias de *Apis mellifera* Linnaeus, 1758 (Hymenoptera: Apidae), pp. 547. In: Congresso Brasileiro de Entomologia 20, 2004, Gramado. Proceedings. Gramado.
- CARVALHO, S. M.; CARVALHO, G. A.; CARVALHO, C. F.; MOSCARDINI, V. F. 2004b. Efeito de produtos fitossanitários recomendados para a cultura de citros sobre operárias de *Apis mellifera* Linnaeus, 1758 (Hymenoptera: Apidae) fornecido por meio de pasta Candi contaminada, pp. 547. In: Congresso Brasileiro de Entomologia 20, 2004, Gramado. Proceedings. Gramado.

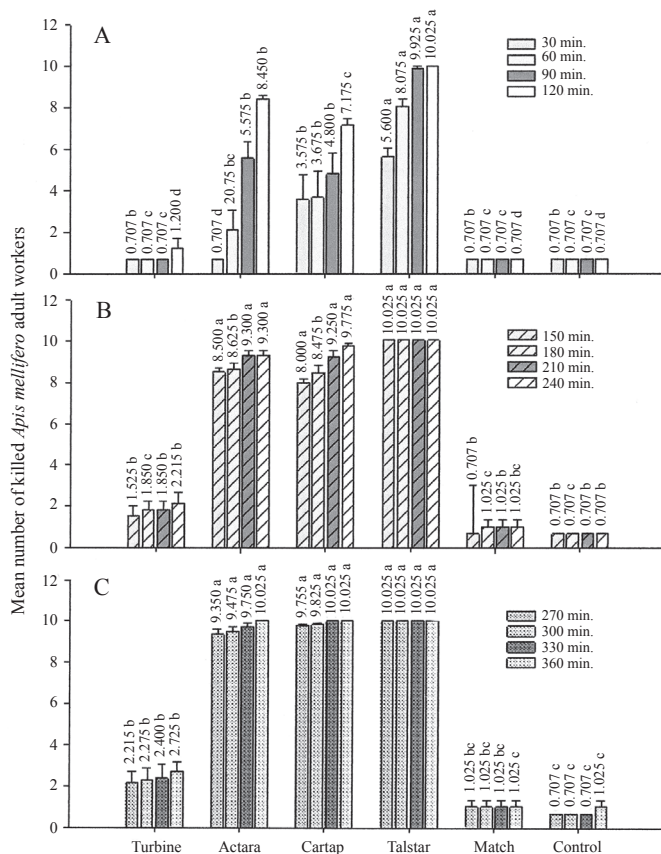


Figure 2. Mean number of *Apis mellifera* adult workers killed by the action of various insecticides applied to cotton in a greenhouse in Dourados, MS, Brazil. 2007. (n=4). Means (±SE) with the same letter within the same period of evaluation are not significantly different (Tukey's test, P < 0.05). **A.** Periods of evaluation at 30, 60, 90 and 120 minutes after insecticide application. **B.** periods of evaluation at 150, 180, 210 and 240 minutes after insecticide application and **C.** Periods of evaluation 270, 300, 330 and 360 minutes after insecticide application.

- CARVALHO, S. M. 2006. Toxicidade de produtos fitossanitários utilizados na cultura de citros a operárias de *Apis mellifera* Linnaeus, 1758 (Hymenoptera: Apidae). M.Sc. thesis – Universidade Federal de Lavras. 72 p.
- DAVIS, A. R. 1989. The study of insecticide poisoning of honeybee brood. *Bee World* 4: 163-174.
- DEGRANDE, P. E. 2000. Manejo de pragas: realidade e desafios, pp. 229-244. In: Congresso Internacional do Agronegócio do Algodão/ V Seminário Estadual da Cultura do Algodão: Negócios e Tecnologias para melhorar a vida. Proceedings of the International Congress of Cotton Agribusiness. Cuiabá-MT-Brasil.
- DEGRANDE, P. E.; GOMES, D. R. S. 1990. Seletividade de produtos químicos no controle de pragas. *Agrotécnica*: 8-13.
- DEGRANDE, P. E.; REIS, P. R.; CARVALHO, G. A.; BELARMINO, L. C. 2002. Metodologia para avaliar o impacto de pesticidas sobre inimigos naturais, pp. 71-93. In: Parra, J. R. P.; Botelho, P. S. M.; Corrêa-Ferreira, B. S.; Bento, J. M. S. Controle Biológico no Brasil. São Paulo, Manole.
- FERMIANO, L. H. M. N. 1981. Estudo do comportamento de coleta de alimento em duas subespécies de *Apis mellifera* (Hymenoptera, Apidae) e nos descendentes F1 resultantes de seu cruzamento. M.Sc. thesis. FMRP-USP, 81 p.
- FREE, J. B. 1980. Organização social das abelhas (*Apis*). Edusp. 79 p.
- GALLO, D.; NAKANO, O.; SILVEIRA, S.; CARVALHO, R.; BAPTISTA, G.; BERTI, E.; PARRA, J. R.; ZUCCHI, R. A.; ALVES, S. B.; VENDRAMIM, J. D.; MARCHINI, L. C.; LOPES, J. S.; OMOTO, C. 2002. Entomologia Agrícola. Biblioteca de Ciências Agrárias Luiz de Queiroz. São Paulo: FEALQ. 920 p.
- GODOY, M. S.; CARVALHO, G. A.; MORAES, J. C.; JÚNIOR, M. G.; MORAIS, A. A.; COSME, L. V. 2004. Seletividade de inseticidas utilizados na cultura dos citros para ovos e larvas de *Chrysoperla externa* (Hagen) (Neuroptera: Chrysopidae). *Neotropical Entomology* 33: 639-646.
- HUNT, G. J. 2000. Using honey bees in pollination. *Beekeeping*: E-216-W. Purdue University Cooperative Extension Service. 5 p.
- IWASA, T.; MOTOYAMA, N.; AMBROSE, J. T.; ROE, R. M. 2004. Mechanism for the differential toxicity of neonicotinoid insecticides in the honey bee, *Apis mellifera*. *Crop Protection* 23 (5): 371-378.
- KARNATAK, A. K.; NEETU, S. 2005. Relative toxicity of some insecticides to the workers of *Apis mellifera* L. *Shashpa* 12 (1): 23-25.
- LORENZON, M. C. A.; MARTINHO, M. R. 1992. Polinizadores da floração da cebola em Igarapé, estado de Minas Gerais. *Anais da Sociedade Entomológica do Brasil* 3: 383-389.
- MAGALHÃES, L. C.; GUEDES, R. N. C.; OLIVEIRA, E. E.; TUELHER, E. S. 2002. Desenvolvimento e reprodução do predador *Podisus distinctus* (Stål) (Heteroptera: Pentatomidae) frente a doses subletais de permetrina. *Neotropical Entomology* 31: 445-448.
- MELKSHAM, K.; JACOBSEN, J. N.; RHODES, J. 1988. Compounds which affect the behaviour of the honeybee, *Apis mellifera* L.: a review. *Bee World* 3: 105-124.
- METCALF, R. L. 1982. Insecticides in pest management, pp. 217-277. In: Metcalf, R. L.; Luckman, W. H. (Ed.). Introduction to insect pest management. 2nd ed. New York: John Wiley.
- MORETI, A. C.; SILVA, C. C.; SILVA, R. M. B.; SILVA, E. C. A. 1996. Aumento na produção de sementes de girassol (*Helianthus annuus*) pela ação de insetos polinizadores. *Scientia Agrícola* 2: 13-16.
- NAKANO, O. 1986. Avanços na prática do controle de pragas. *Informe Agropecuário* 12: 55-59.
- RHODES, J.; SCOTT, M. 2006. Pesticides: a guide to their effects on honey bees. NSW Department of Primary Industries: Primefacts 149. 4 p.
- RIGOTTI, M. 2005. Efeito da pulverização de inseticidas utilizados na cultura de soja sobre adultos de *Apis mellifera* Linnaeus (Hymenoptera, Apidae) em condições de laboratório. M.Sc. thesis – Universidade Federal da Grande Dourados, 34 p.
- SANTANA-REIS, V. P. G.; MARQUES, O. M.; COSTA, J. C. 2002. Seletividade de inseticidas ao predador *Polistes canadensis canadensis* (L., 1758) Hymenoptera: Vespidae. *Acta Biológica Leopoldensia* 2: 141-146.
- SANTOS, W. J. 1999. Monitoramento e controle das pragas do algodoeiro, pp. 133-179. In: Cia, E.; Freire, E. C.; Santos, W. J. dos. Cultura do algodoeiro. POTAFOS: Associação Brasileira para pesquisa da potassa e do fosfato. Piracicaba-SP.
- TEW, J. E. 1996. Protecting honey bees from pesticides. Ohio: The Ohio State University (Ohio State University Extension. Circular HYG-2161-97). 10 p.
- THOMPSON, H. M. 2003. Behavioural effects of pesticides in bees – their potential for use in risk assessment. *Ecotoxicology* 12 (1/4): 317-330.
- WIESE, H. 1995. Novo Manual De Apicultura. Livraria Editora Agropecuária. 292 p.

Received: 14-sep-2008 • Accepted: 11-jul-2009