

# Comparative Optical Performance of Brain Tissue-Clearing Methods Combined with Immunostaining: A Systematic Review with Descriptive Quantitative Synthesis

Comparación del rendimiento óptico de los métodos de aclaramiento tisular cerebral combinados con inmunotinción: revisión sistemática con síntesis cuantitativa descriptiva

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## Declaration of interests

The authors declare no conflicts of interest.

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## Ethics statement

Ethical approval was not required for this study because it did not involve human participants, animals, or identifiable personal data.

## Abstract

**Introduction.** Optical clearing has enabled three-dimensional visualization of brain tissue, but available methods differ in their ability to preserve morphology, achieve maximum effective imaging depth, and maintain fluorescent signal quality while minimizing unwanted background fluorescence.

**Objective.** To evaluate the optical and morphological performance of tissue-clearing protocols applied to immunostained brain structures through a systematic review with descriptive quantitative synthesis of studies published between 2014 and 2024.

**Method.** Experimental studies using human or rodent brain tissue combining optical clearing and immunostaining were identified through searches in Scopus and Web of Science following the PRISMA 2020 guidelines. Data were synthesized using descriptive statistics, group comparisons, non-parametric correlations, and exploratory regression analyses. Bibliometric network analyses were conducted using the software VOSviewer.

**Results.** Twelve experimental studies met the inclusion criteria. Organic solvent-based methods (iDISCO+, uDISCO) showed higher descriptive values of maximum effective imaging depth and lower levels of unwanted background fluorescence compared to hydrogel-based and hydrophilic approaches. Maximum effective imaging depth was significantly associated with reduced unwanted background fluorescence ( $\rho = -0.596$ ;  $p = 0.041$ ), while its association with fluorescent signal quality showed a non-significant positive trend ( $\rho = 0.570$ ;  $p = 0.053$ ).

#### Data availability

All data supporting the findings of this study are available within the article. For additional details, please contact the corresponding author.

#### Author Contributions

##### Jorge L. Anaya-González:

conceptualization, data curation, formal analysis, investigation, methodology, project administration, resources, software, supervision, validation, visualization, writing – original draft, writing – review & editing.

**Claudia Velásquez-Calderón:** data curation, formal analysis, investigation, validation.

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**Juan José Chingal-Cando:** data curation, formal analysis, investigation, software, validation.

#### Generative AI declaration

During the preparation of this manuscript, the authors used ChatGPT as a generative artificial intelligence tool solely for language editing. The tool was not used to generate scientific content, interpret results, create figures, or perform data analyses. The authors reviewed and edited the manuscript and take full responsibility for the accuracy and integrity of the final content.

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**Conclusion.** Organic solvent-based clearing methods exhibited more favorable descriptive optical profiles, particularly with respect to imaging depth and background suppression. However, in the absence of statistically significant between-group differences, these findings should be interpreted as suggestive patterns rather than definitive evidence of superiority.

#### Keywords

Optical clearing; immunostaining; brain tissue; fluorescence; three-dimensional microscopy; tissue transparency.

#### Resumen

**Introducción.** El aclaramiento óptico ha permitido la visualización tridimensional del tejido cerebral; sin embargo, los métodos disponibles difieren en su capacidad para preservar la morfología, alcanzar mayores profundidades de visualización efectiva y mantener una adecuada calidad de señal fluorescente con mínima fluorescencia de fondo no deseada.

**Objetivo.** Evaluar el rendimiento óptico y morfológico de los protocolos de aclaramiento tisular aplicados a tejido cerebral inmunotintado mediante una revisión sistemática con síntesis cuantitativa descriptiva de estudios publicados entre 2014 y 2024.

**Método.** Se identificaron estudios experimentales en tejido cerebral humano o murino que combinaron aclaramiento tisular e inmunotinción, mediante búsquedas en las bases de datos de Scopus y Web of Science, siguiendo las directrices PRISMA 2020. Se realizaron análisis descriptivos, comparaciones de grupos, correlaciones no paramétricas y modelos de regresión exploratorios. Se incluyó además un análisis bibliométrico con VOSviewer.

**Resultados.** Doce estudios cumplieron los criterios de inclusión. Los métodos basados en solventes orgánicos (iDISCO+, uDISCO) mostraron mayores valores descriptivos de profundidad máxima de visualización efectiva y menores niveles de fluorescencia de fondo no deseada en comparación con los métodos hidrofílicos y basados en hidrogel. La profundidad máxima de visualización efectiva se asoció significativamente con la reducción de fluorescencia de fondo no deseada ( $\rho = -0,596$ ;  $p = 0,041$ ), mientras que su relación con la calidad de la señal fluorescente mostró una tendencia positiva no significativa ( $\rho = 0,570$ ;  $p = 0,053$ ).

**Conclusión.** Los métodos de aclaramiento basados en solventes orgánicos presentaron perfiles ópticos descriptivamente más favorables, especialmente en profundidad de visualización y supresión de fondo. No obstante, estos hallazgos deben interpretarse como patrones sugerentes y no como evidencia concluyente de superioridad.

#### Palabras clave

Aclaramiento óptico; inmunotinción; tejido cerebral; fluorescencia; microscopía tridimensional; transparencia tisular.

## Introduction

Achieving a three-dimensional visualization of brain microarchitecture remains a major technical challenge, with direct implications for clinical and translational neuroscience. Volumetric imaging of immunostained brain tissue enables improved detection of pathological changes associated with neurodegenerative disorders, enhances the characterization of neuronal and glial alterations in neuropathology, and supports circuit-level mapping in experimental disease models. These advances contribute to improved diagnostic interpretation, a better understanding of disease progression, and the development of targeted neurorehabilitation and neurological interventions [1–3]. Consequently, methodological advances in tissue clearing and volumetric immunostaining are highly relevant to health sciences and clinical research.

Conventional histological approaches rely on serial sectioning, which disrupts spatial continuity and introduces distortions in neuronal pathways. In contrast, optical tissue-clearing techniques reduce light scattering through lipid removal and refractive index homogenization, allowing deeper imaging of intact samples and improved visualization of neuronal and glial networks [4–7].

Over the past decade, tissue-clearing technologies have expanded considerably, giving rise to four broadly recognized methodological categories. Hydrogel-based protocols (e.g., CLARITY), are generally reported to preserve protein structure and tissue architecture while enabling moderate maximum effective imaging depth. Hydrophilic or aqueous methods such as CUBIC and SeedDB are often described as offering a balance between optical transparency and volumetric stability. Organic solvent-based approaches (e.g., iDISCO+, uDISCO, PEGASOS) are frequently associated with greater optical penetration, although potential trade-offs such as tissue shrinkage or reduced fluorescence retention have been reported. Hybrid or expansion-based techniques, including ProExM, combine chemical clearing with physical expansion to enable subcellular-resolution imaging. Each category presents distinct advantages and limitations related to optical transparency, structural preservation, and compatibility with immunostaining [5,8–13].

Newer rapid-clearing strategies have also been proposed in recent years; however, these were not represented among the studies meeting the eligibility criteria of the present review and are mentioned here solely for contextual completeness.

Despite these methodological advances, substantial variability persists across studies, particularly in antibody penetration efficiency, fluorescent signal quality, unwanted background fluorescence, and microscopy configurations [7,14–18]. In postmortem human brain tissue, adaptations of clearing and immunostaining protocols have enabled multicolour labelling and visualization of deep cortical structures. Nevertheless, challenges related to autofluorescence, incomplete labelling, and structural distortion remain significant [19–21].

The marked heterogeneity across studies (including differences in antibody selection, clearing chemistry, imaging modalities, and definitions of optical outcomes), limits the applicability of conventional effect-size-based meta-analytic approaches. To address this complexity, the present work applies a descriptive quantitative synthesis, integrating pooled summary statistics, correlation analyses, and exploratory regression models to characterize patterns of optical performance across clearing categories. This approach enables structured comparison while respecting the diversity of experimental designs and measurement frameworks reported in the literature.

Although several studies have reported partial quantitative comparisons of optical clearing performance, these analyses are often limited in scope and are not integrated into a unified synthesis focused specifically on immunostained brain tissue. Key outcome measures, such as maximum effective imaging depth, fluorescent signal quality, unwanted background fluorescence, and morphological preservation are inconsistently reported, hindering the development of evidence-informed guidance for protocol selection [22–25].

To address this gap, the present study conducts a systematic review with descriptive quantitative synthesis of tissue-clearing protocols combined with immunostaining in human and rodent brain samples. Maximum effective imaging depth, fluorescent signal quality, unwanted background fluorescence, and morphological preservation are comparatively examined across hydrogel-based, hydrophilic, solvent-based, and hybrid methods. By integrating data from heterogeneous experimental studies, this work aims to identify descriptive patterns of optical performance associated with different clearing strategies, and to support more informed methodological decision-making in three-dimensional neuroimaging research.

## **Objectives, Research Questions and Hypotheses**

### ***General Objective***

To evaluate the optical and morphological performance of tissue-clearing protocols applied to immunostained brain structures (specifically, maximum effective imaging depth, fluorescent signal quality, unwanted background fluorescence, and morphological preservation) through a systematic review with descriptive quantitative synthesis of studies published between 2014 and 2024.

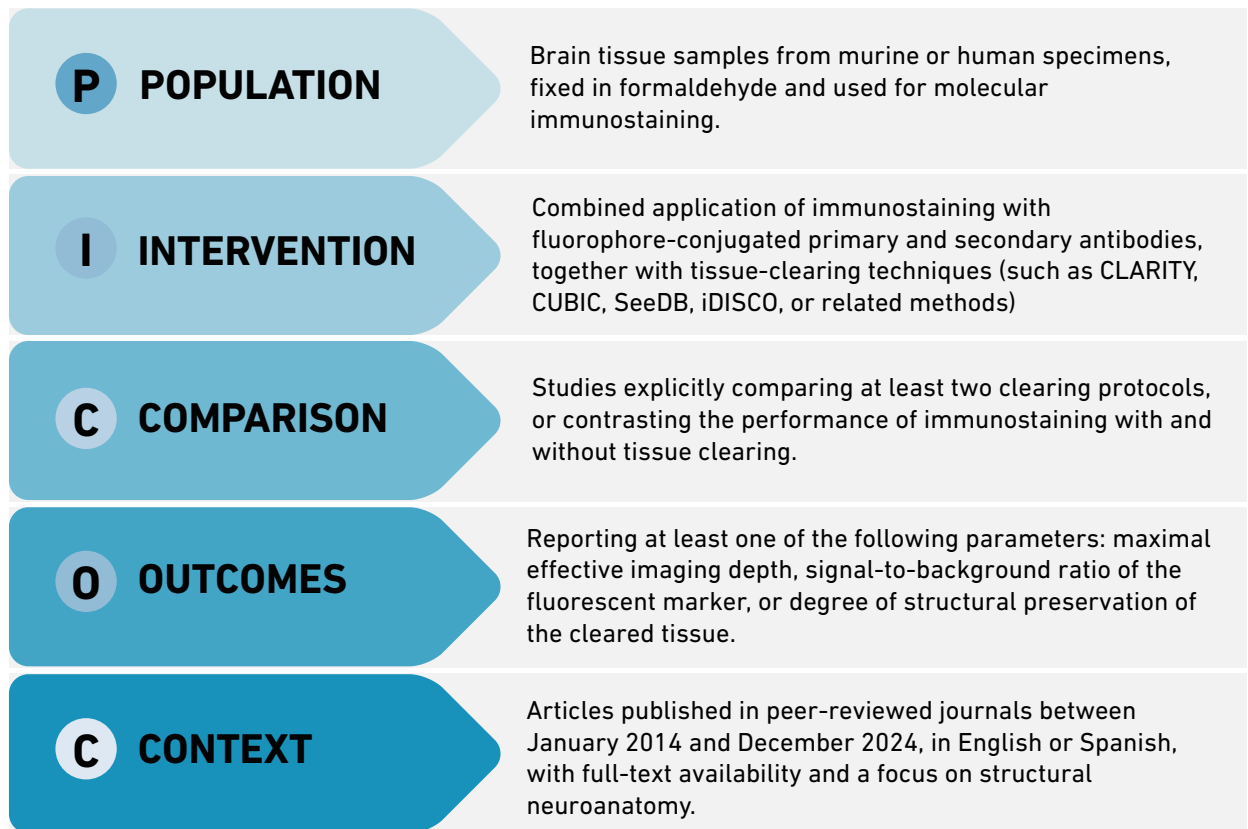
### ***Specific Objectives***

1. To compare maximum effective imaging depth across clearing methods according to chemical class: hydrogel-based, hydrophilic, solvent-based, and hybrid.
2. To evaluate fluorescent signal quality and unwanted background fluorescence in relation to immunostaining conditions, including primary and secondary antibody combinations.
3. To examine morphological preservation, including tissue shrinkage or expansion, and its association with clearing chemistry and optical performance.

### ***Research Questions***

Research questions were structured following the PICOC framework (Population, Intervention, Comparison, Outcome, Context), as outlined in [Figure 1](#).

1. Which clearing chemistry is associated with greater maximum effective imaging depth in immunostained brain tissue?
2. How do different immunostaining conditions influence fluorescent signal quality and unwanted background fluorescence?
3. How does the chemical nature of each clearing method relate to morphological preservation in cleared brain tissue?
4. What associations are observed among maximum effective imaging depth, fluorescent signal quality, unwanted background fluorescence, and morphological preservation across clearing methods?



**Figure 1.** Elements for formulating the research question according to the PICOC framework.

### *Working Hypotheses*

Based on the patterns reported in the literature, the following working hypotheses were formulated:

H1: Organic solvent-based tissue-clearing protocols are expected to be associated with greater maximum effective imaging depth compared with hydrogel-based, hydrophilic, and hybrid methods.

H2: Variations in fluorescent signal quality and unwanted background fluorescence are expected to depend on primary and secondary antibody combinations, with solvent-based methods hypothesized to show higher signal-to-background ratios under comparable conditions.

H3: Hydrogel-based and hydrophilic clearing protocols are expected to demonstrate better morphological preservation than organic solvent-based methods, despite achieving lower optical transparency.

H4: A positive association is hypothesized between maximum effective imaging depth and fluorescent signal quality, whereas an inverse association is anticipated between unwanted background fluorescence and overall optical performance.

## Method

### Study Design

A systematic review with descriptive quantitative synthesis was conducted in accordance with the PRISMA 2020 guidelines (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) [26]. The methodological guidance provided by the *Cochrane Handbook for Systematic Reviews of Interventions* was followed to ensure transparency and reproducibility [27].

The protocol was prospectively registered in the International Prospective Register of Systematic Reviews (PROSPERO; CRD420251160314) [28]. The registration predefined the study objectives, research questions, eligibility criteria, and analytical approach. An administrative update was recorded on October 31, 2025, to clarify eligibility criteria and data synthesis procedures without modifying the original research objectives.

### Study Classification

- Study type: Systematic review with descriptive quantitative synthesis
- Reporting framework: PRISMA 2020
- Methodological guidance: *Cochrane Handbook for Systematic Reviews of Interventions*
- Registration: PROSPERO (CRD420251160314)
- Time frame: January 2014 to December 2024

### Information Sources

A comprehensive literature search was conducted in Scopus and Web of Science Core Collection covering publications from January 2014 to December 2024. Searches were limited to studies published in English or Spanish. Reference lists of all eligible articles were manually screened to identify additional relevant studies.

Although a complete search strategy was initially developed for PubMed/MEDLINE, a preliminary overlap assessment demonstrated that all PubMed-indexed records relevant to optical clearing and immunostaining were also retrieved through Scopus or Web of Science (WOS). Because these two databases yielded additional relevant studies not indexed in PubMed, the final search was restricted to WOS and Scopus to avoid redundancy while maintaining completeness.

### Search Strategy

Search terms were constructed using a combination of controlled vocabulary and free-text keywords, linked with Boolean operators (AND / OR) and adapted to the syntax of each database. Quotation marks were used to delimit exact phrases. The strategy was structured according to the PICOC framework, ensuring coverage of population, intervention, comparison, outcomes, and context (Table 1).

PICOC-derived terms were applied to titles, abstracts, and keywords. The final search strategies are detailed below.

**Table 1.** Classification of search terms according to the PICOC framework.

| PICOC Category                | Search Descriptors                                                                                                                                                                           | Purpose of the search                                                                                                                                        |
|-------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Population                    | <i>"human brain tissue",<br/>"murine brain tissue",<br/>"formaldehyde-fixed tissue"</i>                                                                                                      | Defines the target population: human or murine brain tissue, fixed or postmortem.                                                                            |
| Intervention                  | <i>"CLARITY", "CUBIC", "SeeDB",<br/>"tissue clearing"</i>                                                                                                                                    | Identifies the main intervention: application of different tissue-clearing methods.                                                                          |
| Comparator                    | <i>"comparative studies",<br/>"comparison between<br/>clearing protocols"</i>                                                                                                                | Represents the comparison between distinct tissue-clearing protocols or methods.                                                                             |
| Outcomes                      | <i>"structural preservation",<br/>"imaging depth", "signal-to-noise ratio"</i>                                                                                                               | Corresponds to the primary outcomes defined in the PROSPERO protocol.                                                                                        |
| Context                       | <i>"3D brain visualization with<br/>immunolabeling", "molecular<br/>staining", "neuroimaging<br/>with molecular markers",<br/>"visualizing brain structures<br/>at molecular resolution"</i> | Delineates the experimental and technological context of the studies.                                                                                        |
| Complementary Technical Terms | <i>"fluorophore-conjugated antibodies",<br/>"immunofluorescence",<br/>"immunostaining"</i>                                                                                                   | Refines the search by including fluorescent immunolabeling techniques associated with tissue-clearing protocols                                              |
| Applied Filters               | <i>LIMIT-TO (LANGUAGE, "English") OR LIMIT-TO (LANGUAGE, "Spanish")<br/>PUBYEAR &gt; 2013 AND<br/>PUBYEAR &lt; 2025</i>                                                                      | Restricts the search to recent publications (2014–2024) in English or Spanish, corresponding to the period following the introduction of the CLARITY method. |

**Scopus (TITLE-ABS-KEY)**

*((("3D brain visualization with immunolabeling" OR "molecular staining" OR "neuroimaging with molecular markers" OR "visualizing brain structures at molecular resolution" OR "human brain tissue" OR "murine brain tissue" OR "formaldehyde-fixed tissue")*

*AND ("fluorophore-conjugated antibodies" OR "immunofluorescence" OR "immunostaining" OR "CLARITY" OR "CUBIC" OR "SeeDB" OR "tissue clearing")*

*AND ("comparative studies" OR "comparison between clearing protocols") AND ("structural preservation" OR "imaging depth" OR "signal-to-noise ratio"))*

*AND (LIMIT-TO (LANGUAGE, "English") OR LIMIT-TO (LANGUAGE, "Spanish")) AND (PUBYEAR > 2013 AND PUBEYEAR < 2025))*

**Web of Science (TS)**

*((("3D brain visualization with immunolabeling" OR "molecular staining" OR "neuroimaging with molecular markers" OR "visualizing brain structures at molecular resolution" OR "human brain tissue" OR "murine brain tissue" OR "formaldehyde-fixed tissue")*

*AND (“fluorophore-conjugated antibodies” OR “immunofluorescence” OR “immunostaining” OR “CLARITY” OR “CUBIC” OR “SeeDB” OR “tissue clearing”)*

*AND (“comparative studies” OR “comparison between clearing protocols”)*

*AND (“structural preservation” OR “imaging depth” OR “signal-to-noise ratio”)*

Prior to the final search, the strategy was piloted through scoping searches to ensure adequate sensitivity and confirm retrieval of key studies using alternative terminology. This iterative refinement minimized over-restriction while preserving conceptual specificity.

## **Eligibility Criteria**

### ***Inclusion***

- Original experimental studies.
- Studies combining tissue-clearing protocols or protocols with immunostaining (primary antibody plus fluorophore-conjugated secondary antibody).
- Human or murine brain tissue (fixed or postmortem).
- Reporting at least one predefined outcome: maximum effective imaging depth, fluorescent signal quality, unwanted background fluorescence, or morphological preservation.
- Publications in peer-reviewed journals.

### ***Exclusion***

- Reviews, editorials, or methodological papers without original experimental data.
- Studies not employing optical clearing and immunostaining.
- Non-brain tissues or studies relying solely on endogenous fluorescence without antibody labeling.
- Incomplete reports or lack of full-text availability.
- Grey literature sources, conference abstracts, institutional repositories, and preprint servers.

Grey literature was intentionally excluded because optical performance metrics require validated, peer-review experimental protocols. The risk of publication bias was considered low due to redundancy across major citation databases and the consistent publication patterns observed in this field.

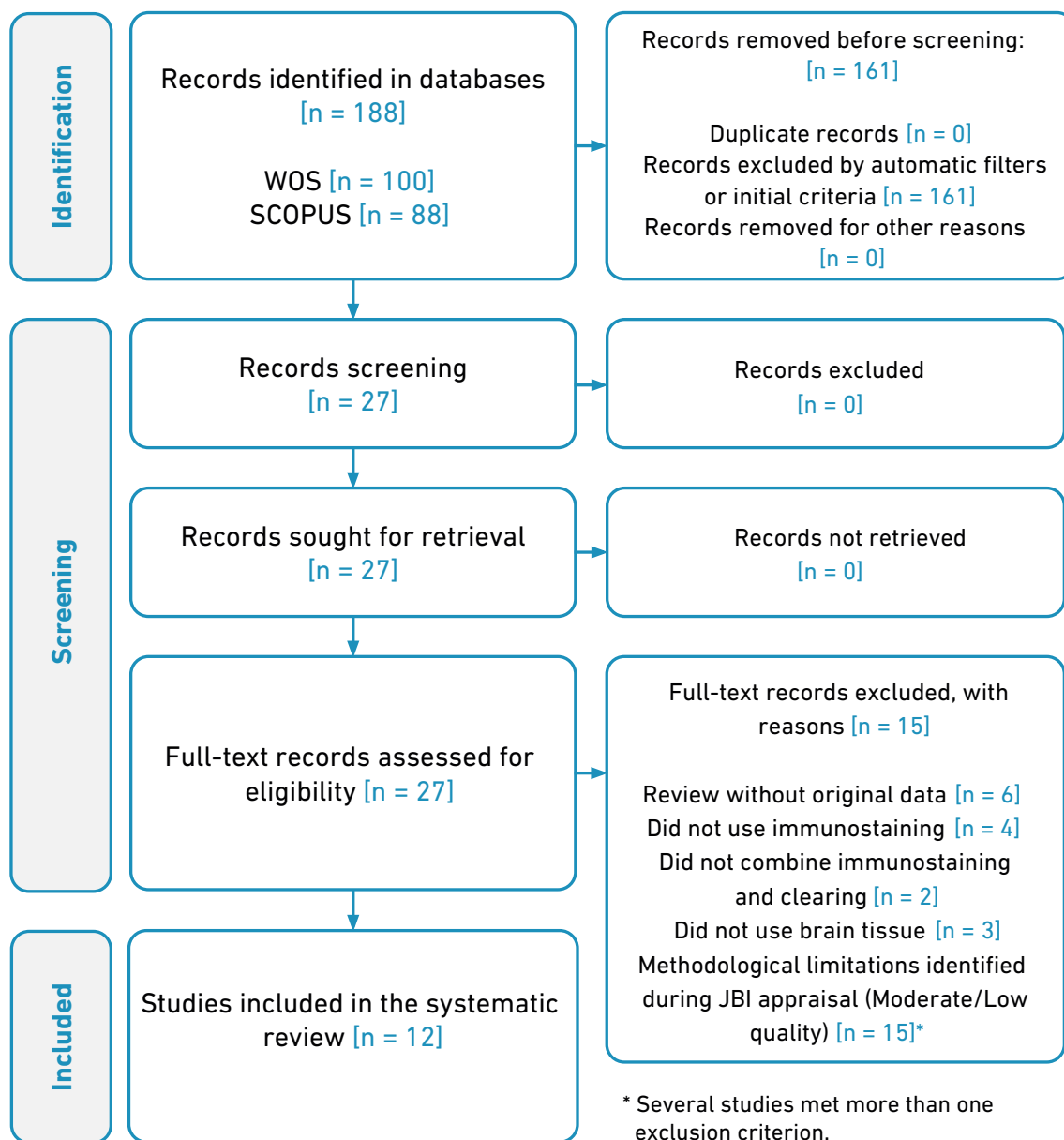
## **Selection Process and PRISMA Flow**

Automatic database filters were applied prior to screening to restrict publication year (2014-2024), language (English or Spanish), and document type (original research articles). After duplicate removal, two independent reviewers screened titles and abstracts. All 27 records retained after automated filtering met the eligibility criteria and proceeded directly to full-text assessment.

At the full-text stage, eligibility criteria and methodological quality were evaluated concurrently. Methodological quality was assessed using the JBI Critical Appraisal Checklist. Studies rated as moderate or low quality, or failing eligibility criteria, were excluded. Fifteen

studies were excluded at this stage. The remaining 12 studies met all inclusion criteria and were rated as high quality, forming the final dataset for quantitative synthesis.

Discrepancies were resolved by consensus. Inter-rater agreement for quality assessment was excellent (Cohen's  $\kappa = 0.883$ ). The selection process is summarized in the PRISMA 2020 flow diagram [26] (Figure 2).



**Figure 2.** PRISMA Flow Diagram for the Study Selection Process.

## Data Extraction and Coding

Data were extracted and coded into a structured database using SPSS v.29. Variables were categorized as follows:

- General variables: year, country, species, brain region, tissue type, processing time.
- Qualitative variables: clearing protocol, chemical principle, antibodies combinations, fluorophores, microscopy modality.
- Ordinal variables: morphological preservation, fluorescent signal quality, unwanted background fluorescence (three-level scale).
- Quantitative variable: maximum effective imaging depth ( $\mu\text{m}$ ).

## Methodological Quality Assessment

Methodological quality was independently assessed using the JBI Critical Appraisal Checklist for Quasi-Experimental Studies [29]. This tool was selected because the included studies consisted of heterogeneous laboratory experiments without randomization or clinical control groups.

Checklist items were operationalized to capture bias sources relevant to optical and imaging studies, including protocol reproducibility, control of technical variability, reliability of outcome measurements, and appropriateness of data analysis. Items were scored as fulfilled (1), partially fulfilled (0.5), or not fulfilled (0). Studies were classified as high (>80%), moderate (60-79%), or low (<60%) quality.

Only high-quality studies with complete optical outcome data were included. Inter-rater agreement was assessed using Cohen's Kappa ( $\kappa = 0.883$ ;  $p < 0.001$ ) (Table 2), indicating excellent agreement [30].

**Table 2.** Cohen's Kappa and observed agreement for JBI quality assessment (n=27).

| Quality    |        | Reviewer 2 |            |            |           |
|------------|--------|------------|------------|------------|-----------|
|            |        | Low        | Middle     | High       | Total     |
| Reviewer 1 | Low    | 4 (80.0%)  | 1 (20.0%)  | 0 (0.0%)   | 5 (100%)  |
|            | Middle | 1 (10.0%)  | 9 (90.0%)  | 0 (0.0%)   | 10 (100%) |
|            | High   | 0 (0.0%)   | 0 (0.0%)   | 12 (100%)  | 12 (100%) |
|            | Total  | 5 (18.5%)  | 10 (37.0%) | 12 (44.4%) | 27 (100%) |

**Note.** Cohen's kappa = 0.883 (SE=0.078),  $p < 0.001$ ; observed agreement (Po): 92.6

## Data Synthesis and Statistical Analysis

All analyses were conducted using SPSS version 29 and were predefined as descriptive and exploratory. Continuous variables were summarized using means and standard deviations, Normality and homoscedasticity were assessed using the Kolmogorov-Smirnov and Levene's tests, respectively.

### ***Operationalization of outcome variables***

Given the heterogeneity in outcome reporting, optical performance variables were harmonized using three-level ordinal scales (low, moderate, high). Quantitative measures and qualitative descriptions were mapped to these categories using predefined criteria based on relative performance within and across studies. When numerical values were unavailable, classification was achieved through independent reviewer consensus.

Ordinal variables were treated as quasi-continuous ones for exploratory comparisons, assuming approximately equidistant categories. This approach was adopted to identify global descriptive patterns rather than estimate precise effect sizes.

Group comparisons across chemical clearing categories were performed using one-way ANOVA or Welch's ANOVA, with post hoc testing (Tukey HSD or Games–Howell) as appropriate. Associations between optical outcomes were assessed using Spearman's rank correlation coefficient. Simple linear regression models were applied to explore potential functional relationships, with results interpreted as descriptive trends.

A p-value < 0.05 was considered statistically significant. Values between 0.05 and 0.10 were interpreted as non-significant trends. No correction for multiple comparisons was applied, given the exploratory nature of the analyses.

Heatmap visualizations were generated by averaging ordinal scores within antibody-protocol combinations. These averages were used exclusively for descriptive visualization and were not treated as continuous measures nor subjected to inferential testing. Direct statistical associations between fluorescent signal quality and unwanted background fluorescence were not independently tested.

### ***Bibliometric Analysis***

A descriptive bibliometric analysis was conducted using VOSviewer version 1.6.20 to examine keyword co-occurrence and co-authorship networks. Full counting was applied with a minimum occurrence threshold of one. Networks were interpreted based on frequency, link strength, and thematic clustering.

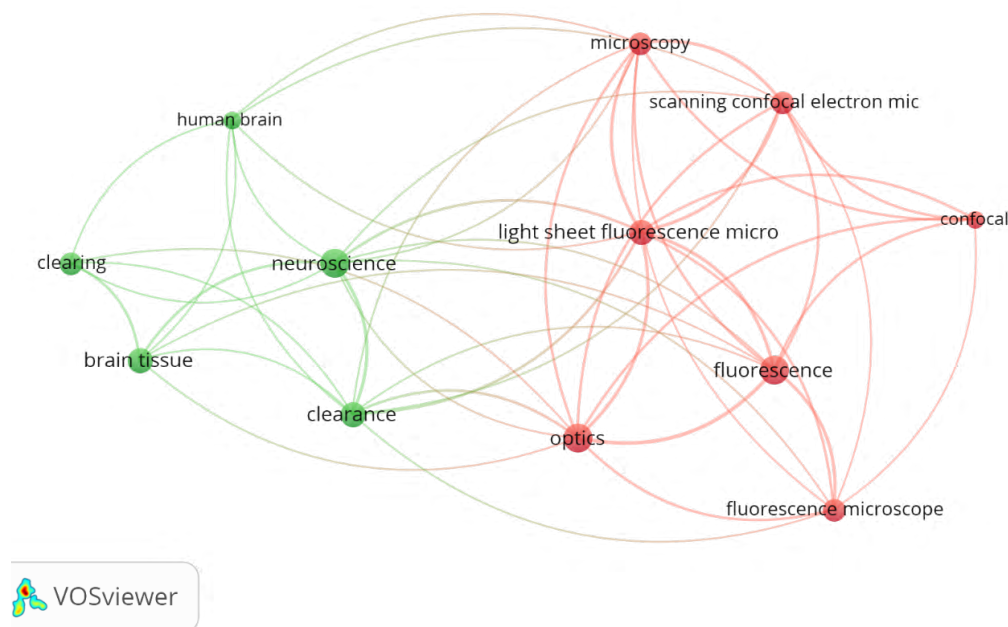
### ***Ethical Considerations***

This study involved no primary experimentation with human or animal subjects. All analyses were based exclusively on previously published data and adhered to international ethical guidelines for systematic reviews.

## **Results**

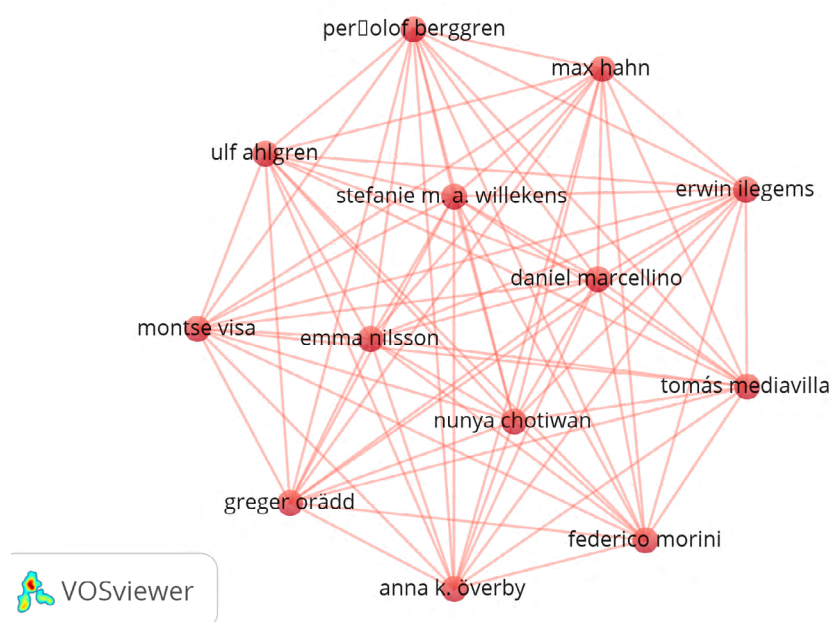
### ***Bibliometric and Network Analyses***

The keywords co-occurrence analysis (Figure 3) identified two main thematic clusters. The red cluster grouped terms related to the physicochemical principles of optical clearing and immunostaining, reflecting the methodological focus of studies assessing optical performance. The green cluster comprised concepts associated with the neuroscientific application of clearing and immunostaining techniques. The connections between both clusters indicate that methodological and application-oriented concepts frequently co-occur within the included studies. Given the limited number of experimental articles included, this network should be interpreted primarily as an exploratory representation of thematic structure rather than as a comprehensive bibliometric mapping of the field.



**Figure 3.** Co-occurrence network of keywords in research on optical clearing and three-dimensional brain imaging.

The co-authorship network (Figure 4) provides a descriptive overview of collaboration patterns among authors of the included studies. Each node represents an author, and links indicate shared publications. The network highlights recurrent collaborations among research groups working at the interface of neuroscience, biomedical optics, and tissue engineering. This analysis is presented for contextual purposes and does not imply differences in scientific contribution or methodological influence.



**Figure 4.** Co-authorship network of researchers involved in studies on brain tissue optical clearing.

Study characteristics

Table 3 summarizes the general characteristics of the 12 experimental studies published between 2014 and 2024 that were included in the quantitative synthesis. Nine studies (75%) were conducted in rodent brain tissue, while three (25%) examined human brain samples. A range of tissue-clearing protocols was represented, including CLARITY, CUBIC, SeeDB, iDISCO+, uDISCO, and ProExM. These methods corresponded to hydrogel-based, hydrophilic solution-based, organic solvent-based, and hybrid chemical principles.

Table 3. Summary of studies included in the systematic review.

| Study                                                                                                                                | Brain structure/<br>Species | Tissue<br>Type | Chemical<br>principle* | Primary/<br>Secondary<br>Antibody    | Duration<br>(days) | Microscopy              |
|--------------------------------------------------------------------------------------------------------------------------------------|-----------------------------|----------------|------------------------|--------------------------------------|--------------------|-------------------------|
| A novel, modernized Golgi-Cox stain optimized for CLARITY cleared tissue                                                             | Hippocampus/<br>Rodent      | Fixed          | Hydrogel               | Anti-NeuN/<br>AF-488                 | 11-14              | Combined,<br>Two-photon |
| An MR-based brain template and atlas for optical projection tomography and light sheet fluorescence microscopy in neuroscience       | Whole brain/<br>Rodent      | Fixed          | Hydrogel               | Anti-NeuN<br>Anti-GFAP/<br>AF-594    | 11-14              | Two-photon<br>and LSM   |
| Brain pericytes acquire stemness via the nrf2-dependent antioxidant system                                                           | Striatum/<br>Rodent         | Fixed          | Organic<br>Solvent     | Anti-TH<br>Anti-GFAP/<br>AF-594/647  | 7-10               | LSFM                    |
| Clarifying clarity: quantitative optimization of the diffusion based delipidation protocol for genetically labeled tissue            | Whole brain/<br>Rodent      | Fixed          | Hydrogel               | Anti-NeuN/<br>AF-488                 | 11-14              | Combined                |
| Comparative analyses of clearing efficacies of tissue clearing protocols by using a punching assisted clarity analysis               | Hippocampus/<br>Rodent      | Fixed          | Hydrophilic            | Anti-NeuN<br>Anti-GFAP/<br>AF-594    | 7-10               | Combined                |
| Comparison of light-sheet fluorescence microscopy and fast-confocal microscopy for three- dimensional imaging of cleared mouse brain | Cortex/<br>Rodent           | Fixed          | Hydrophilic            | Anti-GFAP/<br>AF-488                 | < 7                | Laser<br>Confocal       |
| Fast 3-D imaging of brain organoids with a new single-objective planar-illumination two-photon microscope                            | Whole brain/<br>Rodent      | Fixed          | Organic<br>Solvent     | Anti-TH<br>Anti-GAD67/<br>AF-594/647 | 7-10               | LSFM                    |
| Immunofluorescent visualization of mouse interneuron subtypes.                                                                       | Cortex/<br>Human            | Post<br>mortem | Hydrogel               | Anti-NeuN<br>Anti-GFAP/<br>AF-488    | 11-14              | Combined                |

|                                                                                                              |                        |                    |                 |                                  |       |                |
|--------------------------------------------------------------------------------------------------------------|------------------------|--------------------|-----------------|----------------------------------|-------|----------------|
| On-line optical clearing method for whole-brain imaging in mice                                              | Whole brain/<br>Rodent | Fixed              | Organic Solvent | Anti-NeuN/<br>AF-647             | 7-10  | LSFM           |
| Optimizing tissue clearing and imaging methods for human brain tissue                                        | Cortex/<br>Human       | <i>Post mortem</i> | Hydrogel        | Anti-GFAP/<br>AF-488             | 11-14 | Laser Confocal |
| Protein-retention expansion microscopy for visualizing subcellular organelles in fixed brain tissue          | Hippocampus/<br>Rodent | Fixed              | Hybrid          | Anti-NeuN/<br>AF-488             | <7    | Combined       |
| Scalable labeling for cytoarchitectonic characterization of large optically cleared human neocortex samples. | Cortex/<br>Human       | <i>Post mortem</i> | Hydrophilic     | Anti-NeuN<br>Anti-MBP/<br>AF-647 | >14   | Combined       |

**Notes.** LSFM: Light Sheet Fluorescence Microscopy; AF: Alexa-Fluor; Combined: Laser Confocal + LSFM; \* Chemical principle of optical clearing methods.

All clearing protocols were combined with immunostaining procedures using primary and fluorophore-conjugated secondary antibodies. Three-dimensional imaging was performed using various microscopy modalities, including two-photon microscopy, light-sheet fluorescence microscopy (LSFM), and confocal microscopy. The most frequently analyzed brain regions were the hippocampus, cerebral cortex, and whole brain preparations.

### Volumetric changes and morphological preservation

Regarding volumetric tissue changes, a statistically non-uniform distribution across contraction and expansion categories was observed only among CLARITY-based protocols (Fisher's exact test,  $p = 0.048$ ). All CLARITY studies reported mild tissue contraction (-5% to -10%), with no severe contraction or expansion. Given the small number of studies and multiple categorical comparisons, this finding should be interpreted as an exploratory within-method pattern.

No statistically significant distributional differences were observed for volumetric changes in the remaining clearing methods. Percentages reported in Table 4 reflect relative distributions derived from small method-specific sample sizes. Fisher's exact test was applied because of low expected cell counts, and all results were interpreted descriptively.

**Table 4.** Comparison of morphological and optical variables according to the tissue-clearing method in immunostained brain tissue.

| Variables                                            |          | Optical clearing methods |       |       |       |       |       |        |       |         |       |        |       |
|------------------------------------------------------|----------|--------------------------|-------|-------|-------|-------|-------|--------|-------|---------|-------|--------|-------|
|                                                      |          | CLARITY                  |       | CUBIC |       | SeeDB |       | uDISCO |       | iDISCO+ |       | ProExM |       |
|                                                      |          | %                        | $p^*$ | %     | $p^*$ | %     | $p^*$ | %      | $p^*$ | %       | $p^*$ | %      | $p^*$ |
| Qualitative assessment of signal-to-background ratio | Moderate | 60.0                     | 0.558 | 33.3  | —     | 50.0  | —     | 0.0    | —     | 0.0     | 0.470 | 0.0    | —     |
|                                                      | High     | 40.0                     |       | 66.6  |       | 50.0  |       | 100.0  |       | 100.0   |       | 100.0  |       |

|                                             |                                 |           |       |            |       |            |       |          |       |          |       |          |       |
|---------------------------------------------|---------------------------------|-----------|-------|------------|-------|------------|-------|----------|-------|----------|-------|----------|-------|
| Degree of structural preservation           | Moderate                        | 60.0      | 0.558 | 33.3       | —     | 50.0       | —     | 0.0      | —     | 0.0      | 0.470 | 0.0      | —     |
|                                             | High                            | 40.0      |       | 66.6       |       | 50.0       |       | 100.0    |       | 100.0    |       | 100.0    |       |
| Visible morphological alterations           | Mild                            | 100.0     | 0.205 | 66.6       | —     | 50.0       | 0.455 | 100.0    | —     | 100.0    | —     | 0.0      | —     |
|                                             | Moderate                        | 0.0       |       | 33.3       |       | 50.0       |       | 0.0      |       | 0.0      |       | 100.0    |       |
| Tissue volumetric change (%)                | Severe Contraction (-25 to -30) | 0.0       | 0.048 | 0.0        | 0.503 | 0.0        | 0.167 | 100.0    | 0.141 | 50.0     | 0.526 | 0.0      | 0.141 |
|                                             | Mild Contraction (-5 to -10)    | 100.0     |       | 66.6       |       | 50.0       |       | 0.0      |       | 50.0     |       | 0.0      |       |
|                                             | Mild Expansion (5-10)           | 0.0       |       | 0.0        |       | 50.0       |       | 0.0      |       | 0.0      |       | 0.0      |       |
|                                             | Moderate Expansion (15 - 25)    | 0.0       |       | 33.3       |       | 0.0        |       | 0.0      |       | 0.0      |       | 100.0    |       |
| Unwanted background fluorescence            | Low                             | 20.0      | 0.576 | 33.3       | —     | 0.0        | 0.515 | 100.0    | 0.333 | 50.0     | —     | 0.0      | —     |
|                                             | High                            | 80.0      |       | 66.6       |       | 100.0      |       | 0.0      |       | 50.0     |       | 100.0    |       |
| Fluorescent signal quality                  | Moderate                        | 60.0      | 0.222 | 0.0        | 0.491 | 50.0       | —     | 0.0      | —     | 0.0      | 0.515 | 0.0      | —     |
|                                             | High                            | 40.0      |       | 100.0      |       | 50.0       |       | 100.0    |       | 100.0    |       |          |       |
| Maximum effective imaging depth (μm) [x±SD] |                                 | 2860±1045 |       | 1540 ± 450 |       | 4333 ±1155 |       | 5000 ± — |       | 4000 ± — |       | 1200 ± — |       |

**Notes.** \*Fisher test; — Invalid comparison.

### Comparative statistical analyses

The Kolmogorov–Smirnov test with Lilliefors correction indicated a normal distribution for maximum effective imaging depth values ( $K-S = 0.147$ ;  $p = 0.200$ ), supporting the use of parametric methods for group comparisons.

Group-level comparisons of optical performance according to clearing methods, stratified by chemical principles, are presented in Table 5. Levene's test indicated heterogeneity of variances for fluorescent signal quality ( $p = 0.002$ ), while the remaining variables met the assumption of homoscedasticity ( $p > 0.45$ ). Neither one-way ANOVA nor Welch's ANOVA identified statistically significant differences between clearing categories for any of the analyzed outcomes (all  $p > 0.05$ ). Consequently, observed differences across clearing categories are interpreted as descriptive patterns rather than statistically supported between-group effect.

Post hoc analysis using Games–Howell and Tukey HSD tests likewise showed no statistically significant pairwise differences ( $p \geq 0.133$ ). Effect size estimates ( $\eta^2$ ) ranged from 0.16 to 0.27, corresponding to small-to-moderate magnitudes. These estimates provide only an approximate indication of effect size and should be interpreted as exploratory. Organic solvent-based methods showed the highest descriptive values for fluorescent signal quality and maximum effective imaging depth; however, these trends did not reach statistical significance.

**Table 5.** Comparative analysis of fluorescent signal quality, unwanted background fluorescence level, and maximum effective imaging depth according to the chemical principle of tissue-clearing methods.

| Chemical principle of tissue-clearing methods | Fluorescent signal quality |          |                | Unwanted background fluorescence level |          |                | Maximum effective imaging depth (μm) |          |                     |
|-----------------------------------------------|----------------------------|----------|----------------|----------------------------------------|----------|----------------|--------------------------------------|----------|---------------------|
|                                               | <i>F o Δ</i>               | <i>p</i> | 95% <i>CI</i>  | <i>F o Δ</i>                           | <i>p</i> | 95% <i>CI</i>  | <i>F o Δ</i>                         | <i>p</i> | 95% <i>CI</i>       |
| Levene's test (based on means)                | 15.65                      | 0.002    | -              | 0.387                                  | 0.691    | -              | 0.877                                | 0.452    | -                   |
| ANOVA (classical)                             | 1.455                      | 0.289    | -              | 0.773                                  | 0.493    | -              | 0.996                                | 0.411    | -                   |
| Welch's ANOVA                                 | $\bar{\Delta}=0$           |          |                | 0.618                                  | 0.584    | -              | 1.925                                | 0.255    | -                   |
| Post hoc                                      | Games-Howell               |          |                | Tukey HSD                              |          |                | Tukey HSD                            |          |                     |
| Hydrogel vs. Hydrophilic                      | 0.267                      | 0.805    | [-1.72 – 1.18] | 0.133                                  | 0.938    | [-1.35 – 1.62] | 206.6                                | 0.98     | [-3282.9 – 2869.6]  |
| Hydrogel vs. Organic Solvents                 | 0.600                      |          | [-1.47 – 0.27] | 0.467                                  | 0.521    | [-1.02 – 1.95] | 1473.3                               | 0.4      | [-4549.6 – 1602.93] |
| Hydrophilic vs. Organic Solvents              | 0.333                      |          | [-2.30 – 1.63] | 0.333                                  | 0.773    | [-1.35 – 2.01] | 1204.0                               | 0.567    | [-4706.0 – 2172.7]  |
| Effect size ( $\eta^2$ )                      | 0.266                      |          |                | 0.161                                  |          |                | 0.199                                |          |                     |

**Note.**  $\bar{\Delta} = 0$  (the variance is zero in at least one of the groups compared). The hybrid method was not included in the analysis, as it was applied in only one study.

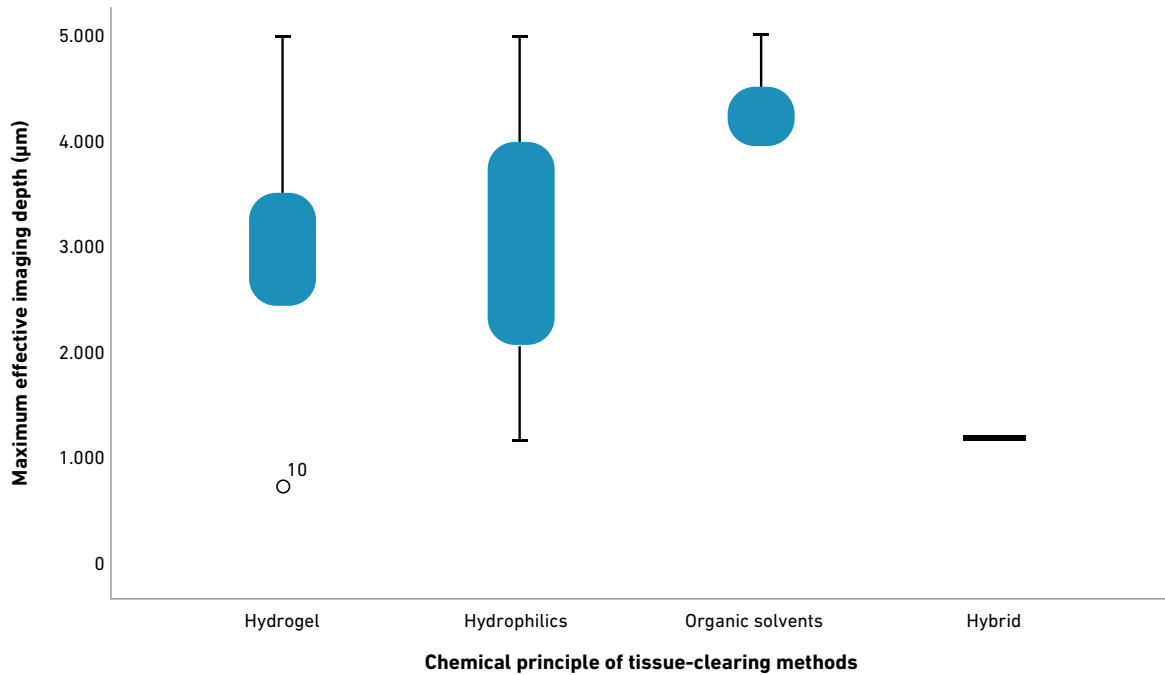
### Imaging depth distribution

Figure 5 presents a boxplot of maximum effective imaging depth by clearing chemistry. Organic solvent-based protocols exhibited the highest median imaging depths (approximately 4000–5000 μm) with relatively compact interquartile ranges. Hydrophilic methods showed the widest dispersion of value (approximately 1500–5000 μm). Hydrogel-based methods displayed intermediate maximum effective imaging depths (approximately 2500–3000 μm), whereas hybrid approaches showed the lowest optical penetration (<1500 μm).

Although an apparent ordering of median maximum effective imaging depth was observed across clearing categories, this pattern remained descriptive and was not supported by statistically significant group differences.

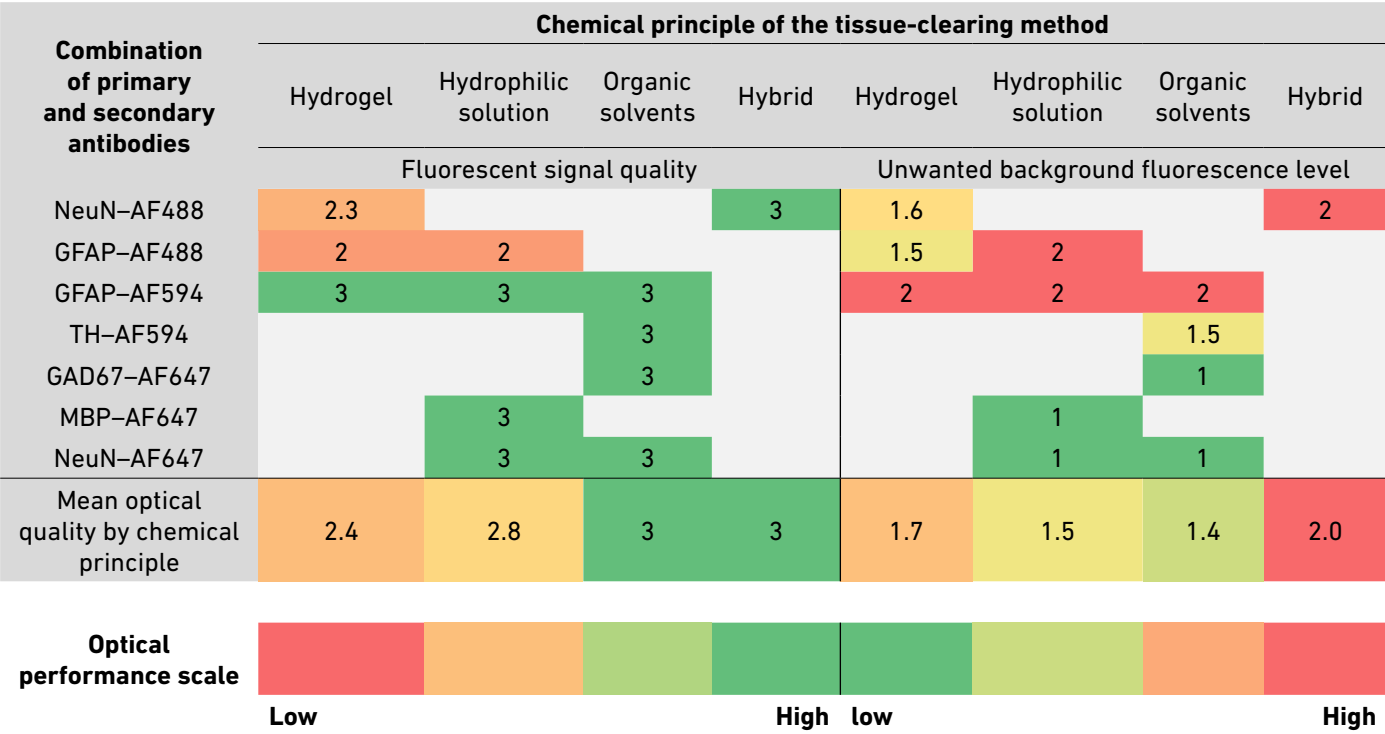
### Heatmap analysis of optical quality

Figure 6 presents a descriptive heatmap illustrating the distribution of ordinal optical quality scores across primary-secondary antibody combinations and clearing chemistry. The displayed values represent mean descriptive scores derived from ordinal classifications and are used exclusively to facilitate visual comparison across categories.



**Figure 5.** Distribution of maximum effective imaging depth (μm) according to the chemical principle of the tissue-clearing method.

**Note:** ANOVA:  $p>0.05$ ; Tukey:  $p=0.426$



**Figure 6.** Optical performance according to the combination of primary and secondary antibodies and the chemical principle of tissue-clearing methods.

For fluorescent signal quality, higher intensity is represented in green and lower intensity in red. For unwanted background fluorescence, green denotes lower levels and red denotes higher levels.

Organic solvent-based protocols (iDISCO+, uDISCO) showed a higher proportion of antibody combinations classified as having high fluorescent signal quality and low unwanted background fluorescence, particularly for Alexa Fluor 594/647 conjugates (e.g., GFAP-AF594, TH-AF594, GAD67-AF647, NeuN-AF647). This distribution suggests a descriptive tendency toward higher fluorescent signal quality combined with lower unwanted background fluorescence under solvent-based clearing conditions.

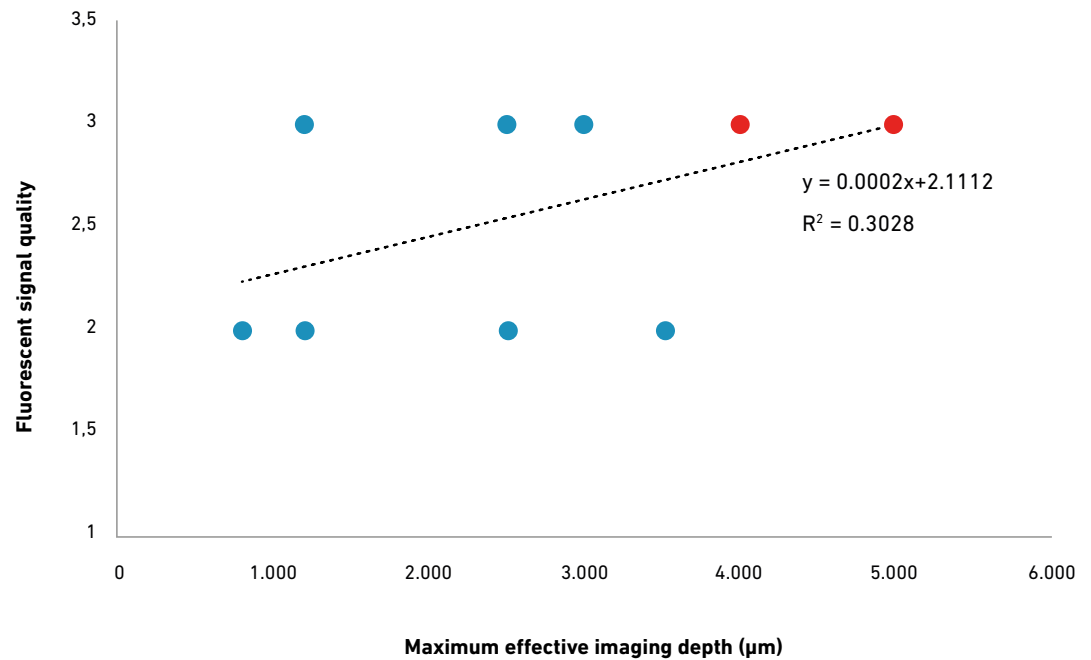
Visual inspection of the heatmap indicates that higher signal quality scores frequently co-occur with lower unwanted background fluorescence levels. However, this apparent inverse association is based on descriptive visualization and was not evaluated using a direct statistical test between the two ordinal variables. Hydrophilic and hydrogel-based protocols more frequently exhibited moderate signal quality and unwanted background fluorescence categories, consistent with their emphasis on structural preservation. These heatmap findings align with, but do not independently confirm, the patterns observed in correlation and regression analyses involving maximum effective imaging depth.

### Correlations and regression analyses

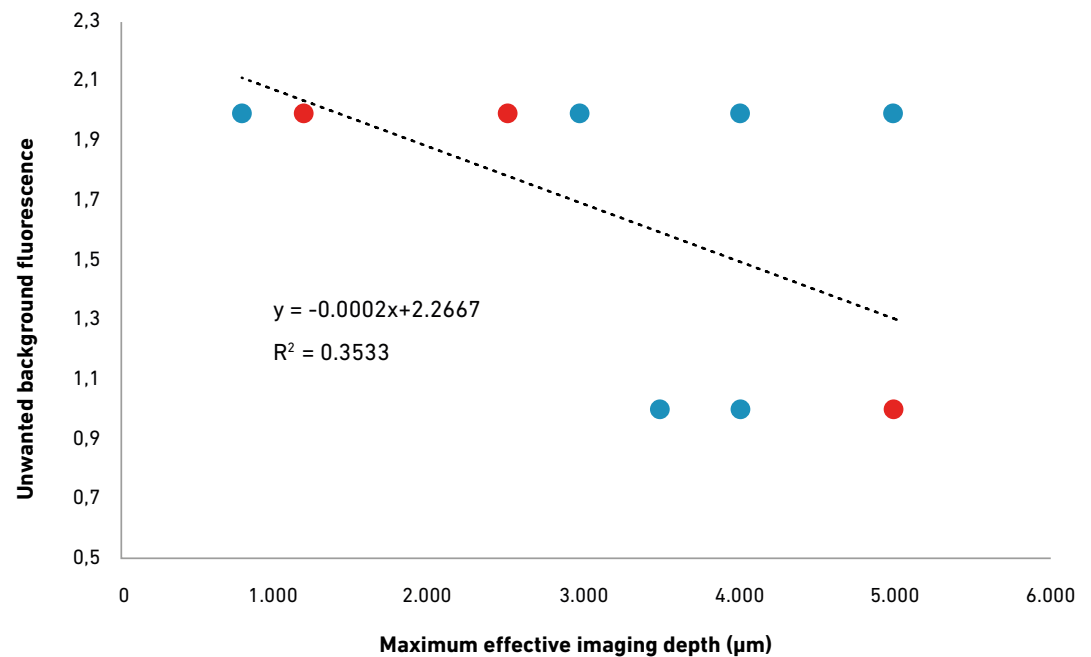
Spearman correlation revealed a moderate negative association between unwanted background fluorescence and maximum effective imaging depth, ( $\rho = -0.596$ ;  $p = 0.041$ ). In contrast, the association between maximum effective imaging depth and fluorescent signal quality did not meet the predefined threshold for statistical significance ( $\rho = 0.570$ ;  $p = 0.053$ ), although it showed a positive non-significant trend.

These results indicate that increased maximum effective imaging depth is significantly associated with reduced unwanted background fluorescence, whereas its relationship with fluorescent signal quality remains statistically inconclusive.

Simple linear regression analyses (Figure 7) showed that approximately 30% of the variability in fluorescent signal quality and 35% of the variability in unwanted background fluorescence were explained by maximum effective imaging depth ( $R^2 = 0.3028$ ;  $R^2 = 0.3533$ , respectively). These models illustrate a direct relationship between maximum effective imaging depth and fluorescent signal quality and an inverse relationship with unwanted background fluorescence. Given the ordinal nature of the variables and the limited number of aggregated observations, these regression results should be considered exploratory and interpreted cautiously.



a) Relationship between maximum effective imaging depth ( $\mu\text{m}$ ) and fluorescent signal quality ( $\rho = 0.570$ ;  $p = 0.053$ ).



b) Relationship between maximum effective imaging depth ( $\mu\text{m}$ ) and Unwanted background fluorescence ( $\rho = -0.596$ ;  $p = 0.041$ ).

**Figure 7.** Linear regression plots showing the bivariate correlations between maximum effective imaging depth, fluorescent signal quality, and unwanted background fluorescence.

**Note:** Overlapping data points are highlighted in red to improve visual clarity.

## Discussion

This systematic review with descriptive quantitative synthesis provides a comparative evaluation of the principal optical clearing methods applied to immunostained brain tissue, integrating morphological and optical parameters. The results indicate that the chemical nature of the clearing method is associated with descriptive differences in maximum effective imaging depth, fluorescent signal quality, and unwanted background fluorescence. Across the analyzed studies, organic solvent-based protocols tended to exhibit higher optical performance in several descriptive outcomes, thereby supporting the study hypothesis at an exploratory level rather than demonstrating definitive superiority.

All comparative statements derived from this review should be interpreted as descriptive trends based on aggregated observations, rather than as evidence of statistically confirmed superiority or reference standards. The present findings are intended to inform methodological decision-making and hypothesis generation and do not establish definitive comparative effectiveness.

In the present descriptive synthesis, iDISCO+ and uDISCO tissue-clearing methods reached the highest reported maximum effective imaging depths (approximately 4000  $\mu\text{m}$ ) and exhibited high fluorescent signal quality. These findings are consistent with previous experimental reports, which have described the compatibility of these protocols with Alexa Fluor-conjugated antibodies in rodent brain tissue [28]. Although mechanistic parameters were not directly evaluated in this review, the higher optical performance observed for solvent-based methods is commonly attributed in the literature to efficient lipid removal and refractive index matching, which reduce internal light scattering and improve optical homogeneity [8,29-31]. These observations are presented as descriptive trends, given the absence of statistically significant differences in variance-based comparisons.

Hydrophilic solution-based methods, such as CUBIC and SeeDB, exhibited intermediate optical outcomes, while maintaining favourable morphological preservation. These patterns align with prior studies emphasizing their structural stability [2,32]. These protocols offer a balance between transparency and fluorophore compatibility, although they are associated with higher unwanted background fluorescence compared with organic solvent-based methods [11,33,34]. Hydrogel-based protocols, such as CLARITY, consistently preserved tissue architecture but achieved lower maximum effective imaging depth, likely due to constraints on light transmission and antibody diffusion imposed by the polymeric matrix [7,35-37]. Within the present datasets, the hybrid expansion-based ProExM technique was associated with high fluorescent signal quality but lower effective imaging depth. This trade-off has been more extensively characterized in previous studies, where physical expansion has been shown to enhance spatial resolution at the expense of total analysable volume [14,38,39].

Regarding associations between optical variables, a statistically significant negative correlation was observed between unwanted background fluorescence and maximum effective imaging depth ( $\rho = -0.596$ ,  $p = 0.041$ ). This finding indicates that increased tissue transparency is reliably associated with reduced unwanted background fluorescence. In contrast, the positive association between fluorescent signal quality and maximum effective imaging depth represented a non-significant trend ( $\rho = 0.570$ ,  $p = 0.053$ ) and should therefore be interpreted cautiously. These results are consistent with observations reported by Ueda et al., who attributed unwanted background fluorescence to the retention of endogenous pigments and aldehydes [1,40-43].

A study conducted in Wuhan reported a multiple linear regression model in which fluorescent signal quality and unwanted background fluorescence jointly accounted for more than 70% of the variance in maximum effective imaging depth. Although these findings were obtained under different experimental conditions, they provide a useful comparative framework for interpreting how clearing efficiency may influence photonic performance. In that context, organic solvents were described as acting as both delipidizing and refractive agents, contributing to enhanced optical homogeneity [19,44,45].

Heatmap analyses indicated that, within the analysed dataset, organic solvent-based methods were more frequently associated with antibody combinations classified as high fluorescent signal quality and low unwanted background fluorescence, particularly for Alexa Fluor 594/647 conjugates. These observations are descriptive in nature and align with, but do not independently confirm, the patterns identified in the statistical analyses. In this context, this tendency is consistent with the recommendations of Mai et al., who highlighted the advantage of high-photostability secondary antibodies in human postmortem brain studies [16,46-48].

Overall, all analysed clearing protocols exhibited satisfactory optical performance within their respective methodological context. Organic solvent-based methods tended to provide greater tissue transparency and higher fluorescent signal quality in descriptive comparisons. Hydrophilic solution-based methods offered a balance between structural preservation and optical penetration. Hydrogel-based protocols favoured morphological stability, whereas hybrid approaches emphasized maximal optical resolution. These patterns are consistent with prior methodological synthesis reported by Richardson and Lichtman [5,46].

Several limitations should be considered when interpreting these findings. Variability in primary and secondary antibody combinations, differences in microscopy techniques, and incomplete quantitative reporting contributed to experimental heterogeneity. In addition, fixation procedures were not consistently described across studies, which may have influenced optical outcomes [49-51]. Despite these constraints, the convergence of descriptive indicators across studies supports the internal coherence and descriptive robustness of the present quantitative synthesis.

In summary, this systematic review with descriptive quantitative synthesis indicates that optical clearing methods play a relevant role in shaping the visualization quality of immunostained brain tissue. Organic solvent-based methods consistently exhibited higher maximum effective imaging depth and favourable fluorescent signal quality in descriptive comparisons, albeit at the cost of moderate volumetric contraction. These methods may be considered a strong methodological option for three-dimensional neuroimaging studies in descriptive terms, particularly when optical transparency is prioritized over maximal morphological preservation. Future research should aim to improve immunological compatibility and minimize structural alterations to achieve an optimal balance between transparency, morphological preservation, and optical quality.

### Limitations and Recommendations

Several limitations should be considered when interpreting the findings of this review.

First, substantial heterogeneity across studies was observed with respect to clearing chemistry, immunostaining conditions, imaging modalities, and definitions of optical outcomes. This variability limited direct comparability between studies and precluded the application of effect-size-based meta-analytic models.

Second, optical performance outcomes were harmonized using three-level ordinal classification schemes to accommodate heterogeneous reporting formats. Although this approach enabled structured synthesis, the subsequent use of descriptive and parametric statistical methods on ordinally coded data represents an additional methodological constraint that should be considered when interpreting comparative results.

Third, the quantitative associations identified between maximum effective imaging depth, fluorescent signal quality, and unwanted background fluorescence were exploratory in nature and derived from a small number of heterogeneous studies ( $n = 12$ ). The exclusion of Moderate-and Low-quality studies based on JBI appraisal further reduced the effective sample size for correlation and regression analyses. Consequently, the reported correlation coefficients and  $R^2$  values should be interpreted as indicative trends rather than stable predictive estimates.

Future research would benefit from standardized measurement criteria for tissue transparency, fluorescent signal quality, and unwanted background fluorescence, as well as from larger, well-controlled experimental datasets. In addition, systematic evaluation of hybrid protocols that aim to balance morphological preservation with optical performance may help address current methodological trade-offs and support more robust comparative analyses.

## Conclusions

This systematic review with descriptive quantitative synthesis indicates that the chemical principle underlying tissue-clearing methods is associated with descriptive differences in optical performance and morphological preservation in immunostained brain tissue.

In descriptive comparisons, organic solvent-based methods tended to exhibit greater maximum effective imaging depth and favourable fluorescent signal characteristics, suggesting a potential advantage for three-dimensional neuroimaging applications. However, in the absence of statistically significant between-group differences and given the limited number of heterogeneous studies, these findings should be interpreted as indicative patterns rather than evidence of proven superiority.

Hydrophilic solution-based methods showed intermediate performance, balancing structural preservation and optical transparency, whereas hydrogel-based approaches consistently favoured morphological stability at the expense of maximum effective imaging depth. Hybrid expansion-based techniques achieved high subcellular resolution but were associated with reduced maximum effective imaging depth and were underrepresented in comparative analyses.

Increased maximum effective imaging depth was significantly associated with reduced unwanted background fluorescence, while its relationship with fluorescent signal quality did not reach statistical significance and should therefore be interpreted cautiously. Overall, the observed patterns suggest a possible functional interplay between tissue transparency and fluorescent signal quality characteristics, which requires confirmation in larger, standardized experimental datasets.

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