

Scientific Article http://doi.org/10.31910/rudca.v25.n2.2022.2312

Influence of microbial consortia on the incidence of grey mold (*Botrytis cinerea*) in strawberry (Monterey variety)

Influencia de consorcios microbianos en la incidencia del moho gris (*Botrytis cinerea*) en fresa (variedad Monterey)

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How to cite: Cano, M.A.; Cuervo, J.L.; Darghan, A.E. 2022. Influence of microbial consortia on the incidence of grey mold (Botrytis cinerea)

in strawberry (Monterey variety). Rev. U.D.C.A Act. & Div. Cient. 25(2):e2312. http://doi.org/10.31910/rudca.v25.n2.2022.2312

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Official publication of the Universidad de Ciencias Aplicadas y Ambientales U.D.C.A, University, Accredited as a High-Quality Institution by the Colombian Ministry of Education.

Received: October 21, 2021

Accepted: November 15, 2022

Edited by: Helber Adrián Arévalo Maldonado

ABSTRACT

Botrytis cinerea, the causal agent of grey mold disease, is one of the most destructive pathogens of strawberry crops, both in vegetative development and postharvest. The control of this pathogen is complex due to its aggressiveness and ability to attack and infect various plant tissues and is mainly based on chemical control; however, the incorrect use of pesticides, mainly due to overdosing, causes the presence of traces of these agrochemicals in the fruits, as well as the selection of pathogen resistance to fungicides, making it a risk to human health and the environment. The objective of the study was to use biological regulation strategies, with the application of microbial consortia made up of mycorrhizal fungi, antagonistic bacteria and Trichoderma harzianum, as an alternative for the management of grey mold in strawberry crops (Monterey variety) under field conditions. Treatments T4 (mycorrhizal fungi), T8 (mycorrhizal fungi, antagonistic bacteria and T. harzianum) and T2 (T. harzianum) presented the lowest incidence of the pathogen with 2.6, 3.1 and 3.6 %, respectively, compared to control plants with 16.6%. The influence of all biological treatments on the regulation of B. cinerea was greater than the control.

Keywords: *Botrytis cinerea*; Biological regulation; Microbial consortia; Mycorrhizae; Antagonistic bacteria; *Trichoderma harzianum*.

Botrytis cinerea, el agente causal de la enfermedad del moho gris, es uno de los patógenos más destructivos del cultivo de fresa, tanto en el desarrollo vegetativo como en poscosecha. El control de este patógeno es complejo, debido a su agresividad y capacidad de atacar e infectar diversos tejidos de la planta y se basa, principalmente, en el control químico; sin embargo, el uso incorrecto de plaguicidas, principalmente por sobredosificación, provoca la presencia de trazas de estos agroquímicos en los frutos, así como la selección de resistencia del patógeno a los fungicidas, convirtiéndolo en un riesgo para la salud humana y el ambiente. El objetivo del estudio fue utilizar estrategias de regulación biológica, con la aplicación de consorcios microbianos, conformados por hongos micorrícicos, bacterias antagonistas y Trichoderma harzianum, como alternativa para el manejo del moho gris, en cultivos de fresa (variedad Monterey), en condiciones de campo. Los tratamientos T4 (hongos micorrízicos), T8 (hongos micorrízicos, bacterias antagonistas y T. harzianum) y T2 (T. harzianum) presentaron la menor incidencia del patógeno, con 2,6, 3,1 y 3,6 %, respectivamente, en comparación con las plantas control, con 16,6 %. La influencia de todos los tratamientos biológicos en la regulación de B. cinerea fue mayor respecto al control.

RESUMEN

Palabras clave: *Botrytis cinerea*; Regulación biológica; Consorcios microbianos; Micorrizas; Bacterias antagonistas; *Trichoderma harzianum*.

INTRODUCTION

Strawberry (*Fragaria* × *ananasa*) is a crop with high economic, nutritional, medicinal and culinary value (Lantz *et al.* 2010; PTP *et al.* 2013), to the point that it is considered a nutraceutical product, because of its high content of flavonoids, anthocyanins, phenolic compounds, and vitamins (Cao *et al.* 2011). Between 2015 and 2020, the area planted in strawberry crops in Colombia increased by 59 %, going from 1,656 hectares to 2,638, with an increase in production from 55,719 tons to 86,534, respectively (Minagricultura, 2021).

However, its production has a negative environmental impact because it depends on frequent applications of chemical fertilizers and products such as insecticides, fungicides, and herbicides, since it is highly susceptible to pathogens (Pritts, 2002; Vestberg et al. 2004). Edaphic pathogens such as Phytophthora, Pythium, Rhizoctonia, and Fusarium attack the crop. Other fungal diseases such as soft rot (Rhizopus stolonifer) and black rot (Mucor spp., Aspergillus niger, and Pythium spp.) are also common in postharvest. Anthracnose is also one of the major diseases caused by several species of the genus Colletotrichum, including C. acutatum, C. fragariae, and C. gloeosporioides (Chalfoun et al. 2011). These diseases have great importance because they can affect all plant tissues, such as stolons, crowns, stems, leaves, flowers, and fruits (Freeman & Katan, 1997), severely affecting productivity and profitability. Similarly, grey mold, caused by Botrytis cinerea, is one of the most destructive diseases during crop development and postharvest, causing severe economic losses, estimated at around 30 % of total production (Zhang et al. 2007). Even in postharvest, this pathogen is even more aggressive, affecting 95 % of the fruits 48 hours after harvesting (Li et al. 2019).

Traditionally, diseases caused by fungi are controlled with synthetic fungicides. However, there is increasing concern over the possible harmful effects these fungicides may have on the environment and human health. For the specific case of grey mold control, farmers use, among other strategies, chemical control with the application of the following active ingredients: Captan, Chlorothalonil, Iprodione, Procymidone, Fenhexamid, Diclofluanid, Tebuconazole, Methyl thiophanate, and Pyrimethanil (Fillinger *et al.* 2008, Leroux *et al.* 2010; FRAC, 2013). However, *B. cinerea* has a high capacity to become resistant to fungicides.

According to the above, the search for biological alternatives, such as the use of antagonistic microorganisms and/or biological control agents (BCA), is relevant for sustainable agriculture systems in order to achieve safe products (Guédez *et al.* 2009). Various studies have proven the individual effectiveness of some microorganisms used as biological control agents, such as fungi of the genus *Trichoderma*, arbuscular mycorrhizal fungi (AMF) and plant growth promoting bacteria (PGPB's) antagonistic bacteria. However, research using functional mixtures of microorganisms to regulate pathogen populations and seek positive effects on crop yield and quality under field conditions have not been sufficient, due to the very complexity of soil-plant-microorganism-ambient interactions.

For the above the purpose of this research was to evaluate the effect of microbial consortia on the incidence of grey mold in strawberry under field conditions.

MATERIALS AND METHODS

Area location and time of the study. The experiment took place on the campus of the Universidad de Ciencias Aplicadas y Ambientales, U.D.C.A, at the research and teaching center "El Remanso". The center is located at an altitude of 2560 m a.s.l, with geographic coordinates of N 4°35 W 74°04. Using a freely exposed production system, with 8 raised beds, 0.6 m width per bed, 0.4 meters between furrows, and 41.22 m long, plants were planted with three 0.3 m arrangements.

Agroclimatic data. Data reported by the Instituto de Hidrología, Meteorología y Estudios Ambientales (IDEAM) at the time that the experiment was developed, showed an average temperature of 14.1 °C, and precipitation of 506.5 mm; however, the high level of rainfall was during the experiment, reporting more than 50 % of the precipitation for that year. This increased the relative humidity, which registered values between 80 and 90 %.

Vegetable material and field distribution. In total, 1920 strawberry plants Monterey variety were used, distributed in eight blocks, and each block had eight treatments with 30 plants per treatment (Table 1).

Cultivation management. The agronomic management of the crop was focused on a clean production system, which completely restricted the use of chemicals for phytosanitary protection, instead, the cultural practices included phytosanitary pruning, with proper management of the remains of the infected plants, which were removed from the crop immediately and transferred to covered compost to decrease the inoculum of the infection and the dispersion of the pathogens. In addition, microbial inoculants were added at the time of transplantation, and re-inoculation was done three months after planting as a strategy for the regulation of the pathogens.

The cultivation system contemplated the elevation of beds or furrows, which were covered with mulch/plastic to reduce the presence of pathogens and weeds as is done in commercial crops on the savanna of Bogota. The irrigation system that was implemented used dripping, which was also used for fertilization. The first fertilization was done three months after sowing and for the study area, according to the soil analysis, with a recommended application of 533 g of Nitrogen (N), 666 g of Phosphorus (P) and 930 g of Potassium (K).

However, the fertilization was reduced to 50 % of the recommended dose because of the inoculation of biological products that are

Treatment	Block	Randomization within blocks							
T1- Control (uninoculated plants)	Ι	T1	T6	Т3	T5	Т8	T4	T2	T7
T2 Trichoderma (T)	II	Т3	T1	Т5	Т8	Т7	T6	T4	T2
T3 Bacteria (B)	III	T3	T8	T6	T2	T5	T1	Τ7	T4
T4 Mycorrhizal fungi (M)	IV	Т5	T1	Т6	Т8	T4	T2	Т7	Т3
T5 (T+B)	V	T8	T4	Τ7	T5	Т3	T6	T2	T1
T6 (T+M)	VI	T2	T5	Т3	Τ7	Т6	T4	T1	T8
T7 (B+M)	VII	T6	T7	T4	Т8	T1	T5	T2	T3
T8 (T+B+M)	VIII	T8	T3	Т2	Τ7	Т6	T5	T4	T1

Table 1. Treatments and disposition of them in the field.

sensitive to optimum levels of fertilizers. In addition, the inclusion of biofertilization practice (the inoculums used as biological control agents, are also known as biofertilizers) decreases the demand for synthetic fertilizers by optimizing soil nutrient mineralization and plant availability (Li *et al.* 2017; Mondal *et al.* 2017). Finally, 250 g of N, 300 g of P and 400 g of K, applications were prepared in a mixture of 11 L of water and applied for five minutes once a week for four weeks. The subsequent fertilization was done six months to the ninth month (after sowing) increasing the dose of calcium nitrate to 300 g (N), 380 g of diammonium phosphate (P) and 480 g of potassium nitrate (K), once a week for 4 weeks.

Experimental inoculum. Microbial inoculants and their bioregulatory effect on pathogens are listed in table 2. The selection of microbial inoculants and consortia and the mixture of these were carried out to potentiate the beneficial effects for plant growth and protection against *B. cinerea*, knowing the multifunctionality as plant growth biostimulants and control agents biological of each of the microorganisms that made up the mixtures. However, the mixture was also made to observe the compatibility and/or functional complementarity of these microorganisms. Each plant per treatment was inoculated with 100 mL of solution plus an adjuvant (Agrotin[®]) 2 cm³L⁻¹ at the time of transplantation and three months after the establishment of the crop.

Presence or absence of grey mold. The incidence of grey mold disease was evaluated in all plants, 90 days after transplantation. Only the incidence of the disease in fruits, as these are the main organ of interest in the production, was taken into account for the collection of the information, and for marking the plants according to diagnosis (diseased). The presence of at least one fruit affected by the disease was considered a diseased plant.

Botrytis cinerea natural inoculum. B. cinerea was not introduced to the crop, but was present in the soil as sclerotia, due to the

previous sowing of strawberry Albión variety, during the semester immediately before the experiment, which presented estimated losses of over 30 % due to the presence of grey mold disease.

To monitor the grey mold disease at the time of sampling, the presence of the disease in fruits. After the field count, samples were taken to the vegetable sanitary laboratory in the Universidad de Ciencias Aplicadas y Ambientales U.D.C.A, to perform the microscopic observation of the pathogen and corroborate the presence of the disease in affected fruits.

Experimental design and associated variables. A simple factorial design was used in a randomized complete block design with eight blocks and eight treatments and 30 plants per treatment. The variable response was associated with the presence-absence dichotomy of grey mold and the explanatory variables involved blocking factors and biological treatments. The variables associated with the diagnosis of the disease (presence-absence) were evaluated in addition to the proportion of dead plants that were excluded from the modeling process since some of them died from stress after transplantation. Table 1 shows the treatments and distribution of the treatments. The diagnosis was disaggregated by blocks and treatments.

Statistical analysis. Methodology for data analysis: The explanatory variables in logistic regression can be quantitative or qualitative, that is, factors such as treatments or blocks and covariates associated with physical and chemical properties of soil, air, and water. One of its advantages is that the interpretation of the model is possible by probabilities or disparity coefficients, which are a function of the model parameter (Stokes *et al.* 2012).

The statistical analysis included the descriptive component as the inferential. In the first case, the diagram of the spatial distribution of the plants was constructed according to the diagnosis of the

Microorganisms Composition and dosification		Antagonistic effect				
Mycorrhizal fungi Kuklospora colombiana Glomus manihotis, G. intrarradices, G. etunicatum	825 spores of each mycorrhiza species per pound of product. 100 g of product/25 liters of water	-Competition for space and nutrients.-Induction to systemic resistance in plants.-Changes in the composition of the rizospheric biota because of the mycorrhizal exudates.				
Antagonistic bacteria Azotobacter chroococcum Pseudomonas aureofaciens Bacillus licheniformis B. megaterium B. subtilis	2X10 ¹⁰ UFC viable per gram 1 g per liter of water	 -Competition for space and nutrients. -Induction to systemic resistance in plants. -Production of antibiotics. -Production of lytic enzymes. -Production of siderophores. 				
Antagonistic fungus Trichoderma harzianum	Each gram has 5X10 ⁸ (500 millions) of Con- idiospores viable 2 g of active ingredient per each100 g of formulation	 -Competition for space and nutrients. -Induction to systemic resistance in plants. -Production of siderophores. -Production of lytic enzymes. -Micoparasitism. 				

Table 2. Pathogen bioregulating microorganisms used in the experiment.

disease, discriminated by treatments and blocks. Subsequently, the crossed tables were obtained for the proportion of plants according to diagnosis and the corresponding block, accompanying each proportion with the respective marginal count. A bar chart was then presented for diagnosis only in the levels of presence and absence of the disease by treatments and blocks. In the inferential component, a logistic regression model for the response associated with the diagnosis was adjusted using treatments and blocking as explanatory variables. Finally, the table of proportions predicted or adjusted by the model was constructed for each explanatory variable, values being compared with the observed values were sufficient reason to validate the model. The diagram for visualization of the spatial pattern as well as the adjustment of the logistic regression model was made using free software R. The likelihood ratio test was used to verify the fit of the model, comparing the complete model with the reduced model (only treatments effect).

RESULTS AND DISCUSSION

The presence of grey mold disease in strawberry fruits was evident, fruits collected in the field were taken to the plant health laboratory of the U.D.C.A, to make the respective macro and microscopic observations (Figure 1).

The spatial representation of the disease diagnosis in healthy (green) and diseased (red) levels is seen in figure 2. The blank spaces between furrows and plants of each line correspond to dead plants (which were discarded from all analyzes once detected). The figure 2 fits the actual scheme of the sowing method and the coordinates (x, y) represents the measurements in centimeters.

For the inferential component, a logistic regression model was fitted using treatments and blocks as a factor and, as a response, the presence or absence of grey mold in fruits. The binary response variable (denoted by y), associated with the presence or absence of grey mold was written as a function of two predictor variables (denoted by x and z), namely the type of treatment (x) and the block (z).

Results showed statistical significance for the intercept and the coefficient associated with the treatments (p < 5 %), not for the effect of the blocks (Table 3). Equation 1 was written as:

 $logit[p(y=1)]=-2,47110-0,08398xi+0,06782z_j$,

$$\begin{split} &\ln\left[\frac{p(y=1)}{1\!-\!p(y=1)}\right] =\!\!-2,\!47110\!-\!0,\!08398x_i\!+\!0,\!06782z_j\,,\\ &\frac{p(y=1)}{1\!-\!p(y=1)} =\!\!e^{\!-2,\!47110\!-\!0,08398x_i\!+\!0,06782z_j}\,,\\ &p(y=1)\!=\,\frac{e^{\!-2,\!47110\!-\!0,08398x_i\!+\!0,06782z_j}}{1\!+\!e^{\!-2,\!47110\!-\!0,08398x_i\!+\!0,06782z_j}}\,, \end{split}$$

Therefore, the incidence of the grey mold disease estimated for all blocks and treatments is obtained by substituting the values of i and j associated with the treatments and blocks, respectively. The estimated values by the model next to a visual diagram (Figure 3), were used to rank the treatments according to the response.

In addition to the spatial distribution, the estimation of statistical significance coefficients and distribution of prevalence according to



Figure 1. Presence of grey mold disease in the experimental field. a) Strawberry fruit with symptoms of grey mold; c) and d) microscopic observation of *Botrytis cinerea* conidiophores (40 and 10X).



Figure 2. Spatial distribution of the disease diagnosis (Grey mold), according to treatment and shaped blocks 90 days after sowing.

	Table	3.	Estimated	coefficients	of	the	mode	l and	their	stati	stica	al s	igni	fican	ce
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Coefficients	Estimate	Standard error	Value Z	Pr(> z)
Intercept	-2,47110	0,26458	-9,340	$< 2*10^{-16}***$
Treatments	-0,08398	0,03947	-2,127	0,0334 *
Blocks	0,06782	0,03935	1,724	0,0848 .

diagnosis (in percentage), healthy, diseased and dead plants frequency per treatment was obtained (Figure 3). The values of the model adjustment (proof of the likelihood ratio) yielded a significance value of less than 5 %, placing the reduced model (without block effect) as a better model, as compared to the complete model. This result was consistent with the value of the null deviation and the residual deviation of the model.



Figure 3. Marginal distribution of plant counts according to diagnosis (healthy, diseased, dead) for treatments: T1 Control (C), T2 *Trichoderma* (T), T3 Bacteria (B), T4 Mycorrhizal fungi (M), T5 (T + B), T6 (T + M), T7 (B + M) and T8 (T + B + M).

The size of the circles is proportional to the frequency of the data, so the higher count is associated with healthy plants in both treatments and blocks, and the smaller points are associated with the low presence of grey mold disease. The treated plants were compared to the control in the line of diseased plants.

In figure 3, it is observed the low presence of grey mold disease, noting the graphic variability in the small size of the circles for all the biological treatments applied, with percentages of incidence of the disease in ascending order of: 2.6 (T4); 3.1 (T8); 3.6 (T2); 8.5 (T6); 8.6 (T7); 11.9 (T5), compared with control plants, which were not inoculated, nor treated with any chemical and had the highest percentage of incidence of the disease 16.6 (T1). Similar results were found in grapes, where *B. cinerea* had effective control with using different biological control agents (Pertot *et al.* 2017).

It was observed that the lowest incidence rates of grey mold disease are possibly related to the presence of mycorrhizal fungal consortium (Figure 3). These results may suggest that mycorrhizal fungi are strong competitors for space and nutrients in the rhizosphere (Brimner & Boland, 2003; Selosse *et al.* 2004; Pozo & Azcón-Aguilar, 2007; Avis *et al.* 2008; Wehner *et al.* 2010), are capable of rapidly colonizing the interior of the roots and extending a network of hyphae to the exterior, thus hindering the passage and mobility of pathogens, and also stimulate the plant defense system (Hause *et al.* 2007; Hause & Schaarschmid, 2009). Indirectly, hyphae produce exudates, which can modify the microbiome, and this modulates interactions with other organisms present in the rhizospheric zone (Finlay, 2004).

Regarding the microbial consortium composed of all microorganisms used in this research (T8), the incidence of *B. cinerea* was low at 3.1

%, placing it as the second-best treatment, compared to the control and the other treatments. This result may suggest the existence of functional compatibility of the consortium, in terms of the biological regulation of pathogens. In addition, a certain number of soil microorganisms stimulate the predetermined, and internal protection and defense system of plants, diminishing the biotic or abiotic stress (Kapoor *et al.* 2008).

Even though the results of this research seem so strong, it is difficult to predict the result of the interactions between beneficial plants and soil microorganisms, even more among the species of microorganisms used (Vázquez *et al.* 2000). However, the interactions between microorganisms are complex and synergic effects can be presented that potentiate the benefits for the plant or, on the contrary, antagonistic effects or simply that no effect occurs (Cano, 2011).

According to Bisutti *et al.* (2017), the biological control is complex and even more so with microbial mixtures, elucidating the mode of action is difficult, but microorganisms directly exert a beneficial effect in the promotion of plant growth and the strengthening of plants, against pathogens and adverse environmental conditions.

The use of biological control agents for managing diseases represents a viable alternative to replace or reduce applications of chemical fungicides (Cotes, 2014). In this investigation, the microbial consortia possibly acted jointly, to a different extent, but with positive results regarding the reduced incidence of grey mold disease, for all biological treatments, compared with plants without inoculation. This result may be due to what was suggested by Pertot *et al.* (2017), who points out that the combined strategy of different microbial biological control agents did not differ from the results

when the individual treatments were applied, for the control of *B. cinerea* in grapes, which indicates that the microorganisms do not interfere negatively between them, and they could occupy different ecological niches. Likewise, similar results were presented by Pertot *et al.* (2008), using a different microbial consortium, composed of *Ampelomyces quisqualis T. harzianum* T39 and *Bacillus subtilis*, but to a lesser extent than chemical fungicides; however, biological control agents, reduce the use of chemical fungicides and can be applied alternately as part of an integrated disease management program.

According to the results of this research, the treatment T2 (T. *harzianum*), showed to be the third best treatment, very similar to the treatments T4 and T8 and compared with the control plants. Similar results were reported by Merchán-Gaitán *et al.* (2014), finding that in strawberry plants, the control obtained a greater incidence value of *B. cinerea*, than the treatments in which the biocontrollers (T. *harzianum* and T. *lignorumpor*) were applied (Ventana and Camino Real varieties).

T. harzianum, is one of the most widely distributed biological control agents in the world with a broad spectrum of pathogens that it regulates. It is also an excellent competitor, with high growth rates that surpasses most pathogens *in vitro* and is characterized by having different mechanisms and modes of action for bioregulation such as competition, mycoparasitism, production of lytic enzymes, antibiosis, production of siderophores, and stimulation of plants defense system (Harman *et al.* 2004; Harman, 2006). The results of this research concur with the literature because *T. harzianum* has a wide antagonistic potential that reduced the incidence of *B. cinerea*, compared with the plants that were not inoculated.

In this regard, Yang *et al.* (2009), in laboratory conditions, tested the bioregulatory efficiency of *T. harzianum* on *B. cinerea* isolated from the strawberry crop, obtaining positive results and mentioned that *T. harzianum* possesses a constitutive elicitor protein that acts against *B. cinerea* and other pathogens such as *Rhizoctonia solani* and *Fusarium oxysporum*. In the same way, Guédez *et al.* (2009), under similar conditions, dual tests in the laboratory, observed that the antagonist *T. harzianum* is a good competitor (rapid growth), and possibly its main mechanism of action is mycoparasitism, in common strawberry postharvest pathogens, such as *Rhyzopus stolonifer, Mucor* spp., *Penicillium digitatum, Rhizoctonia solani, Aspergillus niger* and *Pythium* spp.

In contrast to the above, the treatments T5, T6, and T7, even though they had lower percentages of incidence of the disease 11.9, 8.5 and 8.6 %, respectively, compared to the control (T1) 16.6 %, the response was not as noticeable as the other treatments analyzed. It is possible that, as previously mentioned, these mixtures of microorganisms did not present adequate complementation in the regulation of the disease; some of the different microbial consortia members could alter the bioregulatory effect of others, affecting the effectiveness or efficiency of the consortium in the pathogen biological regulation.

Results of this investigation showed that the inoculation and co-inoculation of mycorrhizal fungi, antagonistic bacteria, and *T. harzianum*, interact positively in favor of the plants' health, managing to reduce the incidence of *B. cinerea* in strawberry fruits.

Acknowledgment. This research was supported by the Universidad de Ciencias Aplicadas y Ambientales U.D.C.A. We thank our colleagues from Universidad Nacional de Colombia, who provided technical advice, insight, and expertise that greatly assisted the research, and to every people that made possible the development of this project.

REFERENCES

- AVIS, T.J.; GRAVEL, V.; ANTOUN, H.; TWEDDELL, R.J. 2008. Multifaceted beneficial effects of rhizosphere microorganisms on plant health and productivity. Soil Biology and Biochemistry. 40(7):1733-1740. https://doi.org/10.1016/j.soilbio.2008.02.013
- BISUTTI, I.L.; PELZ, J.; BÜTTNER, C.; STEPHAN, D. 2017. Field assessment on the influence of RhizoVital[®] 42 fl. and Trichostar[®] on strawberries in the presence of soil-borne diseases. Crop Protection. 96:195-203. https://doi.org/10.1016/j.cropro.2017.02.004
- BRIMNER, T.A.; BOLAND, G.J. 2003. A review of the non-target effects of fungi used to biologically control plant diseases. Agriculture, Ecosystems & Environment. 100(1):3-16. https://doi.org/10.1016/S0167-8809(03)00200-7
- CANO, M.A. 2011. Interacción de microorganismos benéficos en plantas: Micorrizas, *Trichoderma* spp. y *Pseudomonas* spp. una revisión. Revista U.D.C.A Actualidad & Divulgación Científica. 14(2):15-31. https://doi.org/10.31910/rudca.v14.n2.2011.771
- CAO, S.; HU, Z.; ZHENG, Y.; YANG, Z.; LU, B. 2011. Effect of BTH on antioxidant enzymes, radical-scavenging activity and decay in strawberry fruit. Food Chemistry. 125(1):145-149. https://doi.org/10.1016/j.foodchem.2010.08.051
- CHALFOUN, N.R.; CASTAGNARO, A.P.; DÍAZ RICCI, J.C. 2011. Induced resistance activated by a culture filtrate derived from an avirulent pathogen as a mechanism of biological control of anthracnose in strawberry. Biological Control. 58(3):319-329. https://doi.org/10.1016/j.biocontrol.2011.05.007
- COTES, A.M. 2014. Control biológico de enfermedades de plantas en Colombia. En: Bettiol, W.; Rivera, M.C.; Mondino, P.; Montealegre, J.R.; Colmenárez, Y.C. (eds). Control bilógico de enfermedades de plantas en América Latina y el Caribe. Universidad de la República. p.169-179.

- 8. FILLINGER, S.; LEROUX, P.; AUCLAIR, C.; BARREAU, C.; AL HAJJ, C.; DEBIEU, D. 2008. Genetic analysis of Fenhexamid-Resistant field isolates of the phytopathogenic fungus Botrytis cinerea. Antimicrobial agents and chemotherapy. 52(11):3933-3940. https://doi.org/10.1128/aac.00615-08
- 9. FINLAY, R.D. 2004. Mycorrhizal fungi and their multifunctional roles. Mycologist. 18(2):91-96. https://doi.org/10.1017/S0269-915X(04)00205-8
- 10. FREEMAN, S.; KATAN, T. 1997. Identification of Colletotrichum species responsible for anthracnose and root necrosis of strawberry in Israel. Phytopathology. 87(5):516-521. https://doi.org/10.1094/phyto.1997.87.5.516

- 11. FUNGICIDE RESISTANCE ACTION COMMITTEE, FRAC. 2013. List of plant pathogenic organisms resistant to disease control agents. FRAC. 71p. Disponible desde Internet en: https://www.frac.info/docs/default-source/working-groups/s dhi-fungicides/group/list-of-resistant-plant-pathogens_2012 -edition.pdf
- 12. GUÉDEZ, C.; CAŃIZÁLEZ, L.; CASTILLO, C.; OLIVAR, R. 2009. Efecto antagónico de Trichoderma harzianum sobre algunos hongos patógenos postcosecha de la fresa (Fragaria spp.). Revista de la Sociedad Venezolana de Microbiología. 29(1):34-38.
- 13. HARMAN, G.E. 2006. Overview of mechanisms and uses of Trichoderma spp. Phytopathology. 96(2):190-194. https://doi.org/10.1094/phyto-96-0190
- 14. HARMAN, G.E.; PETZOLDT, R.; COMIS, A.; CHEN, J. 2004. Interactions Between Trichoderma harzianum strain T22 and maize inbred line Mo17 and effects of these interactions on diseases caused by Pythium ultimum and Colletotrichum graminicola. Phytopathology. 94(2):147-153. https://doi.org/10.1094/phyto.2004.94.2.147
- 15. HAUSE, B.; MROSK, C.; ISAYENKOV, S.; STRACK, D. 2007. Jasmonates in arbuscular mycorrhizal interactions. Phytochemistry. 68(1):101-110. https://doi.org/10.1016/j.phytochem.2006.09.025
- 16. HAUSE, B.; SCHAARSCHMIDT, S. 2009. The role of jasmonates in mutualistic symbioses between plants soil-born microorganisms. Phytochemistry. and 70(13-14):1589-1599. https://doi.org/10.1016/j.phytochem.2009.07.003
- 17. KAPOOR, R.; SHARMA, D.; BHATNAGAR, A.K. 2008. Arbuscular mycorrhizae in micropropagation systems and their potential applications. Scientia Horticulturae.

116(3):227-239. https://doi.org/10.1016/j.scienta.2008.02.002

- 18. LANTZ, W.; SWARTZ, H.; DEMCHAK, K.; FRICK, S. 2010. Season-long strawberry production with ever bearers for northeastern producers. University of Maryland Extension. 70p.
- 19. LEROUX, P.; GREDT, M.; LEROCH, M.; WALKER, A.-S. 2010. Exploring mechanisms of resistance to respiratory inhibitors in field strains of *Botrytis cinerea*, the causal agent of gray mold. Applied and Environmental Microbiology. 76(19):6615-6630. https://doi.org/10.1128/aem.00931-10
- 20. LI, R.; TAO, R.; LING, N.; CHU, G. 2017. Chemical, organic and bio-fertilizer management practices effect on soil physicochemical property and antagonistic bacteria abundance of a cotton field: Implications for soil biological quality. Soil and Tillage Research. 167:30-38. https://doi.org/10.1016/j.still.2016.11.001
- 21. LI, X.; XIE, X.; XING, F.; XU, L.; ZHANG, J.; WANG, Z. 2019. Glucose oxidase as a control agent against the fungal pathogen Botrytis cinerea in postharvest strawberry. Food Control. 105:277-284. https://doi.org/10.1016/j.foodcont.2019.05.037
- 22. MERCHÁN-GAITÁN, J.B.; FERRUCHO, R.L.; ÁLVAREZ-HERRERA, J.G. 2014. Efecto de dos cepas de Trichoderma en el control de Botrytis cinerea y la calidad del fruto en fresa (Fragaria sp.). Revista Colombiana de Ciencias Hortícolas. 8(1):44-56.
- 23. MINISTERIO DE AGRICULTURA, MINAGRICULTURA. 2021. Cadena de la fresa. Minagricultura. 22p. Disponible desde Internet en: https://sioc.minagricultura.gov.co/Fresa/Documentos/2021-03-31%20Cifras%20Sectoriales.pdf
- 24. MONDAL, T.; DATTA, J.K.; MONDAL, N.K. 2017. Chemical fertilizer in conjunction with biofertilizer and vermicompost induced changes in morpho-physiological and bio-chemical traits of mustard crop. Journal of the Saudi Society of Agricultural Sciences. 16(2):135-144. https://doi.org/10.1016/j.jssas.2015.05.001
- 25. PERTOT, I.; GIOVANNINI, O.; BENANCHI, M.; CAFFI, T.; ROSSI, V.; MUGNAI, L. 2017. Combining biocontrol agents with different mechanisms of action in a strategy to control Botrytis cinerea on grapevine. Crop Protection. 97:85-93. https://doi.org/10.1016/j.cropro.2017.01.010
- 26. PERTOT, I.; ZASSO, R.; AMSALEM, L.; BALDESSARI, M.; ANGELI, G.; ELAD, Y. 2008. Integrating biocontrol agents

in strawberry powdery mildew control strategies in high tunnel growing systems. Crop Protection. 27(3-5):622-631. https://doi.org/10.1016/j.cropro.2007.09.004

- POZO, M.J.; AZCÓN-AGUILAR, C. 2007. Unraveling mycorrhiza-induced resistance. Current Opinion in Plant Biology. 10(4):393-398. https://doi.org/10.1016/j.pbi.2007.05.004
- 28. PRITTS, M. 2002. Growing strawberries, healthy communities, strong economies and clean environments: what is the role of the researcher? Acta Horticulturae. 567:411-417. https://doi.org/10.17660/ActaHortic.2002.567.85
- 29. PROGRAMA DE TRANSFORMACIÓN PRODUCTIVA, PTP.; ASOCIACIÓN HORTIFRUTICOLA DE COLOMBIA, ASOHOFRUCOL.; FONDO NACIONAL DE FOMENTO HORTIFRUTÍCOLA. 2013. Plan de negocios de fresa: Programa de transformación productiva. 171p.
- 30.SELOSSE, M.-A.; BAUDOIN, E.; VANDENKOORNHUYSE, P. 2004. Symbiotic microorganisms, a key for ecological success and protection of plants. Comptes Rendus Biologies. 327(7):639-648. https://doi.org/10.1016/j.crvi.2003.12.008
- STOKES, M.E.; DAVIS, C.S.; KOCH, G.G. 2012. Categorical Data Analysis Using SAS. Third Edition. SAS Institute Inc (Cary, NC). 590p.
- VÁZQUEZ, M.M.; CÉSAR, S.; AZCÓN, R.; BAREA, J.M. 2000. Interactions between arbuscular mycorrhizal fungi and other microbial inoculants (*Azospirillum, Pseudomonas,*

Trichoderma) and their effects on microbial population and enzyme activities in the rhizosphere of maize plants. Applied Soil Ecology. 15(3):261-272. https://doi.org/10.1016/S0929-1393(00)00075-5

33. VESTBERG, M.; KUKKONEN, S.; SAARI, K.; PARIKKA, P.; HUTTUNEN, J.; TAINIO, L.; DEVOS, N.; WEEKERS, F.; KEVERS, C.; THONART, P.; LEMOINE, M.-C.; CORDIER, C.; ALABOUVETTE, C.; GIANINAZZI, S. 2004. Microbial inoculation for improving the growth and health of micropropagated strawberry. Applied Soil Ecology. 27(3):243-258. https://doi.org/10.1016/j.apsoil.2004.05.006

34. WEHNER, J.; ANTUNES, P.M.; POWELL, J.R.;

- 4. WEFINER, J.; ANTUNES, P.M.; POWELL, J.K.; MAZUKATOW, J.; RILLIG, M.C. 2010. Plant pathogen protection by arbuscular mycorrhizas: A role for fungal diversity? Pedobiologia. 53(3):197-201. https://doi.org/10.1016/j.pedobi.2009.10.002
- YANG, H.-H.; YANG, S.L.; PENG, K.-C.; LO, C.-T.; LIU, S.-Y. 2009. Induced proteome of *Trichoderma harzianum* by *Botrytis cinerea*. Mycological Research. 113(9):924-932. https://doi.org/10.1016/j.mycres.2009.04.004
- 36. ZHANG, H.; WANG, L.; DONG, Y.; JIANG, S.; CAO, J.; MENG, R. 2007. Postharvest biological control of gray mold decay of strawberry with *Rhodotorula glutinis*. Biological Control. 40(2):287-292. https://doi.org/10.1016/j.biocontrol.2006.10.008