HS-SPME GC/MS Volatile profile of the onion Allium fistulosum L. variety Pereirana, cultivated in Colombia



Perfil volátil HS-SPME GC/MS de la cebolla Allium fistulosum L. variedad Pereirana, cultivada en Colombia

https://doi.org/10.15446/rfnam.v77n1.105955

Juan Pablo Arrubla Vélez^{1*}, Santiago Uribe Tabares¹ and Norma Patricia Durán²

ABSTRACT

Keywords: GC-MS HS-SPME VOCs Welsh onion

MEFS COVs

The study presents a characterization of the volatile organic compounds found in both raw and the essential oil of the "Pereirana Onion," an endemic Colombian Welsh onion variety for which the composition has not been previously reported. The analysis was conducted using four distinct fibers through the HS-SPME/GC-MS method. The results revealed that chopped Pereirana onions release as many as 29 different compounds, with concentrations up to 20 times higher than those observed in other evaluated onion species (Biónica, Veleña, and Veleña Sonsón). Most of these compounds are sulfur-based, including dipropyl disulfide, (E)-1-(Prop-1-en-1-yl)-2-propyldisulfane, disulfide, methyl 1-(methylthio) propyl, dipropyl trisulphide, and (E)-1-(Prop-1-en-1-yl)-3-propyltrisulfane. Furthermore, steam extraction of essential oils from Pereirana onions led to the identification of up to 70 different compounds. Simple correspondence analysis (SCA) revealed that Veleña and Veleña Sonsón onion species share common compounds but significantly differ from Biónica and Pereirana varieties cultivated in Risaralda, Colombia. These findings suggest potential applications in the pharmaceutical, agricultural, and food industries, paving the way for future research and industrial utilization.

RESUMEN

El estudio presenta una caracterización de los compuestos orgánicos volátiles encontrados en la Palabras clave: "Cebolla Pereirana", una variedad endémica de cebolla de rama en Colombia para la cual no se CG-EM había informado previamente sobre su composición. El análisis se realizó utilizando cuatro diferentes fibras por la técnica HS-SPME/GC-MS. Los resultados revelaron que las cebollas Pereirana picadas liberan hasta 29 compuestos diferentes, con concentraciones hasta 20 veces más altas que las Cebolla de rama observadas en otras especies de cebolla evaluadas (Biónica, Veleña y Veleña Sonsón). La mayoría de estos compuestos son de origen organosulfurado, incluyendo el disulfuro de dipropilo, (E)-1-(Prop-1-en-1-il)-2-propildisulfano, disulfuro, metil 1-(metiltio) propilo, trisulfuro de dipropilo y (E)-1-(Prop-1-en-1-il)-3-propiltrisulfano. Además, la extracción de aceites esenciales mediante arrastre por vapor de las cebollas Pereirana permitió identificar hasta 70 compuestos diferentes. El análisis de correspondencia simple (SCA) reveló que las especies de cebolla Veleña y Veleña Sonsón comparten compuestos comunes, pero difieren significativamente de las variedades Biónica y Pereirana cultivadas en Risaralda, Colombia. Estos hallazgos sugieren aplicaciones potenciales en las industrias farmacéutica, agrícola y alimentaria, allanando el camino para futuras investigaciones v utilización industrial.

10leochemistry Research Group, Universidad Tecnológica de Pereira, Colombia. juanpablo77@utp.edu.co 💿, santiago.uribe@utp.edu.co 💿 2School of Chemical Technology, Universidad Tecnológica de Pereira, Colombia. npatricia@utp.edu.co 回 *Corresponding author



10612

lobal onion production in 2022 was 93.23 million tons (MT), being China (23.91 MT), India (19.42 MT), Egypt (3.12 MT), USA (3.03 MT), and Iran (2.35 MT) as the top five producers (Kumar et al. 2022). In Colombia, Welsh onion (*Allium fistulosum*) has been grown since 1950 and is grown from 1,500 to 3,000 meters above sea level (masl), between 12 and 20 °C. According to the Colombian statistics department, in 2019, in Colombia, 16,922 ha of Welsh onion were harvested, with a production of 414,554 tons (DANE 2019).

The Department of Risaralda is the third largest producer of onions in Colombia, with 50,000 tons per year, cultivated since 1960 (Jaramillo and Vallejo 2014). Given the environmental conditions that did not favor cultivation, the plant adapted to the environment, giving rise to a unique material, widely accepted by consumers, called 'Cebolla Pereirana'. This plant was morpho-agronomically characterized by exhibiting thin, dark, and violet pseudostems, dark green leaves, intermediate foliage, high tillering, and abundant serosity in leaves, as well as by being resistant to breaking and a cause of strong irritation of the eyes when the leaves are squeezed (Polanco and Pérez 2018).

Fresh onions are a cornerstone of global cuisine due to their distinctive flavor, which is derived from sulfur compounds found in the volatile fraction, commonly referred to as onion oil. This oil is obtained via steam distillation of crushed onions, yielding a complex mixture predominantly composed of mono, di, tri, and tetrasulfides with various alkyl groups (Hosoda et al. 2003). As a result, onion oil has been traditionally and extensively utilized in the food industry to augment the flavors of processed foods (including soups, meats, and ready-to-eat meals), sauces, salad dressings, and dips. This method effectively bypasses the difficulties associated with managing large volumes of fresh produce (Lawless 2012). Given the widespread use of both fresh onions and onion essential oil, this study incorporates both materials.

The genus Allium has a wide variety of sulfur compounds responsible for taste and smell, with marked differences in volatile organic compounds (VOCs) in the various onion species. The compound responsible for generating this variety of compounds is S-alk(en)yl-L-cysteine sulfoxides (ACSOs), thanks to the enzymatic activity of allinase. This pattern of sulfur compounds is common to most *Allium* species. The VOCs profile is concentrated, and the result is a mixture of sulfur-containing compounds including thiosulfinates, mono-, di- and tri-sulfides as well as specific compounds such as the lachrymatory or tear factor, thiopropanal S-oxide (Colina-Coca et al. 2013).

The sulfur compounds are extracted using solvent-free liquid-liquid extraction, hydro distillation, or headspace solid-phase microextraction (HS-SPME). The extraction process can be regarded as a dynamic equilibrium involving the target compounds evaporating into the headspace and their subsequent adsorption onto and desorption from the fiber. The process depends partly on the chemistry and physical dimensions of the fiber coating (exposed area and film thickness), in addition to the aerodynamics inside the headspace (Peng et al. 2020).

The HS-SPME/GC-MS technique used to analyze VOCs is performed with small sample amounts, and avoids the use of solvents, promoting sustainable chemistry and detecting a wide variety of compounds simultaneously (Licen et al. 2021). The SPME is based on the equilibrium of partition of compounds of interest, between the matrix and the extraction phase that covers the surface of the fused silica fiber. This fiber should be selected according to the selectivity of the compounds to be assessed, adapting the time of exposure, equilibrium temperature, and dynamic or static mode, which is decisive in obtaining adequate detection limits.

The efficiency of this technique depends on the polarity of the fiber, and polydimethylsiloxane (PDMS) coated fibers have been satisfactory in most cases. Previously, the VOCs of fresh leaves of *Allium fistulosum* and *Allium tuberosum* were studied with HS-SPME/GC-MS and PDMS fibers, finding dipropyl disulfide (67%), 1-propenylpropyl disulfide (23%), and dipropyl trisulphide (6%) (Hori 2007). The genus *Allium* has also been studied with CAR/PDMS fibers, detecting characteristic sulfides such as propyl mercaptan, dimethyl disulfide, 2,5-dimethylthiophene, and allyl propyl disulfide (Zhang and Li 2007).

The present study contributes to the knowledge of the endemic material 'Cebolla Pereirana', which has its production system and is a product highly appreciated by consumers for its attractive spicy flavor and intense smell for culinary applications. However, there is no evidence of studies reporting the volatile compounds present in this onion variety. This is the first study assessing VOCs using HS-SPME/GC-MS, comparing four fibers and four onion varieties (Pereirana, Biónica, Veleña, and Veleña Sonsón) grown in Pereira, Department of Risaralda, Colombia.

MATERIALS AND METHODS Onion samples

The samples were taken in Pereira, Colombia, during the second semester of 2021, at Castilla and Carmela farms, both at coordinates 4°45'21.4"N 75°36'32.0" W (4.7559450, -75.6088910). The sampling was performed randomly in a zig-zag route, taken between four and six shoots (stems, leaves, and roots) per hectare (4 ha) (Silva and Uchida 2000), of each of the four Welsh onion varieties, namely: (a) Pereirana; (b) Veleña; (c) Veleña Sonsón; and (d) Biónica.

Volatile organic compound analysis Fresh sample preparation

The samples were transported at 4 °C and stored in a refrigerator at the same temperature. The assay was conducted three times. The leaves and stems of each variety weighing 1 g were finely cut with a knife and stored in a hermetically-sealed SPME vial (D'Auria and Racioppi 2017). HS-SPME-GC-MS Analysis of onion (*Allium* cepa L.) and shallot (*Allium* ascalonicum L.). Food Res, 1(5), 161-165. For each sample, the HS-SPME analysis was performed at a temperature of 45 °C with a time of 5 min of homogenization and 15 min of exposure to the fibers divinylbenzene/carboxen/polydimethylsiloxane (DVB/CAR/PDMS), polyacrylate, polydimethylsiloxane (PDMS).

Extraction of Essential Oil from the Pereirana Variety Onion

The leaves and stems were chopped and taken to the steam-dragging equipment. The assay was conducted three times (Boutekedjiret et al. 2003; Peredo et al. 2009) with a sample ratio of solvent 1:1. The distillate was re-extracted with ethyl acetate and concentrated in a rotary evaporator. The solvent was evaporated in a rotary evaporator at 40 °C at 240 mbar. Subsequently, 3 μ L of the essential oil were diluted (1:1,000) with distilled water, stirring at 6,000 rpm, and brought into equilibrium for 20 min at 40 °C (Peredo et al. 2009). Then, the SPME

fiber was exposed in the headspace for 30 min at 40 °C (triplicated analysis) (Soto et al. 2015). The qualitative tentative analysis was performed by comparison with the mass spectra of the NIST14 and NIST14s libraries. The quantification method was carried out by integrating the area under the curve, represented as a percentage of the total area (% Area).

Analysis of the fresh sample and essential oil Instrumentation

The SPME fibers used were polydimethylsiloxane (PDMS, 100 μ m), polyacrylate (85 μ m), polydimethylsiloxanedivinylbenzene PDMS/DVB 65, divinylbenzene-carboxenpolydimethylsiloxane (DVB/CAR/PDMS 50/30 μ m) from Supelco[®] Bulletin 925F-SPME (Sigma-aldrich 2009). The fibers were conditioned according to the manufacturer's instructions. GC-MS analyses were performed on Shimadzu GC2010 Plus + QP2020 equipment, with an SH-Rxi-5Sil MS column (30 m x 0.25 mm x 0.25 μ m).

Compounds were desorbed through the injection port with helium as carrier gas (Grade 5.0), at a linear velocity of 35.0 cm s⁻¹. The run starts with a temperature of 40 °C (2 min), increasing 3 °C min⁻¹, up to 90 °C (3 min), and then increasing 3 °C min⁻¹, up to 150 °C (5 min), and finally, there is a heating rate of 10 °C min⁻¹, up to 280 °C (1 min) for a total run time of 60.67 min. Mass spectra were obtained by electron impact (EI) at 70 eV. The temperatures of the ionization source and the transfer line were 220 and 250 °C, respectively. The qualitative tentative analysis was performed by comparison with the mass spectra of the NIST14 and NIST14s libraries. The quantification method was carried out by integrating the area under the curve, represented as a percentage of the total area (% Area).

Statistical analysis

A descriptive analysis of the variables 'type of onion' and 'type of compound' was performed using the IBM® SPSS® Statistics 25 software, both on a nominal scale. The qualitative data was presented through a contingency table, indicating the attributes for each variable, type of onion (Biónica, Pereirana, Veleña, and Veleña Sonsón), and type of compound (present in the chromatographic analysis).

A simple correspondence analysis was performed to jointly study these two qualitative variables. It was considered appropriate for dealing with contingency tables in which the existing cases are represented in the categories of each of the qualitative variables under study, i.e., type of onion and type of compound. The association test used was the chi-square test of independence (χ^2), which provided information on the relationship that could exist between the two variables (López 2004).

RESULTS AND ANALYSIS

Prior studies have similarly characterized volatile organic compounds in onions, utilizing multivariate statistical techniques, including cluster analysis, principal component analysis, and discriminant analysis, to identify compound groups associated with specific onion varieties (Taglienti et al. 2021; Cozzolino et al. 2021). Volatile organic compounds in onions are responsible for their distinct sensory attributes and global culinary popularity. However, these compounds

vary with onion variety, underscoring the importance of their comprehensive characterization (Bello et al. 2013).

This study aimed to characterize four onion varieties in Pereira, Colombia, by employing chromatographic analysis to detect volatile compounds and conducting statistical analyses to correlate compound types with onion varieties.

Analysis of fibers in fresh onion variety Pereirana

Using DVB/CAR/PDMS, PDMS, PDMS/DVB, and polyacrylate fibers, the Pereirana variety was assessed to determine which fiber was the most optimal for the analysis. In the process, it was possible to observe 23 compounds according to their mass spectra of the NIST14 and NIST14s libraries. The VOCs profile determined by the four fibers can be observed in Table 1.

Table 1. Compounds extracted from the Pereirana variety onion through the four fibers evaluated.

Code	Compounds	Reference ions	t _{R (min)}
	Monosulphide		
F1	Propyl mercaptan	76, 47	2.19
	Disulfides		
F2	Dipropyl disulfide	43, 150	18.53
F3	Disulfide methyl propyl	60, 122	10.19
F4	(E)-1-(Prop-1-en-1-yl)-2-propyldisulfane	41, 148	18.92
F5	Disulfide methyl 1-(1-propenylthio) propyl	115, 81	42.13
F6	1-Methyl-2-(1-(propylthio) propyl) disulfane	117, 75	41.53
F7	Diisopropyl disulfide	43, 150	19.05
F8	1-Allyl-2-isopropyldisulfane	41, 43	17.82
	Trisulphides		
F9	Trisulfide dipropyl	43, 75	30.76
F10	(E)-1-(Prop-1-en-1-yl)-3-propyltrisulfane	41, 148	18.90
F11	(Z)-1-(Prop-1-en-1-yl)-3-propyltrisulfane	74, 41	31.09
F12	1-Allyl-3-propyltrisulfane	73, 116	31.25
	Tetrasulphide		
F13	6-Ethyl-4,5,7,8-tetrathianonane	73, 149	50.46
	Other sulphurised compounds		
F14	2,4-Dimethyl-5,6-dithia-2,7-nonadienal	69, 129	50.04
F15	Ethanethioic acid, S-propyl ester	43, 74	7.71
	Other		
F16	Cyclopentaneacetic acid, 3-oxo-2-pentyl-, methyl ester	83, 83	45.43
F17	7-Acetyl-6-ethyl-1,1,4,4-tetramethyltetralin	243, 213	51.62
F18	Brassilato de etileno	55, 98	54.25
F19	Tetracosa-2,6,10,14,18-pentaen-22-ol, 2,6,10,15,19,23-hexamethyl-23-	73, 69	56.30
F20	methoxy-, alltran 1,3-Benzodioxole, 5-propyl	135, 77	41.34
F21	Diethyl Phthalate	149, 177	42.35
F22	Ethylene brassylate	83, 82	45.33
F23	1,4-Benzenediol, 2-[(1,4,4a,5,6,7,8,8a-octahydro-2,5,5,8a-tetramethyl- 1-naphthalenyl)methyl]-, [1R-(1.alpha.,4a.beta.,8a.alpha	191, 119	45.67

Among the 23 compounds observed, 65.22% had the characteristic of being sulfurous, namely: F1 (1 monosulphide); F2 to F8 (7 disulfides); F9 to F12 (4 trisulphides); F13 (1 tetrasulphide); and F13 to F14 (other sulphurised compounds). The other eight compounds belonged to aromatic groups or esters. Disulphides and trisulphides were the most observed compounds in the four fibers with retention times of less than 30 min.

The observed compounds were grouped in Table 2, where it is determined that the most indicated fiber to

perform the analysis of the various varieties of Welsh onion was PDMS, since it had the greatest affinity with most of the compounds present in the onion variety Pereirana, observing 14 mostly sulfurous compounds. On the other hand, with DVB/CAR/PDMS, PDMS/DVB, and polyacrylate fibers, it was possible to observe nine, five, and eight compounds, respectively.

The compounds observed in most fibers were propyl mercaptan, dipropyl disulfide, methyl propyl disulfide, and (E)-1-(Prop-1-en-1-yl)-2-propyldisulfane (Figure 1).

 Fibers
 F1
 F2
 F3
 F4
 F5
 F6
 F7
 F8
 F9
 F10
 F11
 F12
 F13
 F14
 F15
 F16
 F17
 F18
 F19
 F20
 F21
 F22
 F23

 DVB/CAR/PDMS
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I
 I

Table 2. Compounds extracted from the Pereirana Variety Welsh Onion using the four evaluated fibers.

_^s_s^

∕_∕^S`s^

methyl propyl disulfide

HS propyl mercaptan

S-S-S

dipropyl disulfide

(E)-l-(Prop-l-en-l-yl)-2-propyldisulfane

Figure 1. Main compounds observed in the four fibers.

Table 2 shows that compounds F2 (Dipropyl disulfide) and F4 ([E]-1-[Prop-1-en-1-yl]-2-propyldisulfane) were the only ones found in the four assessed fibers. Both are aliphatic sulfur compounds with a very similar molar mass. These compounds, together with F1 (Propyl mercaptan) and F6 (1-Methyl-2-[1-(propylthio)propyl]disulfane) were detected in a study of *Allium ascolanicum* L. as the most abundant (Indrasari et al. 2021).

PDMS fiber is preferred for the extraction of non-polar analytes, whereas, on the other hand, polyacrylate fiber is preferred for more polar compounds. PDMS/DVB or CAR/DVB fibers can be used for the extraction of polar VOCs with low molecular weight and, in addition, for obtaining increased retention capacity. Although the stationary phase is fundamental, factors such as the concentration of the analytes in the sample, temperature, pH, agitation time, and addition of salts should also be considered (Garcia-Esteban et al. 2004).

Analysis of the extraction results with PDMS fiber in the four varieties of Welsh onion

The four varieties of Welsh onion were analyzed by the PDMS fiber. The main components observed, according

to their mass spectra from the NIST14 and NIST14s libraries, are grouped in Table 3, in which 29 compounds were found in the PDMS fiber, mostly sulfurous.

It is possible to observe one monosulphide, nine disulfides, five trisulphides, one tetrasulphide, three sulfur compounds,

and 10 different compounds such as ketones or ester derivatives. Disulphides and trisulphides were the most abundant. A descriptive analysis of the bar graph (Figure 2) allowed observation that the onion variety Biónica had the highest number of VOCs, followed by Pereirana and Veleña Sonsón. The variety Veleña had the lowest VOC content.

Table 3. VOCs observed in the PDMS fiber in the different varieties assessed by GCMS HS-SPME.

Code	Compounds	t _{R (min)}	Compound Area (%)
	Monosulphide		
C18	Propyl mercaptan	2.19	6.61
	Disulfides		
C1	Dipropyl disulfide	19.05	63.56
C2	(E)-1-(Prop-1-en-1-yl)-2-propyldisulfane	19.19	6.07
C3	Disulfide, methyl 1-(methylthio)propyl	24.31	0.56
C7	1-Methyl-2-(1-(propylthio)propyl)disulfane	41.53	0.15
C8	Disulfuro, metil 1-(1-propeniltio)propil	42.13	1.23
C19	Disulfide methyl propyl	10.18	0.90
C21	1-(1-Propenyl)-2-(4-thiohept-5-yl)disulfide	41.52	0.15
C20	Diisopropyl disulfide	19.05	1.95
C25	1-Allyl-2-isopropyldisulfane	17.90	0.34
	Trisulphides		
C4	Trisulfide dipropyl	30.91	12.13
C5	(E)-1-(Prop-1-en-1-yl)-3-propyltrisulfane	31.47	0.61
C26	(Z)-1-(Prop-1-en-1-yl)-3-propyltrisulfane	31.09	0.13
C27	1-Allyl-3-propyltrisulfane	31.25	0.23
C28	1,2,4-Trithiolane, 3,5-diethyl-	31.47	0.61
	Tetrasulphide		
C11	6-Ethyl-4,5,7,8-tetrathianonane	40.46	0.47
	Other sulphurised compounds		
C10	2,4-Dimethyl-5,6-dithia-2,7-nonadienal	50.04	0.91
C17	2,3-Bis(ethylthio)hexane	23.24	0.42
C29	Ethanethioic acid S-propyl ester	7.71	0.34
	Other		
C9	3(2H)-Furanone, 5-methyl-2-octyl-	45.20	0.37
C6	2-Tridecanone	38.67	1.21
C12	Morpholine, 4-octadecyl	50.57	0.12
C13	1-Heptadecanamine, N, N-dimethyl-	51.52	0.09
C14	1,6,10-Dodecatriene-3-carboxylic acid, methyl ester	54.70	0.21
C15	9-Octadecenoic acid, methyl ester, (E)-	54.80	0.18
C16	9,12-Octadecadienoic acid (Z, Z)-, methyl ester	55.14	0.15
C22	3(2H)-Furanone, 2-hexyl-5-methyl-	45.21	0.05
C23	Squalene	56.25	0.17
C24	Heneicosane	58.11	0.08

Compounds such as dipropyl disulfide (E)-1-(Prop-1-en-1-il)-2-propyldisulfane, methyl-1-(1-propenylthio) propyl disulfide, dipropyl trisulphide, and (E)-1-(Prop-1-en-1-il)-3-propyltrisulfan were present in the four varieties, with an area percentage for the Pereirana onion of 65.67, 6.28, 1.08, 12.53, and 0.63%, respectively. This result is in line with those corresponding to a previous study (Bastaki et al. 2021), according to which the active compounds of the genus were



Figure 2. Bar graph of the 29 compounds obtained from the four varieties of Welsh onion.

Allium disulfide of diallyl disulfide, diallyl trisulphide, diallyl sulfide, dipropyl disulfide, dipropyl trisulphide, 1-propenyl propyl disulfide, allyl methyl disulfide, and dimethyl disulfide that could have antioxidant and antimicrobial properties.

The Pereirana onion was differentiated by containing 1-Allyl-3-propyltrisulfane, (Z)-1-(Prop-1-en-1-yl)-3propyltrisulfane, 1-Allyl-2-isopropyldisulfane, ethanethioic acid S-propyl ester, and organo-sulfur compounds, whereas the other varieties differed in having ketones and ester derivatives. Figure 3 presents a chromatogram for the PDMS fiber, whose most abundant peak with t_R of 19.05 min corresponded to dipropyl disulfide, which in the Pereirana onion had 20 times the relative abundance concerning the other varieties. This way, it was necessary to dilute the extract since it saturated the mass detector. The compound had reference ions of 43 and 150 atomic mass units (mass spectrum illustrated in Figure 3).



Figure 3. PDMS chromatogram of the Pereirana onion and the respective dipropyl disulfide ions (19.048 t_p).

10618

The compounds identified in this study are comparable to those found in the Welsh onion *A. Fistulosum*, as reported by Choi (2019) using the HS-SPME/GC-MS technique. The predominant compounds identified by the author included (Z)-1-propenyl propyl sulfide, (Z)-propenyl propyl disulfide, (E)-propenyl propyl disulfide, dipropyl trisulfide, and propyl propanthiosulfonate.

Subsequently, a correspondence analysis was performed to describe the relationship or independence between the two nominal variables under study—type of onion and type of compound—and determine the relationships between categories, defining similarities or dissimilarities

Table 4. Data obtained from the chi-square test (χ^2).

between the two variables. This procedure allowed a grouping described by the relationship between the categories of the variables that were consistent. To that end, the chi-square test (χ^2) was performed, the results of which are illustrated in Table 4.

Based on the results presented in Table 4, it can be observed that the significance obtained through the chi-square statistic (χ^2) was 0.000, which rejects the null hypothesis of independence and accepts that there was a relationship between the two variables. Below is Figure 4 in which the four varieties of onion are observed and two varieties that are closely related are Veleña and Veleña Sonsón.

	Value	df	Asymp. Sig (2- sided)
Pearson's chi-squared	149.58ª	84	0.000
Likelihood ratio	166.33	84	0.000
Linear association	0.146	1	0.702
No valid cases	183	-	-

a116 cells (100% expected a count less than five. The minimum expected count was 0.49).



Figure 4. Graph for the variable 'type of onion'.

The associated compounds are illustrated in a column points chart (Figure 5) for a type of compound. It can be observed that the compounds C25 and C26 were di and trisulphides with a molar mass between 148 and 180 g mol⁻¹, from which 1-Allyl-2-isopropyldisulfane can be

highlighted. This compound has been found in onion oil together with dipropyl disulfide and dipropyl trisulphide; however, the concentrations decreased as the temperature increased, which made the aroma change significantly (Tian et al. 2021).



Figure 5. Column points chart for the type of compound.

Associated compounds C6, C9, C15, and C16 were observed, of which the first two had the characteristics of being methyl ketones. Methyl ketones include 2-undecanone and 2-tridecanone, which are important in the flavor and fragrance industry. These compounds have a variety of natural and commercial functions, including pheromones and natural insecticides in plants. Depending on their concentrations, they can generate a protective effect against insect attacks, which may indicate that the varieties Biónica, Veleña, and Veleña Sonsón had some protection against insects (Antonious 2013).

Compounds C1, C3, C4, and C19 gave rise to another group, and compounds C1, C4, and C19 have been reported in previous studies by HS-GC-MS (D'Auria and Racioppi 2017), for example, the Dipropyldisulphide was reported in 51.41% for *Allium cepa* L. and 58.57% for *Allium ascalonicum* L.

The presence of this group of compounds could be used for soil biofumigation and plant growth stimulation since they can control pathogens and increase fruit productivity by 15 to 20%. This is a potential biofumigant that could be used to replace methyl bromide, a substance that is characterized by destroying the ozone layer (Arnault et al. 2013).

In studies on essential oils of onions, *Allium porrum* L. (*Alliaceae*) was characterized by the presence of dipropyl disulfide, dipropyl trisulphide, and dipropyl tetrasulphide. The results obtained suggest that the presence of the allyl group was essential for the antimicrobial activity of these sulfur derivatives when they are present in *Allium* or other species (Casella et al. 2013).

Compounds such as diallyl disulfide, diallyl trisulphide, diallyl sulfide, dipropyl disulfide, dipropyl trisulphide, 1-propenylpropyl disulfide, allyl methyl disulfide, and dimethyl disulfide correspond to the most active compounds of the *Alliaceae* family, which have been proven to have medicinal properties such as antibacterial and antioxidant activities thanks to the organosulfur compounds, which are believed to prevent the development of cancer, cardiovascular and neurological diseases, diabetes, liver diseases, as well as allergies and arthritis (Bastaki et al. 2021). These organo-sulfur

10620

compounds have demonstrated their versatility in various applications. Patel et al. (2018) explored their potential in promoting hair growth when incorporated into a shampoo formulation. Additionally, Sharma et al. (2018) highlighted their effectiveness as antibacterial agents, while Hannan et al. (2010) investigated their role as bacterial inhibitors in food products. Furthermore, Khater et al. (2009) suggested their suitability as a potential insect repellent. These findings underscore

the multifaceted utility of organo-sulfur compounds in diverse fields.

Through the correspondence analysis, it was possible to show that there is a significant relationship between the variable "onion variety" and the variable "type of compound" and in Figure 6 it can be seen for each variety of onion the compounds that are associated with it, and which therefore characterize it.



Figure 6. Bispace graph: Relationship between the type of compound and the variety of onion.

Study of the essential oil of Pereirana onion

A fluid liquid oil was obtained with a characteristic odor of onion, a light-yellow color, distinctive properties of essential oils (Rojas et al. 2009), and a yield of 0.028%, which was above that reported by other authors 0.0053% (EI-Tohamy et al. 2009), 0.0045% (Yassen and Khalid 2009; Pino et al. 2000) and lower than that reported by Dron et al. (1997), which obtained 0.032% in extraction by supercritical fluids. Table 5 shows the compounds observed with the PDMS and polyacrylate fibers, which were the ones that extracted the greatest number of compounds. The result is presented by comparison with the NIST14 and NIST14s libraries. The compounds with more than 85% coincidence with the spectrum of the library are presented. The relative areas for each compound are also presented concerning the total automatically integrated.

The main compound of the essential oil of Welsh onion was dipropyl disulfide (number 31, with 15.38%). Studies presented by Cantrell et al. (2020) for onion Cepa oil extracted with dichloromethane (DCM) managed to extract up to 27 organosulfur compounds, including dipropyl disulfide in great abundance. According to Gao et al. (2019) who used the SPME fiber with phase 50:30 µm DVB/

CAR on PDMS, sulfocompounds are the main functional compositions of onion and its oils, exhibiting significant bioactivity in human health. According to this experiment,

volatile sulfocompounds, such as disulfide, trisulfide, tetrasulfide, thiophene, and dithiane with dimethyl groups, were the main components of onion oils.

Table 5. Chemical Composition of the Essential Oil from the Pereirana Variety Onion.

No	Compounds	CAS	t _R (min)	SI (%)	PDMS Area (%)	Polyacrylate Area (%)
1	Propyl acetate	109-60-4	3.89	95	0.51	0.25
2	1,1-Dietoxietano	105-57-7	4.19	96	0.33	0.13
3	Sec-butyl acetate	105-46-4	4.88	93	0.10	-
4	Isobutyl acetate	110-19-0	5.28	98	1.57	0.82
5	Ethyl butyrate	105-54-4	6.08	93	0.12	-
6	n-Butilo acetato	123-86-4	6.49	96	-	0.25
7	Ethyl 2-methyl butyrato	7452-79-1	7.76	93	0,09	0.04
8	3-Hexen-1-ol, (Z)	928-96-1	7.98	86	-	0.02
9	3-Hexen-1-ol, (E)	928-97-2	8.42	85	-	0.05
10	Amyl acetate	628-63-7	8.82	93	1.24	-
11	Isoamyl acetate	123-92-2	8.83	95	1.07	0.52
12	m-Xylene	108-38-3	9,40	91	-	0.05
13	Cyclopentane, 1-ethenyl-3-methylene-	61142-07-02	9.40	93	0.11	-
14	3,4- dimethyl thyophene	632-15-5	9,86	92	0.39	0.27
15	Isopropyl methyl disulfide	40136-65-0	11.09	93	0.36	0.22
16	α-Pineno	80-56-8	11.23	92	0.14	-
17	(Z)-Methyl-1-propenyl disulfide	23838-18-8	11.41	88	0.3	0.11
18	Dimethyl trisulfide	3658-80-8	12.76	87	-	0.03
19	1-Heptanol	111-70-6	13.03	88	-	0.05
20	β-Myrcene	123-35-3	13.91	90	0.09	0.03
21	Decane	124-18-5	14.47	90	0.05	-
22	Octanal	124-13-0	14.56	94	0.09	0.07
23	Limonene	138-86-3	15.79	92	0.13	0.05
24	4-methyl decane	2847-72-5	17.41	93	0.05	-
25	2-methyl decane	6975-98-0	17.62	92	0.05	-
26	2-propyl-1-Heptanol	10042-59-8	17.91	93	0.10	-
27	1-Octanol	111-87-5	17.92	94	-	0.10
28	(E)-1-Allyl-2-(prop-1-in-1-yl) disulfide	122156-02-9	19.25	89	0.10	0.06
29	Tridecane	629-50-5	19.49	95	-	0.34
30	Undecane	1120-21-4	19.54	95	1.66	-
31	Dipropyl disulfide	629-19-6	19.87	96	15.38	6.08
32	1-Allyl-2-isopropyl disulfate	67421-85-6	20.19	91	2.65	1.24
33	Metil propil trisulfuro	17619-36-2	22.16	91	0.45	0.28
34	(E)-1-Metil-3-(prop-1-en-1-il) trisulfan	23838-25-7	22.69	87	0.07	0,03
35	3-methyl-1-hexanol	13231-81-7	23.68	90	0.25	-
36	Nonyl acetate	143-13-5	23.73	86		0.39

Table 5

No	Compounds	CAS	t _R (min)	SI (%)	PDMS Area (%)	Polyacrylate Area (%)
37	Methyl 2-oxo nonanoate	56275-54-8	24.42	91	0.16	0.11
38	Dodecane	112-40-3	25.47	93	0.12	-
39	Decanal	112-31-2	25.72	94	0.32	0.15
40	Cyclohexyl Isothiocyanate	1122-82-3	27.00	95	0.43	0.30
41	3-Ethyl-5-Methyl-1,2,4-Trithiolane	116505-59-0	27,13	86	0.39	0.24
42	3,3,8-trimethyl decane	62338-16-3	29.60	88	0.05	-
43	2-Undecanone	0112-12-9	30.73	94	12.16	5.63
44	2-Undecanol	1653-30-1	31.12	96	0.78	0.56
45	2-Methoxy-4-vinylphenol	7786-61-0	31.31	85	-	0.12
46	2,4-Decadienal	25152-84-5	31.72	87	-	0.04
47	Dipropyl trisulfide	6028-61-1	32.14	96	2.15	1.35
48	(E)-1-(Prop-1-in-1-yl)-3-propyltrisulfan	23838-27-9	32.54	90	4.19	3.45
49	2,4-Octanediona	14090-87-0	34.58	92	-	1.79
50	4-Methyl dodecane	6117-97-1	35.76	92	0.12	-
51	Methyl 1-(methyl thio) propyl disulfide	53897-66-8	36.47	89	0.16	0.09
52	2-nonyl acetate	14936-66-4	36.65	85	0.18	0.26
53	2-hexyl-5-methyl-3(2H)-furanon	33922-66-6	37.37	93	1.42	0.99
54	4,6-dimethyl dodecane	61141-72-8	39.60	93	0.14	-
55	2-Tridecanone	0593-08-08	39.96	94	6.82	3.61
56	3,7- dimethyl decane	17312-54-8	40.14	92	0.16	0.04
57	2-Tridecanol	1653-31-2	40.24	93	-	0.15
58	Dipropyl tetrasulfide	52687-98-6	42.67	86	0.20	0.13
59	1-Methyl-2-(1-(propyltio)propyl)disulfide	126876-21-9	42.82	85	0.37	0.14
60	(Z)-3-hexen-1-yl benzoate	25152-85-6	42.96	94	0.17	0.11
61	trans-3,6-diethyl-1,2,4,5-tetratian	934273-77-5	43.89	90	1.99	1.06
62	cis-4,6-diethyl-1,2,3,5-tetratian	137363-91-8	44.04	88	1.92	1.26
63	Hexadecane	544-76-3	44.55	93	0.20	-
64	5-methyl-2-octyl-3(2H)-furanone	57877-72-2	46.83	95	0.63	0.45
65	Dioctyl ether	629-82-3	47.80	86	0.06	0,04
66	2-Nonadecanone	629-66-3	49.02	86	0.06	0.04
67	Pentadecanal	2765-11-9	49.55	86	0.03	-
68	Benzyl benzoate	120-51-4	50.75	90	0.04	0.05
69	1-(1-propenylthio)propyl propyl disulfide	143193-11-7	51.64	87	0.08	-
70	4,7-Diethyl-1,2,3,5,6-pentatiopane	878997-71-8	52.83	89	0.21	-

Area (%)=relative percentage of the compound in the essential oil; SI (%)=percentage of coincidence with the NIST14 and NIST14s libraries.

Essential oils are concentrated and contain more than twice as many components as fresh samples. They can be used to confer the aroma and flavor of onions without the difficulties of handling large quantities of fresh products. It is plausible to anticipate that the acquisition of new knowledge about native species will yield numerous benefits for humanity, offering new services and products, as well as raw materials for various industries (Lead et al. 2005). In this context, the volatile extracts of the Pereira onion are identified as a raw material of interest due to their high concentration of sulfur compounds. These compounds have been attributed with properties useful for insecticide applications (Denloye 2010), and in the pharmaceutical industry as promising anti-allergy, antihistamine, anti-inflammatory, antioxidant agents, and as preventatives for brain edema with neuroprotective potential (Kuete 2017).

Onion extracts can also be utilized in the cosmetic industry due to their propensity to induce tissue regeneration and facilitate wound healing (Celano et al. 2021). Other potential applications include use as an oil oxidation stabilizer, an ingredient in bakery products, pasta, and noodles, a quality improver for meat products, a substrate for enzyme production, and as a raw material for tea. However, extensive research is necessary before these potential applications can be commercialized or scaled up at an industrial level (Kumar et al. 2022).

CONCLUSIONS

The results underscore the efficacy of the HS-SPME/GC-MS method for the analysis of volatile compounds in the essential oil of the Pereirana variety Welsh onion. This method enabled the separation and characterization of up to 70 compounds using four solid-phase microextraction fibers. Notably, the polyacrylate fiber demonstrated a significant affinity for sulfur compounds, with dipropyl disulfide being the most abundant, constituting 65.67% of the total.

The study revealed that the concentration of volatile compounds in the Pereirana variety of Welsh onion exceeds that of other varieties by up to 20 times. As a result, this variety emerges as a promising plant material warranting further research. Its potential applications span across various industries, encompassing food, pharmaceutical, agricultural, and cosmetics.

ACKNOWLEDGMENTS

The authors are thankful to the Universidad Tecnológica de Pereira for financing the research and publication of the present study through the project SI9-22-2.

REFERENCES

Antonious GF (2013) 2-Undecanone and 2-tridecanone in fieldgrown onion. Journal of Environmental Science and Health Part B 48(4): 302-307. https://doi.org/10.1080/03601234.2013.743790 Arnault I, Fleurance C, Vey F, Du Fretay G and Auger J (2013) Use of Alliaceae residues to control soil-borne pathogens. Industrial Crops and Products 49: 265-272. https://doi.org/10.1016/j.indcrop.2013.05.007

Bastaki SM, Ojha S, Kalasz H and Adeghate E (2021) Chemical constituents and medicinal properties of *Allium* species. Molecular and Cellular Biochemistry 476(12): 4301-4321. https://doi.org/10.1007/s11010-021-04213-2

Bello MO, Olabanji IO, Abdul-Hammed M and Okunade TD (2013) Characterization of domestic onion wastes and bulb (*Allium cepa* L.): fatty acids and metal contents. International Food Research Journal 20(5): 2153-2158. https://www.researchgate.net/publication/258223181

Boutekedjiret C, Bentahar F, Belabbes R and Bessiere JM (2003) Extraction of rosemary essential oil by steam distillation and hydrodistillation. Flavour and Fragrance Journal 18(6): 481-484. https://doi.org/10.1002/ffj.1226

Cantrell MS, Seale JT, Arispe SA and McDougal OM (2020) Determination of organosulfides from onion oil. Foods 9(7): 884. https:// doi.org/10.3390/foods9070884

Casella S, Leonardi M, Melai B, Fratini F and Pistelli L (2013) The role of diallyl sulfides and dipropyl sulfides in the *in vitro* antimicrobial activity of the essential oil of garlic, *Allium sativum* L., and leek, *Allium porrum* L. Phytotherapy Research 27(3): 380-383. https://doi. org/10.1002/ptr.4725

Celano R, Docimo T, Piccinelli AL, Gazzerro P, Tucci M et al (2021) Onion peel: Turning a food waste into a resource. Antioxidants 10(2): 304. https://doi.org/10.3390/antiox10020304

Colina-Coca C, González-Peña D, Vega E, de Ancos B and Sánchez-Moreno C (2013) Novel approach for the determination of volatile compounds in processed onion by headspace gas chromatographymass spectrometry (HS GC–MS). Talanta 103: 137-144. https://doi. org/10.1016/j.talanta.2012.10.022

Cozzolino R, Malorni L, Martignetti A, Picariello G et al (2021) Comparative analysis of volatile profiles and phenolic compounds of Four Southern Italian onion (*Allium cepa* L.) Landraces. Journal of Food Composition and Analysis 101: 103990. https://doi.org/10.1016/j. jfca.2021.103990

Choi J (2019) Changes in volatile compounds in Welsh onion (*Allium fistulosum* L.) concentrates processed by reverse osmosis during storage. https://dspace.ewha.ac.kr/handle/2015.oak/262589

DANE (2019) Encuesta nacional agropecuaria (ENA) En: DANE, https://www.dane.gov.co/index.php/estadisticas-por-tema/agropecuario/ encuesta-nacional-agropecuaria-ena. 17 p.; consultado: Octubre 2023.

D'Auria M and Racioppi R (2017) HS-SPME-GC-MS Analysis of onion (*Allium cepa* L.) and shallot (*Allium ascalonicum* L.). Food Research 1(5): 161-165. http://doi.org/10.26656/fr.2017.5.055

Denloye AA (2010) Bioactivity of Powder and Extracts from Garlic, *Allium sativum* L. (Alliaceae) and Spring onion, *Allium fistulosum* L. (Alliaceae) against *Callosobruchus maculatus* F. (Coleoptera: Bruchidae) on Cowpea, *Vigna unguiculata* (L.) Walp (Leguminosae) Seeds. Psyche: A Journal of Entomology 2010: 1-5. https://doi.org/10.1155/2010/958348

Dron A, Guyeru DE, Gage DA and Lira CT (1997) Yield and quality of onion flavor oil obtained by supercritical fluid extraction and other methods. Journal of Food Process Engineering 20(2): 107-124. https://doi.org/10.1111/j.1745-4530.1997.tb00414.x

Garcia-Esteban M, Ansorena D, Astiasarán I and Ruiz J (2004) Study of the effect of different fiber coatings and extraction conditions on dry cured ham volatile compounds extracted by solidphase microextraction (SPME). Talanta 64(2): 458-466. https://doi. org/10.1016/j.talanta.2004.03.007

Gao Y, Wu S, Sun Y, Cong R, Xiao J and Ma F (2019) Effect of freeze dried, hot air dried and fresh onions on the composition of volatile sulfocompounds in onion oils. Drying Technology 37(11): 1427-1440. https://doi.org/10.1080/07373937.2018.1504062

Hannan A, Humayun T, Hussain MB, Yasir M and Sikandar S (2010) *In vitro* antibacterial activity of onion (*Allium cepa*) against clinical isolates of *Vibrio cholerae*. Journal of Ayub Medical College Abbottabad 22(2): 160-163. https://www.ayubmed.edu.pk/JAMC/PAST/22-2/Hannan.pdf

Hosoda H, Ohmi K, Sakaue K and Tanaka K (2003) Inhibitory effect of onion oil on browning of shredded lettuce and its active components. Journal of the Japanese Society for Horticultural Science 72: 451–456. http://doi.org/10.2503/jjshs.72.451

Hori M (2007) Onion aphid (*Neotoxoptera formosana*) attractants, in the headspace of *Allium fistulosum* and *A. tuberosum* leaves. Journal of Applied Entomology 131(1): 8-12. https://doi.org/10.1111/j.1439-0418.2006.01130.x

Indrasari SD, Arofah D, Kristamtini, Sudarmaji and Handoko DD (2021) Volatile compounds profile of some Indonesian shallot varieties. In IOP Conference Series: Earth and Environmental Science 746(1): 012009. IOP Publishing. https://doi.org/10.1088/1755-1315/746/1/012009

Jaramillo O and Vallejo A (2014) História rural de Pereira. Paisaje Cultural Cafetero. Academia Pereirana de Historia, Pereira, Colombia: Instituto municipal de cultura y fomento al turismo.

Khater HF, Ramadan MY and El-Madawy RS (2009) Lousicidal, ovicidal and repellent efficacy of some essential oils against lice and flies infesting water buffaloes in Egypt. Veterinary Parasitology 164(2-4): 257-266. https://doi.org/10.1016/j.vetpar.2009.06.011

Kuete V (2017) Chapter 14 - Allium cepa. In Medicinal spices and vegetables from Africa (pp. 353-361). Academic Press. https://doi. org/10.1016/B978-0-12-809286-6.00014-5

Kumar M, Barbhai MD, Hasan M, Punia S, Dhumal S et al (2022) Onion (*Allium cepa* L.) peels: A review on bioactive compounds and biomedical activities. Biomedicine & Pharmacotherapy 146: 112498. https://doi.org/10.1016/j.biopha.2021.112498

Kumar M, Barbhai MD, Hasan M, Dhumal S, Singh S et al (2022) Onion (*Allium cepa* L.) peel: A review on the extraction of bioactive compounds, its antioxidant potential, and its application as a functional food ingredient. Journal of Food Science 87(10): 4289-4311. https://doi. org/10.1111/1750-3841.16297

Lawless J (2012) Encyclopedia of essential oils: The complete guide to the use of aromatic oils in aromatherapy, herbalism, health and well-being. HarperCollins Publishers,

Lead C, Beattie AJ, Barthlott W and Rosenthal J (2005) New products and industries from biodiversity. Ecosystems and human well-being: Current State and Trends: Findings of the Condition and Trends Working Group 1: 271.

Licen S, Muzic E, Briguglio S, Tolloi A, Barbieri P and Giungato P (2021) Derivatized volatile organic compound characterization of Friulano wine from Collio (Italy–Slovenia) by HS-SPME-GC-MS and discrimination from other varieties by chemometrics. British Food Journal 123(8): 2844-2855. https://doi.org/10.1108/BFJ-08-2020-0690

López CP (2004) Técnicas de análisis multivariante de datos. Pearson Education, London, UK.

Patel NR, Mohite SA and Shaha RR (2018) Formulation and

evaluation of onion hair nourishing shampoo. Journal of Drug Delivery and Therapeutics 8(4): 335-337. https://doi.org/10.22270/jddt.v8i4.1810

Peng J, Yang Y, Zhou Y, Hocart CH et al (2020) Headspace solidphase microextraction coupled to gas chromatography-mass spectrometry with selected ion monitoring for the determination of four food flavoring compounds and its application in identifying artificially scented rice. Food Chemistry 313: 126136. https://doi.org/10.1016/j.foodchem.2019.126136

Peredo-Luna HA, Palou-García E and López-Malo A (2009) Aceites esenciales: métodos de extracción. Temas Selectos de Ingeniería de Alimentos 3(1): 24-32.

Pino JA, Rosado A and Fuentes V (2000) Volatile flavor compounds from *Allium fistulosum* L. Journal of Essential Oil Research 12(5): 553-555. https://doi.org/10.1080/10412905.2000.9712157

Polanco MF and Pérez JFB (2018) Caracterización morfoagronomica de seis clones de cebolla rama (*Allium fitulosom* L) cultivados en el municipio de Pereira. RIAA 9(2): 3. https://hemeroteca.unad.edu.co/ index.php/riaa/article/view/2195

Rojas LI JP, Perea A and Stashenko EE (2009) Obtención de aceites esenciales y pectinas a partir de subproductos de jugos cítricos. Vitae 16(1): 110-115.

Sharma K, Mahato N and Lee YR (2018) Systematic study on active compounds as antibacterial and antibiofilm agent in aging onions. Journal of Food and Drug Analysis 26(2): 518-528. https://doi.org/10.1016/j. jfda.2017.06.009

Sigma-aldrich (2009) Bulletin 925F-SPME Applications Guide. Available at: https://www.sigmaaldrich.com/deepweb/assets/sigmaaldrich/ marketing/global/documents/334/578/t199925.pdf. Accessed: 23 September 2022.

Silva JA and Uchida RS (2000) Plant nutrient management in Hawaii's soils: Approaches for Tropical and Subtropical Agriculture. http://hdl.handle.net/10125/1908

Soto VC, Maldonado IB, Jofré VP et al (2015) Direct analysis of nectar and floral volatile organic compounds in hybrid onions by HS-SPME/ GC–MS: Relationship with pollination and seed production. Microchemical Journal 122: 110-118. https://doi.org/10.1016/j.microc.2015.04.017

Taglienti A, Araniti F, Piscopo A and Tiberini A (2021) Characterization of volatile organic compounds in 'Rossa di Tropea' onion by means of headspace solid-phase microextraction gas chromatography–mass spectrometry (HS/SPME GC–MS) and sensory analysis. Agronomy 11: 874. https://doi.org/10.3390/agronomy11050874

Tian P, Zhan P, Tian H, Wang P, Lu C et al (2021) Analysis of volatile compound changes in fried shallot (*Allium cepa* L. var. *aggregatum*) oil at different frying temperatures by GC–MS, OAV, and multivariate analysis. Food Chemistry 345: 128748. https://doi. org/10.1016/j.foodchem.2020.128748

El-Tohamy WA, Khalid AK, El-Abagy HM and Abou-Hussein SD (2009) Essential oil, growth and yield of onion (*Allium cepa* L.) in response to foliar application of some micronutrients. Australian Journal of Basic and Applied Sciences 3(1): 201-205. ISSN 1991-8178

Yassen AA and Khalid KA (2009) Influence of organic fertilizers on the yield, essential oil and mineral content of onion. International Agrophysics 23(2) 183-188.

Zhang ZM and Li GK (2007) A preliminary study of plant aroma profile characteristics by a combination sampling method coupled with GC–MS. Microchemical Journal 86(1): 29-36. https://doi.org/10.1016/j. microc.2006.09.003