# Comparative Efficacy of Different Chemotherapeutic Regimens in the Treatment of *Peste des Petits Ruminants* in West African Dwarf Goats\*

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#### Abstract

The comparative efficacy of different chemotherapeutic regimes in the treatment of natural clinical infection of Peste des petits ruminants (PPR) infection in West African Dwarf (WAD) goats were determined. Twenty WAD male goats divided into five groups, of four each were used. Group 1 was the combined therapy group treated with Acyclovir, Oxytetracycline and Ivermectin, Group 2 was treated with Acyclovir only, Group 3 was treated with Ivermectin only, Group 4 was treated with Oxytetracycline only and Group 5 was left untreated. Clinical signs such as nasal and ocular discharges, emaciation, anorexia, pyrexia, ulcerative oral lesions, respiratory distress and diarrhoea were observed in these goats during the period of their acclimatization. The prominent post-mortem lesion observed was discontinuous streaks of congestion (Zebra markings) in the mucosa of the colon and rectum. The physiologic parameters (temperature, pulse rate, heart rate, and respiratory rate) were taken daily. It was observed that there was a significant increase in the temperature above normal in the untreated group, suggesting pyrexia. It was also observed that the combined therapy group showed a smaller number of mortality and thus, had a better efficacy when compared to the other chemotherapeutic agents used singly.

Keywords: Peste des petits ruminants, Acyclovir, Oxyteteracycline, Ivermectin, mortality.

## Eficacia comparativa de diferentes regímenes quimioterapéuticos en el tratamiento de la Peste des petits ruminants (PPR) en cabras enanas de África occidental

#### Resumen

Se determinó la eficacia comparativa de diferentes regímenes quimioterapéuticos en el tratamiento de la infección clínica natural con la *Peste de los pequeños rumiantes* (PPR) en cabras enanas de África occidental (WAD). Se usaron veinte cabras WAD machos, distribuidas en cinco grupos, de cuatro cabras cada uno. El grupo 1 fue el grupo de terapia combinada, tratado con aciclovir, oxitetraciclina e ivermectina. El grupo 2 fue tratado con solo con aciclovir. El grupo 3 fue tratado con ivermectina solamente. El grupo 4 fue tratado con oxitetraciclina y el grupo 5 no recibió tratamiento. Se observaron signos clínicos como secreciones nasales y oculares, emaciación, anorexia, pirexia, lesiones orales ulcerativas, dificultad respiratoria y diarrea en estas cabras durante su periodo de aclimatación. La lesión post-mórtem más notable que se observó fueron las manchas descontinuas de congestión (como las rayas de la cebra) en la mucosa del colon y del

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**Palabras clave:** *Peste des petits ruminants,* aciclovir, oxitetraciclina, ivermectina, mortalidad.

#### INTRODUCTION

Peste des petits ruminants (PPR) is endemic across the African and Asian continents causing economic and veterinary important disease leading to considerable economic losses (1,2). It is known to be endemic in Nigeria as confirmed by Shamaki et al. (3). It is extremely contagious viral disease of sheep and goats, clinically manifested by pyrexia, purulent ocular and nasal discharges, dyspnoea, sneezing, pneumonia, ulceration of mucous membranes of the mouth, gastroenteritis and severe diarrhoea (4-7). The disease is caused by an RNA virus, the PPR virus of the Genus Morbilivirus and the Family Paramyxovirus. Genetic characterization of the virus has grouped it into four distinct lineages: three from Africa and one from Asia. One of the African lineages is also found in Asia (8-10). The aetiologic virus is easily transmitted by direct contact with the aerosols, secretions and/or excretions of infected animals (2,4,11). PPR is a transboundary animal disease causing enormous economic losses due to the high morbidity and mortality associated with the disease and has therefore been classified among World Organisation for Animal Health (OIE) notifiable diseases (12). It poses a significant threat to the development of livestock production in areas where it is endemic (13,14).

Programs aimed at controlling PPR, have costed farmers, governments and aid agencies so much money. Hence, this disease remains as a major constraint in the productivity of small ruminants and is responsible for severe economic losses according to Ogunsanmi *et al.* (15) and Libeau *et al.* (16). In addition to this, drug development provides possible solutions to some tropical health diseases, although pharmaceutical research is very exorbitant and a lack of return on investment has constrained the production of new drugs (17).

Vaccination has been the main method of control but despite it, sporadic outbreaks have also been reported by Şevik and Sait (18). In Nigeria, homologous vaccine for PPR was developed from the attenuation of Nig 75/1 PPRV strain Diallo (19) but recently, Luka *et al.* (20) reported that Nigerian strains of PPRV demonstrate 93-94% homology with PPRV homologous vaccine. Hence, there is no doubt that it is the vaccine of choice to control the PPR in Nigeria. However, the thermolabile nature of the vaccine may increase the likelihood of low sero-conversion following vaccination with the vaccine Sen *et al.* (21).

Hence this research was conducted to use clinical signs, Post-mortem lesions and Monoclonal antibody Blocking ELISA method to serologically diagnose PPR in West African Dwarf goats and to assess the efficacy of different chemotherapeutic regimens including Acyclovir for the treatment of PPR in West African Dwarf goats.

## **MATERIALS AND METHODS**

#### **Study Animals**

Twenty WAD male goats, whose ages ranges from six months to one year with no history of PPR vaccination adjudged by clinical signs and gross lesions to be suffering from natural PPR infection were purchased from the local markets in Ibagwa and Orba, Nsukka and Udenu Local Government Area, Enugu State, Nigeria. The WAD goats were kept together in a well aerated pen in the Department of Veterinary Surgery, University of Nigeria, Nsukka (UNN) in the experimental animal house unit. They were left to acclimatize for 7 days (1 week) prior to the commencement of the study.

During this period of acclimatization, the animals were thoroughly checked for the manifestation of characteristic clinical signs that are suggestive and points to the scope of the study. Also, during this period, the WAD goats were fed on (fresh and green) grasses consisting of Guinea grass (*Panicum maximum*) and Elephant grass (*Pennisetum purpureum*), which is the usual practice in this eco-zone. Water was also made available *ad libitum*.

## **Experimental Drugs and Administration**

Chemotherapeutic drugs used include Acyclovir tablet 200 mg (Virest 200<sup>®</sup>) manufactured by HOVID Bhd., Malaysia. It was administered orally (Per.Os) to the animals for 5 days. Oxytetracycline Hydrochloride Short acting (Oxy-pet<sup>®</sup>/SA) manufactured by Shanghai Gongyi Vet Medicine Plant, Shanghai (People's Republic of. China). It was administered intramuscularly at the dose of 20 mg of the drug per kg of body weight for five days and Ivermectin (Kepromec<sup>®</sup>) manufactured by KEPRO B.V., Maagdenburgstraat (Holland) was used for treatment at 1ml per 25kg and was administered subcutaneously once in two weeks according to their body weight. These treatments were instituted from the 7<sup>th</sup> day, post infection and based on confirmatory PPR diagnosis.

## **Experimental Design**

Twenty WAD goats were randomly assigned into five treatment groups – 1, 2, 3, 4 and 5, of four goats per group. Each goat was duly identified using neck tags: Group 1- The goats were treated with Acyclovir, Oxyte-tracycline and Ivermectin; Group 2- The goats were treated with Acyclovir only; Group 3- The goats were treated with Ivermectin only; Group 4- The goats were

treated with Oxytetracycline hydrochloride only; and Group 5- The goats were left untreated.

## **Experimental Infection**

The WAD goats after a period of stabilization and acclimatization were infected by keeping together in a common pen with goats confirmed by haemagglutination inhibition test and Monoclonal antibody based blocking ELISA to be suffering from PPR infection. This infection was induced according to standard procedures as described by Ugochukwu et al. (6). Three to five days after, all the animals came down with the infection, showing classical clinical signs of PPR. These animals were also subjected to Monoclonal antibody-based blocking ELISA and were all positive for PPR infection.

## **Blood Sample Collection**

Whole blood (2ml) was collected from all experimental animals prior to their separation into groups. This procedure was aseptically done via the jugular vein by venipuncture for serology. The blood samples collected were put in sample bottles not containing anticoagulant and could clot and later centrifuged at 3000 rpm for 15 minutes. The sera obtained were harvested and stored in a deep freezer prior to the start with the serological analysis.

## **Physiological Parameters**

The major physiological parameters, namely, temperature, pulse rate, heart rate and respiratory rate were obtained using standard procedures as described by Chakrabarti (22) and Ugochukwu (6). These physiological parameters were obtained in the mornings throughout the experiment.

## **Serological Diagnosis**

Monoclonal Antibody-Based Blocking Enzyme-Linked Immunosorbent Assay was done by the following procedure and using commercially available Enzyme-linked immunosorbent assay (ELISA) kit (CIRAD-EMVT, Montpellier, France) for PPRV. This diagnostic kit detects antibodies directed against the nucleoprotein of PPRV and was developed by a FAO reference laboratory, CIRAD-EMVT, Montpellier, France. The reaction was stopped after 10 minutes with 50  $\mu$ l of 1 M sulphuric acid. Results were read on an ELISA reader at 492 nm. The absorbance was converted to percentage inhibition (PI). Sera showing PI greater than 50% were regarded as positive. This procedure was done according to standard procedures as described by Saliki *et al.* (23) and Munir *et al.* (24).

## Survivability and Clinical Signs

The goats were observed once daily throughout the experimental period for clinical signs of the disease (e.g., palpable lymph node, body condition/weight changes, appetite, mucus membrane colour, ocular discharges, nasal discharges, etc.) as well as mortality and these were recorded.

## **Statistical Analysis**

The data obtained were computed into means and analysed using Analysis of Variance (ANOVA). The means were separated using Duncan's multiple range tests at 5% level of probability. The results were presented as tables and charts.

## RESULTS

### **Mortality Rate**

Table 1 shows the percentage mortality rate of the different groups. Percentage mortality per group was: Group 1, 25%; Group 2, 75%; Group 3, 75%; Group 4, 75%; and Group 5, 100%. Additionally, Groups 2, 3 and 4 had comparable mean mortality rates across the groups.

### Temperature

Table 2 shows the mean rectal temperature of the different groups. There was no significant difference (P>0.05) across the groups in week 0 and week 1. In weeks 2 and 3, Groups 1, 2, 3 and 4 had a comparable mean temperature and the mean temperatures were significantly lower than (P<0.05) Group 5 (untreated) in week 2.

It was observed that the rectal temperatures in the WAD goats were within physiological range  $(38^{4}40^{\circ}C)$  in Groups 1, 2, 3 and 4 throughout the experimental period. Also, Group 5, in weeks 0 and 1, had temperatures within the physiologic range, but, in week 2, there was an increase in the temperature value suggesting pyrexia.

#### **Pulse Rate**

The mean pulse rates of the different groups are shown in Table 3. There was no significant differences (P>0.05) across all the groups in week 0 and week 1. In week 2, Groups 1, 2, 3 and 4 had a comparable mean pulse rate and were significantly (P<0.05) higher than the mean pulse rate for Group 5 (untreated).

In week 3, the mean pulse rate for Group 2 (treated with Acyclovir only) was significantly (P<0.05) higher than Group 5 (untreated). Also, there was a significant difference (P<0.05) in Groups 1, 2 and 3 although they had a comparable mean pulse rate across the groups.

At weeks 0 and 1, the pulse rates across the groups were within the physiologic range (70-90), except for Groups 2 and 3, in week 0, which had slight increase, but, from week 2 throughout the experimental period, the pulse rates were higher than normal in Groups 1, 2, 3 and 4. Group 5 showed pulse rate within the normal range in week 1, but decreased drastically throughout the experiment.

## **Heart Rate**

Table 4 shows the mean heart rate of the different groups. There was no significant difference (P>0.05) across the groups in week 0 and week 1. In week 2, there was a significant increase (P<0.05) in Groups 1, 2, 3 and 4 when compared with Group 5 (untreated) groups. In week 3, Group 2 (treated with Acyclovir only) was

DAY	GROUP 1	GROUP 2	GROUP 3	GROUP 4	GROUP 5
0	0/4	0/4	0/4	0/4	0/4
7	0/4	0/4	0/4	0/4	2/4
11	0/4	2/4	0/4	0/4	3/4
13	0/4	2/4	3/4	0/4	3/4
15	0/4	2/4	3/4	2/4	3/4
17	0/4	3/4	3/4	2/4	3/4
18	1/4	3/4	3/4	2/4	4/4
19	1/4	3/4	3/4	3/4	4/4

## Table 1. Survivability and mortality of WAD goats infected naturally with PPR and treated with different chemotherapeutic regimens

Figures represent number of mortality/number of animals in the group Source: own work

## Table 2. Mean±SD of temperature (OC) inWAD goats infected naturally with PPR and treated with different chemotherapeutic regimens

Week	Group 1 (combined)	Group 2 (Acyclovir)	Group 3 (Ivermectin)	Group 4 (Oxytetracycline)	Group 5 (Untreated)
0	38.33 ± 0.25ª	38.23 ± 0.32ª	38.42 ± 0.30ª	37.84 ± 0.16ª	38.28 ± 0.15ª
1	38.58 ± 0.26ª	38.07 ± 0.32ª	37.86 ± 0.20ª	38.44 ± 0.16ª	38.29 ± 0.31ª
2	38.31 ± 0.20ª	38.88 ± 0.45ª	38.89 ± 0.23ª	38.58 ± 0.27ª	40.84 ± 0.47 <sup>b</sup>
3	37.65 ± 0.31 <sup>b</sup>	39.60 ± 0.15 <sup>b</sup>	38.50 ± 0.31 <sup>b</sup>	38.25 ± 0.18 <sup>b</sup>	$0.00 \pm 0.00^{a}$

Superscript<sup>a and b</sup> represent the order of significance. Source: own work

## Table 3. Mean ± SD of pulse rates (beats/minute) inWAD goats infected naturally with PPR and treated with different chemotherapeutic regimens

Week	Group 1 (combined)	Group 2 (Acyclovir)	Group 3 (Ivermectin)	Group 4 (Oxytetracycline)	Group 5 (Untreated)
0	87.71±5.10ª	93.71 ± 5.10ª	92.57 ± 5.97ª	89.14 ± 5.08ª	83.35 ± 5.66ª
1	84.86 ± 5.56ª	76.86 ± 2.98ª	87.24 ± 4.77ª	84.66 ± 3.30ª	85.43 ± 8.30ª
2	95.14 ± 6.47 <sup>b</sup>	93.71 ± 4.77 <sup>b</sup>	94.86 ± 5.07 <sup>b</sup>	103.71 ± 4.91 <sup>b</sup>	66.67 ± 10.06ª
3	102.50 ± 3.59ªb	123.00 ± 18.00 <sup>b</sup>	$108.00 \pm 8.04^{ab}$	112.00 ± 21.17 <sup>ab</sup>	0.00 ± 0.00ª

Superscripts <sup>a and b</sup> represent the order of significance. Source: own work significantly higher than (P<0.05) Group 5 (untreated). Also, there was a significant difference (P<0.05) in Groups 1, 2, 3 and 4 and had a comparable mean heart rate across the groups.

## **Respiratory Rate**

It was observed that the respiratory rates across the groups were higher than normal. Normal respiratory rate ranges from 15cpmto 20cpm. Table 5 shows the mean respiratory rate in the different groups. In week 0,

Group 5 (untreated) was significantly higher than (P<0.05) Group 2 (treated with Acyclovir only). Also, Groups 1, 3 and 4 had a comparable mean respiratory rate across the groups. In week 1, Group 2 (treated with Acyclovir only) was significantly higher than (P<0.05) Group 5 (untreated). Also, Groups 1, 3 and 4 had a comparable mean respiratory rate across the groups. In week 2, Groups 1, 2, 3 and 4 was significantly higher than (P<0.05) Group 5 (untreated). In week 3, Groups 1, 2 and 3 was significantly higher than (P<0.05) Group 5 (untreated).

Table 4. Mean±SD of the heart rates (beats/minute) inWAD goats infected naturally with PPR and treated with different chemotherapeutic regimens

Week	Group 1 (combined)	Group 2 (Acyclovir)	Group 3 (Ivermectin)	Group 4 (Oxytetracycline)	Group 5 (Untreated)
0	95.85 ± 5.20ª	106.28 ± 5.31ª	$108.57 \pm 6.70^{a}$	$99.62 \pm 4.40^{a}$	101.81 ± 5.57ª
1	97.71 ± 4.37ª	98.28 ± 5.63ª	100.19 ± 5.94ª	100.14 ± 5.12ª	86.57 ± 2.92ª
2	107.05 ± 10.12 <sup>b</sup>	$119.71 \pm 8.94^{\rm b}$	$126.29 \pm 9.64^{ m b}$	117.71 ± 4.29 <sup>b</sup>	81.33 ± 4.81ª
3	113.00 ± 9.31ªb	144.00 ± 6.73 <sup>b</sup>	125.00 ± 5.51ªb	120.00 ± 4.32 <sup>ab</sup>	$0.00 \pm 0.00^{a}$

Superscripts <sup>a and b</sup> represent the order of significance. Source: own work

## Table5. Mean±SD of the respiratory rates (cycles/minute) inWAD goats infected naturally with PPR and treated with different chemotherapeutic regimens

Week	Group 1 (combined)	Group 2 (Acyclovir)	Group 3 (Ivermectin)	Group 4 (Oxytetracycline)	Group 5 (Untreated)
0	029.90 ± 1.40ªb	28.19 ± 1.35ª	$32.76 \pm 1.67^{ab}$	$31.24 \pm 1.45^{ab}$	$33.52 \pm 1.81^{b}$
1	31.43 ± 2.63ªb	35.14 ± 1.65 <sup>b</sup>	$31.90 \pm 2.16^{ab}$	33.23 ± 1.46ªb	28.57 ± 2.03ª
2	31.19 ± 2.14ª	30.29 ± 2.29ª	37.14 ± 4.16ª	27.57 ± 2.40ª	13.14 ± 6.69ª
3	32.50 ± 1.71 <sup>b</sup>	30.50 ± 2.99 <sup>b</sup>	$34.00 \pm 2.58^{ab}$	25.00 ± 1.00ªb	$0.00 \pm 0.00^{a}$

Superscripts <sup>a and b</sup> represent the order of significance. Source: own work

## **Clinical Signs and Post-Mortem Lesions**

During the experiment, other clinical signs of PPR were observed. The clinical signs initially observed were respiratory distress, ocular and nasal discharges as shown in Figure 1 and 2, lacrimation, pyrexia, dehydration, anorexia and terminally, diarrhoea as shown in Figure 3. Meanwhile, other clinical signs later developed includ-

Figure 1. Catarrhal nasal discharges



Source: own work

ing ulcerative stomatitis, gingivitis and labial Orf-like lesions as shown in Figure 4.

Furthermore, some Post-mortem lesions were observed including oral ulcerative lesions, gastro-enteritis and the characteristic discontinuous streaks of congestion, also called Zebra-markings, in the colon and rectum as shown in Figure 5.

Figure 2. Ocular discharges



Source: own work

#### Figure 4. Labial Orf-like lesions

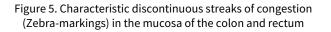


Source: own work



Figure 3. Diarrhoea (observed at the terminal stage of PPR)

Source: own work





Source: own work

## DISCUSSION

The results from this experiment showed that the mortality rate of West African Dwarf (WAD) goats infected with Peste des petits ruminants without supportive therapy and/or treatment was high, which is in agreement with the reports by Kitching, (25) and Jaisree *et al.* (26). Clinically, the affected animals showed signs of pyrexia, anorexia, emaciation, catarrhal nasal discharge, severe lacrimal and ocular discharges, conjunctival encrustation in the medial canthus ulcerative stomatitis and severe diarrhoea. These symptoms agree with those described by the FAO (27), Wolhsein and Saliki (28), Islam *et al.* (29), Abubakar and Irfan (5) and Ugochukwu *et al.* (30) in goats with this disease.

It is also worth noting that the findings from this study is consistent with the report of Blood *et al.* (31), who observed that the disease was very fatal in young goats (60-80%) especially in the age group of 7-12 months. This occurrence is observed in Groups 2, 3, 4 and 5, since these groups showed a mortality rate range between 75%-100%, with the untreated infected PPR goats having 100% mortality rate. This fact agrees with the findings by Dhar *et al.* (1) and Das *et al.* (32).

The findings of this experiment are in tandem with the report by Gupta *et al.* (33), who reported that broad spectrum antibiotics (especially Oxytetracycline and Sulphadimidine) when administered parenterally, have therapeutic action against PPR infection, which suggests taking preventive measures in terms of vaccination and supportive therapy with oxytetracycline will help to effectively control the disease.

According to Jackson and Cockcroft (34) and Radostits *et al.* (35) the per-rectum temperature was within normal reference values. This is in accordance with the initial findings of this work but, however, later on, there was a significant increase in the mean temperature in the untreated group, suggesting pyrexia associated with the disease as reported by Lefevre and Diallo (36). It can thus be deduced that the chemotherapeutic agents (Acyclovir, Oxytetracycline short-acrting Hydrochlo-

ride and Ivermectin) had effects on the treated groups by stabilizing the temperature to remain within the normal range.

The heart rate, pulse rate and respiratory rate were also high, deferring from the normal reference values reported by Jackson and Cockcroft (34), Radostits *et al.*, (35), but this occurrence agrees with the report by Naznin *et al.* (37), who reported that the heart rate, pulse rate and respiratory rate in PPR affected goats are high. It could be postulated, also, that the significant increase in the respiratory rate was due to harsh environmental and weather conditions as well as the presence of the virus.

Ivermectin, a veterinary drug, was incorporated into the combined treatment regimen to kill internal helminth parasites that were observed in some of the goats suffering from PPR used in this experiment. It agrees with findings by Crump and Omura (38). They stated the efficacy of ivermectin, "wonder drug" against external and internal parasites.

Naznin *et al.* (37) reported that the recovery rate from clinical signs was higher in goats treated with parenteral administration of oxytetracycline but as seen in the findings of this work, it was observed that the combined therapy using Acyclovir, Oxytetracycline and Ivermectin, had a better efficacy than the other groups treated singly. The recovery rate of the combined therapy and the single-Oxytetracycline treated groups were comparable one week post-treatment. It can be deduced that the Oxytetracycline effects were against bacteria becoming opportunistic due to the immunosuppressive effects of the virus. It was also postulated that the combined therapy group also symptomatically combated opportunistic bacterial infection (Oxytetracycline) as well as the PPR virus (Acyclovir) and parasitic infections (Ivermectin) that may complicate the disease condition. It provides a platform for rationale for survivability and lower mortality rate in this group.

It was reported by Nawathe (39) that the use of chemotherapeutic agents combined with hygiene, sanitation, isolation, and quarantine of the infected goats had positive effects in managing the disease. The present findings of this work are in agreement with this report. Also, this work is in tandem with the report by Islam *et al.* (2012) and Bupasha *et al.* (40) who investigated and reported the efficacy of Oxytetracycline in managing PPR infection.

## Conclusions

Acyclovir has not been widely used in PPR cases, but it is proven in this work that it produced positive efficacy when combined with Oxytetracycline and Ivermectin as palliative or supportive therapy. This in turn, may allow for immune responses and antibody build up which might lead to recovery and resistance of the goats against the virus.

Combination of Acyclovir, Oxytetracycline and Ivermectin is effective in achieving recovery from clinical signs when compared to the use of these chemotherapeutic agents singly. The presence of the virus had a significant effect on the temperature, pulse rate and respiratory rate. It also caused high mortality in untreated PPR infected goats.

A prolonged study should be done to determine the point of full recovery post-treatment. Although not specific for Paramyxoviruses, it is recommended that more investigations and assessments should be done on the use and efficacy of Acyclovir in the treatment of PPR.

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## **ETHICAL CONSIDERATIONS**

The ethical conditions governing the use and conduct of experiments on live animals were strictly observed and the experimental protocol was approved by the University of Nigeria, Nsukka Senate Committee on Medical and Research Ethics.

## **CONFLICT OF INTEREST**

None of the authors of this paper has a financial or personal association with any organizations that could inappropriately create bias to the content of this research work.

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## References

- Dhar P, Sreenivasa BP, Barrett T, Corteyn M, Singh RP, Bandyopadhyay SK. Recent epidemiology of Peste des petitsruminants virus (PPRV). Vet Microbiol. 2002;88(2):153-59. https://doi.org/10.1016/S0378-1135(02)00102-5
- Ugochukwu ICI, Ezeasor CK, Agina OA, Anyogu DC, Chukwudi IC, Idoko SI, Ugochukwu EI. Peste des petits ruminants: aetiology, pathology, immunology, disease status in Africa, diagnosis, control, prevention and treatment: a review. Not Sci Biol. 2019a;11(1):12-20. https://doi.org/10.15835/nsb11110355
- 3. Shamaki D, Olaleye OD, Obi TU, Diallo A, Majiyagbe KA, Lombin LH, Barrett T. Peste des petits ruminants

(PPR) in Nigeria; serological and molecular epidemiology. Vom J Vet Sci. 2004;1(1):8-23.

- Ozkul A, Akca Alkan F, Barrett T, Karaoglu T, Dagalp SB, Anderson J, Yesilbag K, Cokcaliskan C, Gencay A, Burgu I. Prevalence, distribution, and host range of Peste des petits ruminants virus, Turkey. Emerg Infect Dis. 2002;8(7):708-12. https://doi.org/10.3201/ eid0807.010471
- Abubakar M, Irfan M. An overview of treatment options to combat Peste des petits ruminants in endemic situations. Res J Vet Pract. 2014;2(15):4-7. https://doi. org/10.14737/journal.rjvp/2014/2.1s.4.7
- Ugochukwu ICI, Ugochukwu EI, Chukwu CC. Haematological parameters and serum biochemical assay of West African dwarf goats infected with Peste des petits ruminants virus in Nsukka, Enugu State. Comp Clin Pathol. 2018;27(1):13-9. https://doi.org/10.1007/ s00580-017-2545-9
- Ugochukwu ICI, Idoko SI, Omeke JN, Aneke CI, Sani NA, Emejuo NT, Ikem CF, Ugochukwu EI. The clinicopathological evaluations of the efficacy of incorporation of amantadine hydrochloride in the treatment of West African Dwarf Goats experimentally infected with Peste des petits ruminants virus. Not Sci Biol. 2019b;11(4):379-86. https://doi.org/10.15835/ nsb11410526
- Barrett T, Pronab D, Sreenivasa BP, Corteyn M. Recent epidemiology of Peste des petits ruminants virus (PPRV). Vet Microbiol. 1997;88:125-30.
- Kerur N, Jhala MK, Joshi CG. Genetic characterisation of Indian Peste des petits ruminants virus (PPRV) by phylogenetic analysis of fusion protein and nucleoprotein gene segments. Res Vet Sci. 2008;85:176-83. https://doi.org/10.1016/j.rvsc.2007.07.007
- Berguido FJ, Bodjo SC, Loitsch A, Diallo A. Specific detection of Peste des petits ruminants virus antibodies in sheep and goat sera by the luciferase immunoprecipitation system. J Virol Met. 2016;227:40-6. https://doi. org/10.1016/j.jviromet.2015.10.008
- Ezeibe MCO, Okoroafor ON, Ngene AA, Eze JI, Eze IC, Ugonabo JAC. Persistent detection of Peste des petits ruminants antigen in the faeces of recovered goats. Trop Anim Hlth Prod. 2008;40:517-19. https:// doi.org/10.1007/s11250-008-9128-3

- Balamurugan V, Hemadri D, Gajendragad MR, Singh RK, Rahman H. Diagnosis and control of Peste des petits ruminants: a comprehensive review. Virus Dis. 2014;25(1):39-56. https://doi.org/10.1007/s13337-013-0188-2
- 13. Perry BD, Randolph TF, McDermott JJ, Sones KR, Thornton PK. Investing in animal health research to alleviate poverty. Nairobi, Kenya: ILRI; 2002.
- Asim M, Rashid A, Chaudhary AH, Noor NS. Production of homologous live attenuated cell culture vaccine for the control of Peste des petits ruminants in small ruminants. Pak Vet J. 2009;29(2):72-4.
- Ogunsanmi AO, Awe EO, Obi TU, Taiwo VO. Peste des petits ruminants (PPR) virus antibodies in African Grey Duiker (*Sylvicapra grimmia*). Afr J Biomedl Res. 2003;6(1):59-61. https://doi.org/10.4314/ajbr. v6i1.54025
- Libeau G, Diallo A, Parida S. Evolutionary genetics underlying the spread of peste des Petits ruminants virus. Anim Front. 2014;4:14-20. https://doi.org/10.2527/ af.2014-0003
- Stephenson I, Wiselka M. Drug treatment of tropical parasitic infections: recent achievements and developments. Drug. 2000;60(5):985-95. https://doi. org/10.2165/00003495-200060050-00002
- Şevik M, Sait A. Genetic characterization of peste des petitsruminants virus, Turkey, 2009-2013. Res Vet Sci. 2015;101:187-95. https://doi.org/10.1016/j. rvsc.2015.05.005
- Diallo A. Peste des petits ruminants (PPR) virus. Int J Anim Sci. 2003;15(2):171-73.
- Luka PD, Erume J, Mwiine FN, Ayebazibwe C. Seroprevalence of peste des petits ruminants antibodies in sheep and goats after vaccination in Karamoja, Uganda: implication on control. Int J Anim Vet Adv. 2011;3(1):18-22.
- Sen A, Saravanan P, Balamurugan V, Rajak KK, Sudhakar SB, Bhanuprakash V, Parida S, Singh RK. Vaccines against Peste des petits ruminants virus. Expert Rev Vaccines. 2010;9(7):785-96. https://doi. org/10.1586/erv.10.74
- 22. Chakrabarti A. Textbook of clinical veterinary medicine. 3<sup>rd</sup> Ed. New Delhi: Kalyani Publishers; 2006.

- 23. Saliki JT, Libeau G, House JA, Charles A, Mebus CA, Dubovi EJ. Monoclonal antibody-based blocking enzyme-linked immunosorbent assay for specific detection and titration of peste-des-petits-ruminants virus antibody in caprine and ovine sera. J Clin Microbiol. 1993;31(5):1075-82. https://doi.org/10.1128/ JCM.31.5.1075-1082.1993
- Munir M, Abubakar M, Zohari S, Berg M. Serodiagnosis of Peste des petits ruminants virus. In: Al-Moslih M, editor. Serological diagnosis of certain human, animal and plant diseases, Vol. 1. Croatia: InTech Publishers; 2012. p. 37-58. https://doi.org/10.5772/36464
- Kitching RP. The economic significance and control of small ruminant viruses in North Africa and West Asia. In: Thomson EF, Thomson FS, editors. Increasing small ruminant productivity in semi-arid areas. Dordrecht, Netherlands: Kluwer Academic Publishers; 1988. p. 225-236. https://doi.org/10.1007/978-94-009-1317-2\_19
- 26. Jaisree S, Hemalatha S, Muthuramalingam T, Manimaran K, Mahaprabhu R, Tensingh Gnanaraj P, Kumanan K, Roy P. Investigation on outbreak of Peste des petits ruminants (PPR) in an organized farm among Tellicherry breed of goats. Int J Livest Res. 2017;7(1):100-6. https://doi.org/10.5455/ijlr.20170104095534
- 27. FAO. Recognising Peste des petits ruminants. A field manual. EMPRES (Livestock) Animal Health Service FAO Animal Production and Health Division. Rome, Italy: 1999.
- Wolhsein P, Saliki J. Rinderpest and Peste des petits ruminants-the diseases: clinical signs and pathology. In: Barrett T, Pastoret PP, Taylor WP. editors. Rinderpest and Peste des Petits ruminants: virus plagues of large and small ruminants. Elsevier; 2006. p. 68-85. https://doi.org/10.1016/B978-012088385-1/50034-4
- 29. Islam MS, Khan MSI, Kader HA, Begum MR, Asgar MA. Prevalence of PPR of goat and their response to antibiotic treatment at Mirzaganj Upazila of Patuakhali Distrtict. J Envirom Sci Nat Res. 2012;5(2):181-4. https://doi.org/10.3329/jesnr.v5i2.14811
- 30. Ugochukwu ICI, Ugochukwu, EI, Chukwu CC. Therapeutic response of West African Dwarf goats infected with Peste des petits ruminants whose oral

lesions were treated with oxytetracycline long acting and gentian violet topical spray. Asian Paci J Trop Dis. 2017;7(12):771-4. https://doi.org/10.12980/ apjtd.7.2017D7-234

- Blood DC, Radostits, OM, Gay CC. Veterinary medicine. 8<sup>th</sup> ed. UK: Balliere Tindall; 1995.
- 32. Das KK, Shil NK, Islam MR. Sero-epidemiological investigation on Peste des Petits ruminants in black Bengal goats. Bangl J Microbiol. 2007;(2):143-5. https://doi.org/10.3329/bjm.v24i2.1260
- Gupta SD, Biswas PK, Habib S, Debnath NC. Prevalence of PPR in the greater Chittagong district of Bangladesh. Int J Sust Agri Techno. 2007;3(2):36-41.
- Jackson PGG, Cockcroft PD. Clinical examination of the respiratory system. In: Jackson PGG, Cockcroft PD, editors. Clinical examination of farm animals. Oxford: Blackwell Science Ltd.; 2002. p. 320-325. https://doi.org/10.1002/9780470752425
- 35. Radostits OM, Gay CC, Hinchcliff KW, Constable PD. Veterinary medicine: A textbook of the diseases of cattle, horses, sheep, pigs, and goats. 10<sup>th</sup> ed. Philadelphia: Saunders; 2007.
- Lefevre PC, Diallo A. Peste des petits ruminants. Revue Scientific et Technique. 1990;(9):951-65. https://doi.org/10.20506/rst.9.4.532
- 37. Naznin M, Ahaduzzaman M, Chowdhury S, Biswas PK. Prevalence and clinico-pathological parameters of PPR infected goats and their response to antibiotic treatment at Panchlaish, Chittagong. Bangl Int J Nat Sci. 2014;4(2):1-7. https://doi.org/10.3329/ijns. v4i2.28597
- Crump A, Omura S. Ivermectin, 'Wonder drug' from Japan: the human use perspective. Proceedings of Japan Academy Series B Physical and Biological Sciences. 2011;87(2):13-28. https://doi.org/10.2183/pjab.87.13
- 39. Nawathe DR. The control of peste des petits ruminants in Nigeria. Prev Vet Med. 1984;(2):147-55. https://doi. org/10.1016/0167-5877(84)90058-8
- 40. Bupasha ZB, Hossain F, Sarker MS, Ahaduzzaman M, Biswas PK. Variability in prevalence and therapeutic effectiveness in PPR affected goats of Thakurgoan, Bangladesh. AVAS. 2015;2(1):15-9.