

# CONJUGATED LINOLEIC ACID IN MILK AND FERMENTED MILKS: VARIATION AND EFFECTS OF THE TECHNOLOGICAL PROCESSES

## ÁCIDO LINOLEICO CONJUGADO EN LECHE Y LECHE FERMENTADAS: VARIABILIDAD Y EFECTOS DE LOS PROCESOS TECNOLÓGICOS

Luis Felipe GUTIÉRREZ, MSc, PhD<sup>1\*</sup>

Received: January 19, 2015 Approved: August 08, 2016

### ABSTRACT

**Background:** Conjugated linoleic acid (CLA) is a generic term used to describe a group of geometric and positional isomers of linoleic acid with a conjugated double bond system. CLA-isomers have been widely studied because of their important biological activity and their protective effects against several diseases, such as obesity, atherosclerosis, chronic inflammatory diseases, and cancer. Consequently, these biomolecules have attracted much attention from the dairy industry, since they are naturally found in ruminants' milk, and because the development of CLA-enriched dairy foods can be a good economic opportunity given the growth of the functional foods market, in which the dairy industry plays an important role. **Objectives:** This work presents a comprehensive review of the following aspects: (i) The synthesis and concentration of the CLA-isomers in milk, and the main strategies employed to increase their content in a natural manner; (ii) The influence of the main technological treatments applied to milk on the concentration of CLA-isomers; and (iii) The effects of milk fermentation on the content of CLA-isomers, and the challenges of this technological process, which has been thought as a promissory alternative to naturally increase the content of CLA in fermented dairy products. **Methods:** Information available in various databases was reviewed. A total of 103 articles were selected on the basis of their relevance and scientific-technical quality. **Results:** The CLA concentration in cows' milk normally ranges between 2 and 37 mg/g fat, and is mainly affected by the dietary regime offered to the animals. From the total CLA-isomers, rumenic acid represents between 75 and 90%. The technological processes normally applied to milk (thermal processing, high pressure processing, and fermentation) might cause slight changes on the CLA concentration, but the mechanisms causing these changes have not been still established. The increase in CLA concentration by milk fermentation is strain-dependent, because of the different linoleate isomerase activity of the species. **Conclusions:** Although several studies have reported increases in the concentration of CLA in milk and fermented milks in a natural manner, they are fairly moderate, and the obtained levels of CLA are significantly lower than those recommended to achieve therapeutic effects.

**Keywords:** Conjugated linoleic acid, functional foods, milk, probiotics, yogurt.

<sup>1</sup> Grupo de investigación en biomoléculas alimentarias. Instituto de Ciencia y Tecnología de Alimentos (ICTA), Universidad Nacional de Colombia, Cra. 30 No. 45-03, Edificio 500A, Bogotá, Colombia

\* Autor de correspondencia: [lfgutierrez@unal.edu.co](mailto:lfgutierrez@unal.edu.co)

## RESUMEN

**Antecedentes:** El ácido linoleico conjugado (CLA) es un término genérico utilizado para describir el grupo de isómeros geométricos y posicionales del ácido linoleico con un sistema de dobles enlaces conjugados. Los isómeros de CLA han sido ampliamente estudiados debido a su importante actividad biológica y a sus efectos protectores contra varias enfermedades, como la obesidad, arteriosclerosis, enfermedades inflamatorias crónicas y el cáncer. En consecuencia, estas biomoléculas han despertado mucho interés en la industria láctea, dado que están presentes naturalmente en la leche de rumiantes, y porque el desarrollo de productos lácteos enriquecidos con CLA puede representar una oportunidad económica interesante, dado el crecimiento del mercado de alimentos funcionales, en el cual la industria láctea tiene una participación importante. **Objetivos:** En este trabajo se presenta una revisión detallada de los siguientes aspectos: (i) El origen de los isómeros de CLA en la leche, sus concentraciones promedio y las principales estrategias utilizadas para incrementar su contenido de manera natural; (ii) La influencia de los principales tratamientos tecnológicos aplicados a la leche en la concentración de los isómeros del CLA; y (iii) Los efectos de la fermentación de la leche en el contenido de isómeros de CLA, y los retos de este proceso tecnológico concebido como promisorio para aumentar naturalmente el contenido de CLA en leches y productos lácteos fermentados. **Métodos:** Se revisó la información disponible en varias bases de datos. Un total de 103 artículos fueron seleccionados con base en su pertinencia y calidad técnico-científica. **Resultados:** La concentración de CLA en la leche de vaca normalmente oscila entre 2 y 37 mg/g grasa, y es afectada principalmente por la dieta suministrada a los animales. Del total de isómeros de CLA, el ácido ruménico representa entre 75 y 90%. Los procesos tecnológicos aplicados normalmente a la leche (procesado térmico, procesamiento por altas presiones y fermentación) pueden causar ligeras modificaciones en la concentración de CLA, pero los mecanismos que causan estos cambios no han sido aún establecidos. El incremento en la concentración de CLA por fermentación de la leche es dependiente de la cepa, debido a diferencias en la actividad de la linoleato isomerasa de las especies. **Conclusiones:** Aunque varios estudios han reportado incrementos en la concentración de CLA en leche y leches fermentadas de manera natural, estos son bastante moderados, y los niveles de CLA obtenidos son significativamente menores que los recomendados para alcanzar los efectos terapéuticos.

**Palabras clave:** Ácido linoleico conjugado, alimentos funcionales, leche, probióticos, yogur.

## INTRODUCTION

Milk and dairy products are daily consumption foodstuffs, considered as important sources of energy, and of a variety of bioactive substances positively associated with human health, such as proteins and peptides, oligosaccharides, lipids, minerals and vitamins (1). Milk fat is mainly composed of triacylglycerols (~98%), minor amounts of mono and diacylglycerols (~2%), phospholipids (~1%), sterols (~0.5%), free fatty acids (0.1%) and traces of fat-soluble vitamins (2). The high concentration of saturated fatty acids (mainly that of palmitic, myristic and lauric acids) in the milk's lipid fraction has generated some concern, because of their negative effects on human health, especially related to the increased risk of acquiring cardiovascular diseases (3). However, milk's lipid fraction also contains mono and polyunsaturated fatty acids, such as oleic acid (C18:1 *cis*9), and fatty acids belonging to the omega-3 (C18:3) and omega-6 families (C18:2),

which are considered beneficial to human health, especially for reducing the level of triglycerides and LDL cholesterol, and for enhancing the levels of HDL cholesterol (4, 5). Since the concentration of these fatty acids is relatively low for providing health benefits, significant efforts have been devoted to increasing their content in milk and dairy products, in order to produce foodstuffs with improved nutritional value. The main employed strategies are based on the manipulation of the feed and dietary regimen of the animals, and on the modification of the technological processes used in the manufacture of dairy products (6-8).

Conjugated linoleic acid is a generic term used for describing the geometrical and positional isomers of linoleic acid (C18:2 *cis*9, *cis*12) with a conjugated double bond system (9). Milk fat is the richest natural source of CLA, with concentrations typically ranging between 2 and 37 mg/g fat (10). The Table 1 presents the predominant distribution of the CLA-isomers in milk and dairy products.

**Table 1.** Distribution of the main CLA-isomers in milk and dairy products (Adapted from Bauman and Lock (11)).

Isomer	Percentage of the total isomers
C18:2 <i>cis</i> 9, <i>trans</i> 11	72.6-91.2
C18:2 <i>trans</i> 7, <i>cis</i> 9	1.2-8.9
C18:2 <i>trans</i> 9, <i>trans</i> 11	0.8-2.0
C18:2 <i>trans</i> 11, <i>trans</i> 13	0.3-4.2
C18:2 <i>trans</i> 12, <i>trans</i> 14	0.3-2.8
C18:2 <i>trans</i> 10, <i>trans</i> 12	0.3-1.3
C18:2 <i>cis</i> 11, <i>trans</i> 13	0.2-4.7
C18:2 <i>trans</i> 8, <i>trans</i> 10	0.2-0.4
C18:2 <i>trans</i> 11, <i>cis</i> 13	0.1-8.0
C18:2 <i>trans</i> 9, <i>trans</i> 9	<0.1-2.4
C18:2 <i>trans</i> 8, <i>cis</i> 10	<0.1-1.5
C18:2 <i>trans</i> 10, <i>cis</i> 12	<0.1-1.5
C18:2 <i>cis</i> 12, <i>trans</i> 14	<0.01-0.8
Other <i>cis-cis</i>	0.1-4.8

As it can be appreciated in Table 1, the *cis*-9, *trans*-11-octadecadienoic acid (C18:2 *cis*9, *trans*11), also known as rumenic acid, is the predominant isomer, representing between 75 and 90% of the total CLA-isomers (12). The second most abundant isomer is the C18:2 *trans*7, *cis*9, represents about 10% of the total CLA-isomers. The remaining isomers, including the *trans*-10, *cis*-12-octadecadienoic acid (C18:2 *trans*10, *cis*12), are present in small concentrations, mostly around 0.5% (11).

Among the identified CLA-isomers, the C18:2 *cis*9, *trans*11 and the C18:2 *trans*10, *cis*12, are considered as molecules biologically active, due to their protective effects against various common diseases such as obesity (13, 14), atherosclerosis (15, 16), diabetes (17, 18), some chronic inflammatory diseases (1) and cancer (4, 19, 20). Anticarcinogenic effects have been observed in different cancer types, with doses varying between 55 mg and 3.5 g CLA/day (21-23), but the most important results have been reported regarding breast cancer (11).

Numerous studies have evaluated the factors affecting the content of CLA in milk and dairy products, because these foodstuffs are the main source of CLA in the human diet, providing about 70% of the total CLA daily intake, which varies between 70 and 430 mg/day (11, 24). However, including the conversion of vaccenic acid in the human body by the  $D^9$ -desaturase enzyme, an average intake of 650 mg/day can be achieved (24). Since these intake values are lower than those recommended for achieving beneficial effects on human health (3-4 g CLA/day), various technological alternatives have been investigated in order to increase the CLA

concentration in milk and dairy products, and hence the daily CLA intake of the population. The main investigated alternatives comprise the manipulation of the feed and dietary regimen of the animals, and the milk fermentation with CLA-producing bacteria (bacteria possessing linoleate isomerase activity), alone or in combination with specific starter cultures (24).

Taking into account that the development of dairy products with high concentrations of CLA might represent an interesting economic opportunity, due to important participation of the dairy industry in the functional foods market, the objective of this work is to present a comprehensive review of: **(i)** The content of CLA in milk and the main factors affecting its concentration; **(ii)** the influence of the technological treatments normally applied to milk on the concentration of CLA; and **(iii)** the effects of the milk fermentation on the CLA concentration of fermented dairy products, and the main challenges of this technological process, considered as promissory for obtaining dairy products naturally enriched with CLA. To the best of our knowledge, articles combining these topics have not been yet published.

## METHODS

The data presented in this review were collected from scientific publications and thesis published between 2000 and 2014. The literature search was achieved using the following keywords: conjugated linoleic acid, milk, fermented milks and yogurt. From a total of about 300 documents found, 103 were selected and analyzed on the basis of their pertinence, relevance and technical-scientific quality. Taking into account that the concentration of CLA was not always expressed in the same units in the published papers, in some cases conversion factors were used, for making the comparisons between the data of the selected references.

## CLA IN MILK: SYNTHESIS, VARIABILITY AND EFFECTS OF THE TECHNOLOGICAL PROCESSES

### CLA synthesis

The formation of CLA-isomers in milk is carried out by means of isomerization and biohydrogenation reactions of the unsaturated fatty acids presented in the feed and dietary regimen of the animals. These

reactions are produced by the rumen bacteria and the enzymatic activity of the  $\Delta^9$ -desaturase in the mammary gland (10). The linoleic acid ingested in the diet is firstly isomerized to rumenic acid (C18:2 *cis*9, *trans*11) by the *cis*-12, *trans*-11 isomerase enzyme, and then biohydrogenated to vaccenic acid (C18:1 *trans*11) in the rumen by the bacteria *Butyrivibrio fibrisolvens* (10). This anaerobic bacteria is also responsible of the formation of the rumenic and vaccenic acids from the  $\alpha$ -linolenic (C18:3 *cis*9, *cis*12, *cis*15) and  $\gamma$ -linolenic (C18:3 *cis*6, *cis*9, *cis*12) acids (25). If the initial isomerization of the C18:2 *cis*9, *cis*12 acid involves the *cis*12 double bond, the isomer C18:2 *cis*9, *trans*11 is produced, whereas if the initial double bond isomerized is the *cis*9, the isomer C18:2 *trans*10, *cis*12 is mainly produced (11).

In the mammary gland, the biochemical reaction of the  $\Delta^9$ -desaturase enzyme with the vaccenic acid leads to the production of 70-90% of the C18:2 *cis*9, *trans*11 isomer (26). The other CLA-isomers, including the C18:2 *trans*10, *cis*12, are intermediate products of the biohydrogenation reactions in the rumen. Thus, their concentration in milk are lower than those of the rumenic acid (27). Detailed studies on the lipid metabolism in the rumen and its effects on the CLA-isomers production have been recently published (10, 28).

### Variability

The concentration of CLA in cows' milk normally ranges between 2 and 37 mg/g fat (10). Recent studies have reported CLA concentrations of about 13.5 mg/g fat, in milks from "Sabana de Bogotá" (29). However, pasteurized milks with high content of CLA (30 mg/g fat) can be found in markets of various countries such as Canada and Spain (30).

The lactation stage, the rumen microflora and the type of feed offered to the animals are some of factors influencing the concentration of CLA in milk (31, 32). However, taking into account that the CLA-isomers are mainly synthesized from the unsaturated fatty acids ingested by the animals, the diet is the factor affecting the most the content of CLA in milk fat (10). According to Bell and Kennelly (33), the concentration of CLA in milk can be increased up to 10 fold, by the manipulation of the animal's diet. Various studies have demonstrated that the CLA content in milk increases significantly by using feed systems based on the consumption of fresh pasture, whose content of polyunsaturated fatty acids is high (32, 34). On the contrary, diets based on the supply of grains and preserved forage, lead to a reduction in the CLA

concentration in milk, due to the pH decreases and changes in the microbial composition of the rumen (35). The fodder type and the regrowth age would also have an effect on the concentration of CLA isomers in milk (35). Other researches have shown that the addition of supplements rich in linoleic acid, such as soybean, olive, canola, sunflower and fish oils, can slightly increase both the CLA content and the concentration of unsaturated fatty acids in milk and dairy products (3, 9, 36). Nevertheless, is important to note that the use non-protected oils, such as vegetable and fish oils, generally produces milk fat depression and significant decreases in the milk yield, as well as increases in some *trans* fatty acids, such as C18:1 *trans*11, C18:1 *trans*10 y C18:2 *trans*9, *cis*11 (37). The complete details of the main diet effects on the concentration of CLA isomers in milk have been the subject of recent review articles (10, 38-40).

### Effects of the technological processes

The studies of the influence of the technological processes normally applied to milk for manufacturing dairy products on the CLA concentration are very diverse. The reported results are controversial due to their remarkable differences, suggesting that the CLA concentration in dairy products mainly depends on the CLA content in the raw milk, which, as mentioned above, is principally influenced by the diet regimen of the animals, and in a less manner by the reactions produced during technological processes used in milk processing.

#### Thermal processing

Milk is thermally processed in order to eliminate pathogens and make it safe for human consumption. The effects of the thermal treatments on the main milk components (lactose, proteins, fat and vitamins) have been extensively studied (41). However, specific studies related to the effect of the thermal processing of milk on the CLA concentration are scarce and their results present much variation.

According to some authors, the heating of milk fat in the presence of proteins in aqueous solution can generate an increase in the concentration of CLA (42). Other studies indicate that the microwave heating of milk and the UHT treatment (140°C, 4 s), can produce significant decreases in the CLA content, due to oxidation reactions of the fat and the generation of hydroperoxides that could cause conversion or degradation of the CLA isomers (43, 44). Similar results were found after heating milks by different

pasteurization treatments (HTST: 77.2°C, 16 s; LTLT: 60°C, 20 min; and ultra-pasteurization: 138°C, 3 s) (45-47). On the contrary, it has been also reported that various pasteurization treatments (85°C, 16 s; 95°C, 5 min; 63°C, 30 min; 70-90°C, 5 min) and UHT processing do not generate significant changes on the CLA content of milk (9, 43, 48). Recent investigations evaluated the influence of various thermal treatments (pasteurization (72°C, 30 s); HTST (85°C, 30 s); UHT (135°C, 30 s); UHT (150°C, 5 min); sterilization (121°C, 15 min) and microwave heating (650 W, 1.30 min)) on the milk' CLA concentration (49). The HTST pasteurization produced a sigmatropic rearrangement of the CLA-isomers, increasing the concentration of the C18: 2 *cis*9, *trans*11 isomer. The sterilization treatment generated the isomerization of the linoleic acid into the C18:2 *trans*9, *trans*11 isomer, while the content of the C18:2 *cis*9, *trans*11 isomer in the milk heated by microwaves was significantly higher than that of the untreated milk. In addition, the concentration of CLA in milks treated by means of the UHT process gradually decreased during the storage period (50). These results were attributed to the presence of oxygen during the HTST pasteurization and the microwave treatment, allowing the formation of protein radicals, which can react with the linoleic acid, thus increasing the concentration of the C18:2 *cis*9, *trans*11 isomer (49). These results are in agreement with previous findings (51), which indicate that the thermal processing of milk may alter the CLA-isomers' distribution, but their total concentration remains constant.

The retention kinetics of CLA and *trans*-vaccenic acid of milks thermally treated within the temperature interval between 90 and 120°C, was recently investigated (23). Both lipids were oxidized in great extent when the temperature increased from 90 to 120°C. After 60 minutes of heating at 90°C, the retention percentage of CLA varied between 68 and 71%, while in the milks treated at 120°C during 15 minutes, the retention percentage was only 15-21%.

Although there is no a rule concerning the effects of thermal processing on the CLA concentration in milk, the normal thermal processes applied to milk could affect not only the concentration of CLA in milk, but also the CLA-isomer distribution. More research is needed in order to establish the mechanisms of these possible changes.

#### *High pressure processing*

The potential of emerging technologies such as high hydrostatic pressure has been widely studied

as non-thermal alternative to milk pasteurization. However, few studies have evaluated the influence of this technology on the concentration of CLA in milk and dairy products. The fatty acid composition (including that of CLA-isomers) of milks homogenized at high pressures (up to 350 MPa), did not present significant variations in comparison to the control samples (52). Milks naturally enriched with CLA, did not show significant changes in the fatty acid composition, including the CLA-isomers, after processing at high pressures (400 MPa, 15 min, 25°C) (49). Recent reports indicate that the combined effect of high pressure with temperature may produce an increase in the retention of CLA (30, 53). The retention kinetics of the CLA-isomers of milks treated by high pressure sterilization (100-600 MPa, 90-120°C) was well described by the Weibull model (6). The retention of CLA decreased with an increase of temperature, but more CLA was retained with an increase of pressure. In addition, the processing conditions at which commercial sterilization may be achieved (120°C and 600 MPa with 3 min of holding time) allowed to retain more than 80% of the CLA-isomers. This high retention value was attributed to isomerization reactions, instead oxidation reactions, taking into account that in the presence of oxygen the CLA isomerization is thermally induced through a free radical mechanism in which oxygen is consumed during the reaction (51). However, the retention of CLA decreased progressively during storage, being approximately 36% after 60 days at 25°C (30).

#### *Refrigeration*

Some studies have evaluated the influence of the refrigeration process of milk and dairy products on the content of CLA-isomers. In general, the results indicate that refrigerated storage does not significantly affect the concentration of CLA in milk and dairy products (54, 55). However, there are reports that indicate important decreases of the C18:2 *cis*9, *trans*11 and C18:2 *cis*10, *trans*12 isomers of skim milk enriched with CLA after three weeks of refrigerated storage, possibly due to the activity of microbial lipases (45). Leite, Lima and Baptista (46) found a 12% reduction of the C18:2 *cis*9, *trans*11 isomer in commercial UHT milk after two months of refrigerated storage at 6-7°C. Similarly, significant decreases of the C18:2 *cis*9, *trans*11; C18:2 *cis*11, *trans*13; C18:2 *cis*10, *cis*12 and C18:2 *cis*10, *trans*12 isomers were found during refrigerated storage of fresh cheese, as a consequence of an excessive microbial growth (54).

### Fermentation

The ability of microorganisms for producing CLA from linoleic acid depends on the activity of their linoleate isomerase, enzyme catalyzing this reaction (24). Species expressing linoleate isomerase activity can be divided in two groups: bacteria that produce mainly C18:2 *cis*9, *trans*11 from linoleic acid, and bacteria able to produce mainly C18:2 *trans*10, *cis*12 from linoleic acid. In addition to the rumen bacteria, this enzyme has been found in strains of *Bifidobacterium*, *Enterococcus*, *Lactobacillus*, *Lactococcus*, *Propionibacterium* y *Streptococcus*, which have been classified as potential CLA-producers (56-58). Genetic sequences of the linoleate isomerase enzymes from strains of *B. dentium*, *B. breve*, *Lc. lactis* ssp. *lactis*, *L. acidophilus*, *L. plantarum*, *L. reuteri*, *P. acnes*, and *Rhodococcus erythropolis* are available in GenBank (24).

The influence of the processing conditions for producing of CLA *in vitro* using microorganisms has been largely studied (24, 58-60). The concentration of linoleic acid, pH, temperature and the microbial growth stage are the most important factors on the bioproduction yield of CLA (24). Although most of the investigated microorganisms are normally used as starter cultures or as probiotics in the manufacture processes of fermented milks, is important to note that the obtained results of *in vitro* essays in MRS medium, may not be extrapolated to the milk fermentation processes, because of the changes and multiple interactions characterizing the milk-based substrates. Some recent findings are discussed below.

Studies have shown that strains of *B. breve* could be used as starter cultures for the development of functional milk products with high concentrations of bioactive lipids such as CLA (61-66). Conversions of linoleic acid into CLA up to 74% were obtained using three strains of *B. breve* (ZL12-28, 29M2 and M7-70) isolated from breast milk (62). These strains cultured in MRS medium and reconstituted skimmed milk, produced different CLA-isomers (C18:2 *cis* 9, *trans* 11; C18:2 *trans* 10, *cis* 12 y C18:2 *trans* 9, *trans* 11) in amounts varying between 160-170 mg CLA/mL. Coakley et al. (64) obtained CLA (mainly the C18:2 *cis*9, *trans*11 isomer) by using strains of *B. breve* and *B. dentium*. Among 36 strains of *Bifidobacterium*, *B. breve*, *B. bifidum*, and *B. pseudolongum* ssp. *pseudolongum* converted 20 to 54% of linoleic acid into CLA (C18:2 *cis*9, *trans*11 y C18:2 *trans*9, *trans*11) in MRS medium (63). Similarly, strains of *B. breve* were able to produce CLA (mainly the C18:2 *cis*9, *trans*11 isomer) in skimmed

milk supplemented with linoleic acid (0.5 mg/mL), with conversions up to 23% (61). The conversion was higher (39%) when strains of *B. bifidum* were cultured in buffalo milk supplemented with linoleic acid at low concentrations (0.2 mg/mL) (67). *B. breve* LMG 13194 and *B. pseudolongum* ssp. *pseudolongum* LMG 11595 produced higher conversion of linoleic acid into CLA in MRS medium, in comparison to *B. breve* LMG 11084, *B. breve* LMG 11040, and *B. breve* LMG 11613, confirming that the CLA production is strain-dependent, probably because of the differences in the genetic sequences of the linoleate isomerase enzyme of each microorganism (68).

Numerous strains of *Lactobacillus* have shown high linoleate isomerase activity and can biosynthesize CLA from linoleic acid (69-76). *L. plantarum* ZS2058 converted more than 50% of linoleic acid into CLA (C18:2 *cis*9, *trans*11 y C18:2 *trans*9, *trans*11) in MRS medium, while strains of *L. bulgaricus*, *L. crispatus*, *L. gasseri* and *L. helveticus* converted only 10-20% of linoleic acid in three CLA-isomers (C18:2 *cis*9, *trans*11; C18:2 *trans*9, *trans*11 and C18:2 *trans*10, *cis*12) (69). Recently, high conversions (86.4%) of linoleic acid into CLA using permeabilized *L. acidophilus* cells were reported (77). The whole cells converted only 38.5%. The permeabilized cells were recycled ten times without showing a significant decrease in their catalytic activity. Similar results were obtained by Lee et al. (78), who reported 35% conversion of linoleic acid into CLA by using immobilized *L. reuteri* cells. The cells were recycled five times without presenting negative effects on the production of CLA. Lin et al. (70) evaluated the CLA production capacity of strains of *L. acidophilus*, *L. delbrueckii* ssp. *bulgaricus* and *L. delbrueckii* ssp. *lactis*, in MRS medium enriched with 12% skimmed milk and linoleic acid in concentrations ranging between 0 and 5000  $\mu$ g/mL. An increase in the concentration of CLA was observed in the substrates added with linoleic acid. *L. acidophilus* presented the highest CLA production capacity (23.0-105.5  $\mu$ g/mL), when incubated during 24 hours at 37°C, with a linoleic acid addition of 1000  $\mu$ g/mL. Further increases in the concentration of linoleic acid (from 1000 to 5000  $\mu$ g/mL), as well as in the incubation time (from 24 to 48 hours), did not conduce to significant increases in CLA production. Similarly, strains of *L. acidophilus*, *L. plantarum*, *L. casei*, *L. delbrueckii* ssp. *bulgaricus*, and *Streptococcus thermophilus* produced CLA in MRS medium enriched with linoleic acid at 0.50 mg/mL (75).

The CLA production capacity of more than 250 strains from the genera of *Lactobacillus*, *Streptococcus*, *Pediococcus*, *Leuconostoc*, *Propionibacterium*, *Bifidobacterium*, *Weissella*, *Aquaspirillum*, *Enterococcus*, *Tetragenococcus*, *Aerococcus*, *Butyrivibrio* and *Lactococcus* was studied by Kishino et al. (79). Strains from the genera of *Enterococcus*, *Pediococcus*, *Propionibacterium*, and *Lactobacillus* produced considerable amounts of CLA in MRS medium supplemented with linoleic acid. Two strains of *P. freudenreichii* ssp. *freudenreichii* and one strains of *P. freudenreichii* ssp. *shermanii* converted 60 to 90% of linoleic acid into CLA (C18:2 *cis*9, *trans*11) in MRS medium (80), while strains of *Lc. lactis* ssp. *lactis* biovar *diacetylactis* and *Leuconostoc mesenteroides* ssp. *mesenteroides* were able to produce CLA (0.20 mg/mL) in skimmed milk added with hydrolyzed sesame oil (81).

## CLA IN FERMENTED MILKS

Fermented milks such as yogurt are considered healthy foods, due to their beneficial effects on human health. The concentration of CLA in fer-

mented milks normally varies between 3.4 and 8.8 mg/g fat (24, 82). Recent reports indicate that the content of CLA in commercial yogurt and kumis from Colombia ranges between 4.5 and 8.2 and between 7.6 y 22.6 mg/g fat, respectively (82, 83). Table 2 presents the concentration of CLA in various fermented milks, as well as the starter cultures employed in the fermentation process.

Various studies have evaluated the influence of the milk fermentation on the concentration of CLA. The reported results are controversial. While some authors report that fermentation of milk does not affect the CLA content (84-86), others suggest that the use of lactic acid bacteria may increase (8, 55, 57) or even reduce (47) the concentration of CLA in fermented milk products. More research is needed in order to elucidate if the fermentation of milk can be used as an alternative method for manufacturing dairy products with high CLA content. However, from the published results it can be summarized that:

**Table 2.** Concentration of CLA (mg/g fat) in various fermented milks

Product	Starter cultures	CLA	Ref.
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i>	4.7-7.6	(87)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i>	2.4-4.5	(87)
Fermented milk	<i>Lc. lactis</i> ssp. <i>lactis</i>	5.1-5.5	(8)
Fermented milk	<i>L. plantarum</i>	4.9	(8)
Fermented milk	<i>L. buchneri</i>	4.3	(8)
Fermented milk	<i>L. reuteri</i>	4.8	(8)
Fermented milk	<i>L. brevis</i>	5.0	(8)
Fermented milk	<i>L. casei</i>	6.3	(8)
Fermented milk	<i>L. helveticus</i>	5.4	(8)
Fermented milk	<i>Lactobacillus</i> sp.	6.3	(8)
Fermented milk	<i>Lactococcus</i> sp.	5.6-8.7	(8)
Fermented milk	<i>Lc. lactis</i>	11.2	(8)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i>	16.5	(88)
Fermented milk	<i>B. animalis</i> ssp. <i>lactis</i>	15.5-18.5	(88)
Fermented milk	<i>B. bifidum</i>	2.7-4.2	(89)
Fermented milk	<i>B. breve</i>	2.3-4.4	(89)
Fermented milk	<i>B. pseudolongum</i> ssp. <i>pseudolongum</i>	2.7-4.2	(89)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>B. bifidum</i>	2.7-4.2	(89)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>B. breve</i>	2.3-4.4	(89)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>B. pseudolongum</i> ssp. <i>pseudolongum</i>	3.0-4.4	(89)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>L. acidophilus</i> L10	4.7-11.0	(90)
	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>B. animalis</i> ssp. <i>lactis</i> (BI04, HN019, B94)	4.7-9.6	(90)
Fermented milk	<i>L. viridescens</i>	1.8-5.7	(76)
Fermented milk	<i>L. lactis</i>	1.9-9.2	(76)
Fermented milk	<i>L. brevis</i>	1.7-10.5	(76)
Dahi	<i>L. acidophilus</i> + <i>L. casei</i>	10.5	(91)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i>	2.1-2.5	(85)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>L. acidophilus</i>	3.3-5.6	(85)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>B. animalis</i>	3.6-6.0	(85)
Yogurt	<i>S. thermophilus</i> + <i>L. delbrueckii</i> ssp. <i>bulgaricus</i> <i>L. acidophilus</i>	8.8-20.8	(86)

(i) The nature of milk can influence either the amount or the type of CLA-isomers in fermented milks. Studies have shown that the CLA content of yogurts from sheep milk is higher than that of yogurts from cow's milk (4.7-7.6 vs. 2.4-4.5 mg/g fat) (87), while the CLA concentrations of Greek yogurts elaborated with milks from cow, sheep and goat varied between 12.8-15.1, 4.1-12.5 and 4.3-9.8 mg CLA/g fat, respectively (92). After 14 days of refrigerated storage at 5°C, the concentration of CLA significantly increased in yogurts from goat milk, but it was significantly reduced in those from cow's milk (87). During the manufacturing of Dahi (fermented product from buffalo milk, similar to yogurt) Yadav et al. (91) found a significant increase (about twice) in the amount of CLA, when using *L. acidophilus* and *L. casei* as starter cultures. The refrigerated storage at 4°C did not affect the concentration of the CLA-isomers. On the other hand, the CLA content of organic fermented milks, produced using strains of *B. animalis* ssp. *lactis* in combination with *S. thermophilus*, was slightly higher than the CLA concentration of the starting milk (88, 93, 94). In contrast, there was no formation of CLA in fermented milks with strains of *Bifidobacterium*, despite having been chosen for their ability to produce CLA *in vitro*, and being cultured in substrates rich in linoleic acid (89).

(ii) The modification of the substrate can also affect the concentration of CLA in fermented milks. For example, the production of CLA was increased in milks fermented with *Lc. lactis* I-01 from substrates added with sunflower oil (0.1 mg/mL) (95). Also, the fermentation of milk enriched with linoleic acid (0.1%) with lactic cultures and *L. acidophilus*, showed a significant increase in the CLA content, without negative effects on the sensory properties (96). High values of CLA (5.58 and 7.06 mg/g fat) were obtained in low fat yogurts, in comparison to the reported in other studies (1.7 mg/g fat (97) and 5.25 mg/g fat, from milk added with 4.40 mg CLA/g fat (55)). Slightly elevated concentrations of the C18:2 *cis*9, *trans*11 and C18:2 *trans*10, *cis*12 isomers were obtained in fermented milks elaborated from substrates added with hydrolyzed soybean oil, which were cultured with *P. freudenreichii* ssp. *freudenreichii* 23, *P. freudenreichii* ssp. *shermanii* 56, *P. freudenreichii* ssp. *shermanii* 51, *L. rhamnosus* and yogurt starter cultures (*L. delbrueckii* ssp. *bulgaricus* and *S. thermophilus*) (98). The combination of probiotics and yogurt starter cultures led to higher concentrations of CLA than

those obtained without probiotic cultures. It has been also found that the addition of monolinolein favors the production of CLA in milks fermented with *B. breve* LMC 017 (66). Moreover, the fatty acid profile of yogurts fermented with *S. thermophilus* and *L. delbrueckii* ssp. *bulgaricus* and strains of *B. lactis* Bl04 or B94 was enhanced by the addition of açai pulp to the starting milk, which provided an extra amount of unsaturated fatty acids, which may act as CLA precursors (99). On the contrary, the addition of oils rich in unsaturated fatty acids did not modify the CLA concentration of creams fermented with various probiotic cultures (100). Similar results were obtained by Xu et al. (101), who did not observe significant increases in the concentration of CLA when milk added with 1% fat was cultured with *L. rhamnosus* and yogurt starter cultures (*L. bulgaricus* and *S. thermophilus*). The content of the C18:2 *cis*9, *trans*11 isomer remained constant during the refrigerated storage, while the concentration of the C18:2 *trans*10, *cis*12 isomer increased up to 3.3 fold after two weeks of storage. The addition of dietary fibers from apple, banana and passion fruit processing by-products led to a significant increase of the polyunsaturated and short chain fatty acids of yogurts fermented with four different strains of probiotic cultures: *L. acidophilus* L10 and *B. animalis* ssp. *lactis* BL04, HN019 and B94 (90). A synergistic effect between the type of fiber and the probiotic strain on the CLA concentration was found. In yogurts without fiber addition, the content of the C18:2 *cis*9, *trans*11 isomer varied between 4.7 and 8.0 mg/g fat, while in the fiber-added yogurts, the values were slightly higher, varying between 5.4 and 11.2 mg/g fat. The control yogurt samples, co-fermented with *B. animalis* ssp. *Lactis*, showed CLA concentrations higher than those co-fermented with *L. acidophilus* L10 (8.0 vs. 4.7 mg/g fat), suggesting again that the CLA products are strain-dependent. In spite of this, the highest increases in the CLA concentration were obtained in the samples added with fibers from banana and passion fruit, co-fermented with *L. acidophilus* L10. The fiber from passion fruit promoted the increase in the CLA content of all yogurt samples. These results are in agreement with previous studies in which the combination of strains of *L. acidophilus* and *B. animalis*, and prebiotics such as fructooligosaccharides (FOS), favored the production of CLA during the manufacture of fermented milks (85, 102). Milks added with FOS (2%) and co-fermented with *L. acidophilus* and *B. animalis* presented concentrations of CLA almost



three fold higher (2.7 and 2.9 fold, respectively), in comparison to the control yogurt samples without prebiotic addition (85). However, the addition of FOS did not cause significant variations in the yogurt samples without the addition of probiotic cultures, indicating that there is a synergistic effect, because the use of *L. acidophilus* and *B. animalis* led to a significant increase in the concentration of CLA, when comparing with the control samples fermented only with yogurt starters (3.3 vs. 2.1 and 3.6 vs. 2.1 mg/g fat, respectively). During refrigerated storage, the CLA concentration of the yogurts remained constant after 28 days. Moreover, the supplementation (4% w/w) with other prebiotic compounds such as maltodextrin, oligofructose and polydextrose was reported to generate significant increases in the concentration of CLA in milks fermented with *S. thermophilus* and *L. acidophilus* (102). On the contrary, it has been reported that the addition of sucrose, fructose and lactose (60 g/L) led to an inhibition of the CLA production in yogurts elaborated using *L. acidophilus* and *L. delbrueckii* ssp. *bulgaricus* (103).

In one study evaluating the CLA production capacity of 155 different dairy starter cultures (*Lactococcus*, *Lactobacillus*, *Streptococcus* y *Bifidobacterium*), and 11 commercial yogurt starter cultures, it was found that 13 species from *Lactococcus* and seven from *Lactobacillus*, produced CLA in whole milk, in concentrations ranging between 4.3 and 11.2 mg/g fat (8). On the contrary, none of the commercial starter cultures produced CLA during the fermentation process.

## CONCLUDING REMARKS

In this work a comprehensive review on the concentration of CLA in milk, and on the influence of different technological processes normally applied to milk on the content of these bioactive compounds has been presented. Special emphasis was done in fermented milks. Taking into account the biological importance of the CLA-isomers, an increase in their intake would be beneficial to the human health. Likewise, the development of dairy products with high concentrations of CLA could have a great impact on the market of functional foods.

Because milk and dairy products are the foodstuffs providing the highest amounts of CLA in the human diet, various technological alternatives have been evaluated for increasing the concentration of CLA in these products in a natural manner. The manipulation of the animals' diet has allowed

increasing the content of CLA in milk, while the fermentation using microorganisms having linoleate isomerase activity has been employed as a promissory technological alternative for the manufacture of fermented milk products with high content of CLA. However, many factors can influence the concentration of CLA in milk and fermented milks. Although the mechanisms governing the biosynthesis of CLA during the milk fermentation are still unknown, it is evident that this reaction mainly depends on the type of strain. Various strains of food grade microorganisms such as *Bifidobacterium*, *Lactobacillus* and *Propionibacterium* have showed CLA production capacity from linoleic acid in milk-based culture mediums, but it is still unknown why some strains produce greater amounts than others.

The substrate composition is also an important factor in the production of CLA in fermented milks. The addition of fibers from fruits and prebiotics such as maltodextrine and FOS, has showed to be an effective method for enhancing the concentration of CLA in fermented milks manufactured using traditional yogurt starter cultures in combination with probiotic strains such as *L. acidophilus*, which could be an indicator of synergetic effects taking place in the production of CLA in this kind of dairy products. Likewise, the supplementation with linoleic acid has led to significant increases in the concentration of CLA in yogurt and other fermented milks. However, the addition of free fatty acids can on one hand increase the production costs, and on the other hand, be restricted because of the toxicity that this kind of compounds may provide at high concentrations.

The use of permeabilized cells may be a low cost technological alternative for the production of CLA, because it overcomes the problems related to the purification of the linoleate isomerase enzyme from microorganisms, and to the low permeability of the linoleic acid into the microbial cells. In addition, these biocatalysts can be reused several times without noticeable loss of activity.

Finally, although several studies have reported increases in the concentration of CLA in milk and fermented milk products, they are fairly moderate, and the obtained levels of CLA are significantly lower than those recommended to achieve therapeutic effects. More research is needed on the characterization of the linoleate isomerase enzymes produced by microorganisms, and on the appro-

priate process conditions (*i.e.* type of strains and substrate supplementation) to produce fermented dairy products with high concentrations of CLA.

## ACKNOWLEDGEMENTS

Institute de Ciencia y Technology de Alimentos (ICTA) of the Universidad Nacional de Colombia Sede Bogotá is acknowledged for supporting this work. The author declares that declare there is no conflict of interests, with the results presented in this article.

## REFERENCIAS

- Mills S, Ross RP, Hill C, Fitzgerald GF, Stanton C. Milk intelligence: Mining milk for bioactive substances associated with human health. *International Dairy Journal*. 2011;21(6):377-401.
- Pereira PC. Milk nutritional composition and its role in human health. *Nutrition*. 2014;30(6):619-27.
- Stergiadis S, Leifert C, Seal CJ, Eyre MD, Steinshamn H, Butler G. Improving the fatty acid profile of winter milk from housed cows with contrasting feeding regimes by oilseed supplementation. *Food Chemistry*. 2014;164:293-300.
- Yuan G-F, Chen X-E, Li D. Conjugated linolenic acids and their bioactivities: a review. *Food & Function*. 2014;5(7):1360-8.
- Gutierrez L-F, Rosada L-M, Jiménez A. Chemical composition of Sacha Inchi (*Plukenetia volubilis* L.) seeds and characteristics of their lipid fraction. *Grasas y Aceites*. 2011;62(1):76-83.
- Martínez-Monteagudo SI, Saldaña MDA. Modeling the retention kinetics of conjugated linoleic acid during high-pressure sterilization of milk. *Food Research International*. 2014;62(0):169-76.
- Glover KE, Budge S, Rose M, Rupasinghe HPV, MacLaren L, Green-Johnson J, et al. Effect of feeding fresh forage and marine algae on the fatty acid composition and oxidation of milk and butter. *Journal of Dairy Science*. 2012;95(6):2797-809.
- Pandit A, Anand S, Kalscheur K, Hassan A. Production of conjugated linoleic acid by lactic acid bacteria in milk without any additional substrate. *International Journal of Dairy Technology*. 2012;65(4):603-8.
- Jones EL, Shingfield KJ, Kohen C, Jones AK, Lupoli B, Grandison AS, et al. Chemical, physical, and sensory properties of dairy products enriched with conjugated linoleic acid. *Journal of Dairy Science*. 2005;88(8):2923-37.
- Collomb M, Schmid A, Sieber R, Wechsler D, Ryhanen EL. Conjugated linoleic acids in milk fat: Variation and physiological effects. *International Dairy Journal*. 2006;16(11):1347-61.
- Bauman DE, Lock AL. Conjugated Linoleic Acid: Biosynthesis and Nutritional Significance. In: Fox PF, McSweeney PLH, editors. *Advanced Dairy Chemistry Volume 2: Lipids*: Springer; 2006.
- Parodi PW. Milk fat in human nutrition. *Australian Journal of Dairy Technology*. 2004;59(1):3-59.
- Kennedy A, Martinez K, Schmidt S, Mandrup S, LaPoint K, McIntosh M. Antiobesity mechanisms of action of conjugated linoleic acid. *Journal of Nutritional Biochemistry*. 2010;21(3):171-9.
- Whigham LD, Watras AC, Schoeller DA. Efficacy of conjugated linoleic acid for reducing fat mass: a meta-analysis in humans. *American Journal of Clinical Nutrition*. 2007;85(5):1203-11.
- Toomey S, Harhen B, Roche HM, Fitzgerald D, Belton O. Profound resolution of early atherosclerosis with conjugated linoleic acid. *Atherosclerosis*. 2006;187(1):40-9.
- Mitchell PL, McLeod RS. Conjugated linoleic acid and atherosclerosis: studies in animal models. *Biochemistry and Cell Biology-Biochimie Et Biologie Cellulaire*. 2008;86(4):293-301.
- Dilzer A, Park Y. Implication of Conjugated Linoleic Acid (CLA) in Human Health. *Critical Reviews in Food Science and Nutrition*. 2012;52(6):488-513.
- McCrorie TA, Keaveney EM, Wallace JMW, Binns N, Livingstone MBE. Human health effects of conjugated linoleic acid from milk and supplements. *Nutrition Research Reviews*. 2011;24(2):206-27.
- Evans NP, Misyak SA, Schmelz EM, Guri AJ, Hontecillas R, Bassaganya-Riera J. Conjugated Linoleic Acid Ameliorates Inflammation-Induced Colorectal Cancer in Mice through Activation of PPAR gamma. *Journal of Nutrition*. 2010;140(3):515-21.
- Ip C, Chin SF, Scimeca JA, Pariza MW. Mammary-cancer prevention by conjugated dienoic derivative of linoleic-acid. *Cancer Research*. 1991;51(22):6118-24.
- Ip C, Singh M, Thompson HJ, Scimeca JA. Conjugated Linoleic Acid Suppresses Mammary Carcinogenesis and Proliferative Activity of the Mammary Gland in the Rat. *Cancer Research*. 1994;54(5):1212-5.
- Knekt P, Jarvinen R, Seppanen R, Pukkala E, Aromaa A. Intake of dairy products and the risk of breast cancer. *Br J Cancer*. 1996;73(5):687-91.
- Martínez-Monteagudo SI, Saldaña MDA. Retention of bioactive lipids in heated milk: Experimental and modelling. *Food and Bioproducts Processing*. 2014(0).
- Gorissen L, Leroy F, De Vuyst L, De Smet S, Raes K. Bacterial production of conjugated linoleic and linolenic acid in foods: a technological challenge. *Critical Reviews in Food Science and Nutrition*. 2013 doi: 10.1080/10408398.2012.706243.
- Irmak S, Dunford NT, Gilliland SE, Banskaliava V, Eisenmenger M. Biocatalysis of linoleic acid to conjugated linoleic acid. *Lipids*. 2006;41(8):771-6.
- Griinari JM, Corl BA, Lacy SH, Chouinard PY, Nurmela KVV, Bauman DE. Conjugated linoleic acid is synthesized endogenously in lactating dairy cows by Delta(9)-desaturase. *Journal of Nutrition*. 2000;130(9):2285-91.
- Butler G, Nielsen JH, Slots T, Seal C, Eyre MD, Sanderson R, et al. Fatty acid and fat-soluble antioxidant concentrations in milk from high- and low-input conventional and organic systems: seasonal variation. *Journal of the Science of Food and Agriculture*. 2008;88(8):1431-41.
- Palmquist DL, Lock AL, Shingfield KJ, Bauman DE. Biosynthesis of Conjugated Linoleic Acid in Ruminants and Humans. In: Steve LT, editor. *Advances in Food and Nutrition Research*: Academic Press; 2005. p. 179-217.
- Rico JM, Moreno B, Pabón ML, Carulla JE. Composición de la grasa láctea de la sabana de Bogotá con énfasis en ácido ruménico - CLA cis-9, trans-11. *Revista Colombiana de Ciencias Pecuarias*. 2007;20(1):30-9.
- Martínez-Monteagudo SI, Gänzle MG, Saldaña MDA. High-pressure and temperature effects on the inactivation of *Bacillus amyloliquefaciens*, alkaline phosphatase and storage stability of conjugated linoleic acid in milk. *Innovative Food Science & Emerging Technologies*. 2014;http://dx.doi.org/10.1016/j.ifset.2014.05.003.
- Khanal RC, Dhiman T, Boman RL. Changes in fatty acid composition of milk from lactating dairy cows during transition to and from pasture. *Livestock Science*. 2008;114(2-3):164-75.
- Stanton C, Murphy J, McGrath E, Devery R. Animal Feeding Strategies for Conjugated Linoleic Acid Enrichment of Milk. In: William W. Christie J-LS, and Richard Adlof, editor. *Advances in Conjugated Linoleic Acid Research*. Champaign, IL, USA: AOCS Publishing; 2003. p. 123-45.
- Bell JA, Kennelly JJ. Conjugated linoleic acid enriched milk: A designer milk with potential. *Advances in Dairy Technology*. 2001;13:213-28.

34. Floris R, Dekker R, Slangen C, Ellen G. Influence of pasture feeding and stall feeding on CLA and other fatty acids in bovine milkfat. *Australian Journal of Dairy Technology*. 2006;6(1):13-20.
35. Pabón ML, Carulla JE. Compuestos lipídicos benéficos para la salud humana asociados a la nutrición animal. *Revista Colombiana de Ciencias Pecuarias*. 2008;21:136-45.
36. Stergiadis S, Leifert C, Seal CJ, Eyre MD, Nielsen JH, Larsen MK, et al. Effect of Feeding Intensity and Milking System on Nutritionally Relevant Milk Components in Dairy Farming Systems in the North East of England. *Journal of Agricultural and Food Chemistry*. 2012;60(29):7270-81.
37. Toral PG, Frutos P, Hervás G, Gómez-Cortés P, Juárez M, de la Fuente MA. Changes in milk fatty acid profile and animal performance in response to fish oil supplementation, alone or in combination with sunflower oil, in dairy ewes. *Journal of Dairy Science*. 2010;93(4):1604-15.
38. Glasser F, Ferlay A, Chilliard Y. Oilseed Lipid Supplements and Fatty Acid Composition of Cow Milk: A Meta-Analysis. *Journal of Dairy Science*. 2008;91(12):4687-703.
39. Schroeder GF, Gagliostro GA, Bargo F, Delahoy JE, Muller LD. Effects of fat supplementation on milk production and composition by dairy cows on pasture: a review. *Livestock Production Science*. 2004;86(1-3):1-18.
40. Chilliard Y, Martin C, Rouel J, Doreau M. Milk fatty acids in dairy cows fed whole crude linseed, extruded linseed, or linseed oil, and their relationship with methane output. *Journal of Dairy Science*. 2009;92(10):5199-211.
41. Fox PF, McSweeney PLH. *Advanced Dairy Chemistry Volume 2: Lipids*. 3rd ed. Springer, editor: P.F. Fox and P.L.H. McSweeney; 2006.
42. Aneja RP, Murthi TN. Conjugated linoleic acid contents of Indian curds and ghee. *Indian Journal of Dairy Science*. 1990;43:231-8.
43. Herzallah SM, Humeid MA, Al-Ismael KM. Effect of heating and processing methods of milk and dairy products on conjugated linoleic acid and trans fatty acid isomer content. *Journal of Dairy Science*. 2005;88(4):1301-10.
44. Costa EN, Lacerda ECQ, Santos SMS, Santos CMS, Franco M, Silva RR, et al. Action of successive heat treatments in bovine milk fatty acids. *Journal of the Brazilian Chemical Society*. 2011;22:2115-20.
45. Campbell W, Drake MA, Larick DK. The impact of fortification with conjugated linoleic acid (CLA) on the quality of fluid milk. *Journal of Dairy Science*. 2003;86(1):43-51.
46. Leite J, Lima E, Baptista J. Azorean bovine milk conjugated linoleic acid. Effect of green pasture diet, storage and processing temperature. *Lait*. 2007;87(2):167-79.
47. Santos-Junior OO, Pedrao MR, Dias LF, Paula LN, Coro FAG, De Souza NE. Fatty acid content of bovine milkfat from raw milk to yoghurt. *Am J Applied Sci*. 2012;9(8):1300-6.
48. Zengin G, Cakmak YS, Guler GO, Oguz E, Aktumsek A, Akin M. The effect of pasteurisation temperature on the CLA content and fatty acid composition of white pickled cheese. *International Journal of Dairy Technology*. 2011;64(4):509-16.
49. Rodríguez-Alcalá LM. *Lípidos bioactivos en productos lácteos: Estrategias para su incremento y efectos del procesado y la conservación [PhD Thesis]: Universidad Autónoma de Madrid; 2009.*
50. Martínez-Monteagudo SI. Kinetic studies of chemical reactions and quality changes in conjugated linoleic acid (CLA) enriched milk treated with high-pressure sterilization. [Doctoral dissertation]: University of Alberta.; 2013.
51. Destailats F, Angers P. Thermally induced formation of conjugated isomers of linoleic acid. *European Journal of Lipid Science and Technology*. 2005;107(3):167-72.
52. Rodríguez-Alcalá LM, Harte F, Fontecha J. Fatty acid profile and CLA isomers content of cow, ewe and goat milks processed by high pressure homogenization. *Innovative Food Science & Emerging Technologies*. 2009;10(1):32-6.
53. Martínez-Monteagudo SI, Saldaña MDA, Torres JA, Kennelly JJ. Effect of pressure-assisted thermal sterilization on conjugated linoleic acid (CLA) content in CLA-enriched milk. *Innovative Food Science & Emerging Technologies*. 2012;16(0):291-7.
54. Rodríguez-Alcalá LM, Fontecha J. Hot topic: Fatty acid and conjugated linoleic acid (CLA) isomer composition of commercial CLA-fortified dairy products: Evaluation after processing and storage. *Journal of Dairy Science*. 2007;90(5):2083-90.
55. Shantha NC, Ram LN, O'Leary J, Hicks CL, Decker EA. Conjugated linoleic-acid concentrations in dairy-products as affected by processing and storage. *Journal of Food Science*. 1995;60(4):695-&.
56. Bisig W, Eberhard P, Collomb M, Rehberger B. Influence of processing on the fatty acid composition and the content of conjugated linoleic acid in organic and conventional dairy products - a review. *Lait*. 2007;87(1):1-19.
57. Sieber R, Collomb M, Aeschlimann A, Jelen P, Eyer H. Impact of microbial cultures on conjugated linoleic acid in dairy products - A review. *International Dairy Journal*. 2004;14(1):1-15.
58. Andrade JC, Ascencao K, Gullon P, Henriques SMS, Pinto JMS, Rocha-Santos TAP, et al. Production of conjugated linoleic acid by food-grade bacteria: A review. *International Journal of Dairy Technology*. 2012;65(4):467-81.
59. Adamczak M, Borscheuer UT, Bednarski W. Properties and biotechnological methods to produce lipids containing conjugated linoleic acid. *European Journal of Lipid Science and Technology*. 2008;110(6):491-504.
60. O'Shea EF, Cotter PD, Stanton C, Ross RP, Hill C. Production of bioactive substances by intestinal bacteria as a basis for explaining probiotic mechanisms: Bacteriocins and conjugated linoleic acid. *International Journal of Food Microbiology*. 2012;152(3):189-205.
61. Choi NJ, Park HG, Kim YJ, Kim IH, Kang HS, Yoon CS, et al. Utilization of Monolinolein as a Substrate for Conjugated Linoleic Acid Production by *Bifidobacterium breve* LMC 520 of Human Neonatal Origin. *Journal of Agricultural and Food Chemistry*. 2008;56(22):10908-12.
62. Villar-Tajadura MA, Rodríguez-Alcalá LM, Martín V, Gómez de Segura A, Rodríguez JM, Requena T, et al. Production of Conjugated Linoleic and Conjugated  $\alpha$ -Linolenic Acid in a Reconstituted Skim Milk-Based Medium by Bifidobacterial Strains Isolated from Human Breast Milk. *BioMed Research International*. 2014;2014:6.
63. Gorissen L, Raes K, Weckx S, Dannenberger D, Leroy F, De Vuyst L, et al. Production of conjugated linoleic acid and conjugated linolenic acid isomers by *Bifidobacterium* species. *Appl Microbiol Biotechnol*. 2010;87(6):2257-66.
64. Coakley M, Ross RP, Nordgren M, Fitzgerald G, Devery R, Stanton C. Conjugated linoleic acid biosynthesis by human-derived *Bifidobacterium* species. *Journal of Applied Microbiology*. 2003;94(1):138-45.
65. Coakley M, Banni S, Johnson M, Mills S, Devery R, Fitzgerald G, et al. Inhibitory Effect of Conjugated  $\alpha$ -Linolenic Acid from *Bifidobacteria* of Intestinal Origin on SW480 Cancer Cells. *Lipids*. 2009;44(3):249-56.
66. Chung SH, Kim IH, Park HG, Kang HS, Yoon CS, Jeong HY, et al. Synthesis of conjugated linoleic acid by human-derived *Bifidobacterium breve* LMC 017: Utilization as a functional starter culture for milk fermentation. *Journal of Agricultural and Food Chemistry*. 2008;56(9):3311-6.
67. Van Nieuwenhove CP, Oliszewski R, González SN, Pérez Chaia AB. Conjugated linoleic acid conversion by dairy bacteria cultured in MRS broth and buffalo milk. *Letters in Applied Microbiology*. 2007;44(5):467-74.
68. Gorissen L, De Vuyst L, Raes K, De Smet S, Leroy F. Conjugated linoleic and linolenic acid production kinetics by bifidobacteria differ among strains. *International Journal of Food Microbiology*. 2012;155(3):234-40.

69. Yang B, Chen H, Gu Z, Tian F, Ross RP, Stanton C, et al. Synthesis of conjugated linoleic acid by the linoleate isomerase complex in food-derived lactobacilli. *Journal of Applied Microbiology*. 2014;117(2):430-9.
70. Lin TY, Lin CW, Lee CH. Conjugated linoleic acid concentration as affected by lactic cultures and added linoleic acid. *Food Chemistry*. 1999;67(1):1-5.
71. Kishino S, Ogawa J, Yokozeki K, Shimizu S. Linoleic Acid Isomerase in *Lactobacillus plantarum* AKU1009a Proved to Be a Multi-Component Enzyme System Requiring Oxidoreduction Cofactors. *Bioscience, Biotechnology, and Biochemistry*. 2011;75(2):318-22.
72. Kishino S, Takeuchi M, Park S-B, Hirata A, Kitamura N, Kunisawa J, et al. Polyunsaturated fatty acid saturation by gut lactic acid bacteria affecting host lipid composition. *Proceedings of the National Academy of Sciences*. 2013;110(44):17808-13.
73. Gorissen L, Weckx S, Vlaeminck B, Raes K, De Vuyst L, De Smet S, et al. Linoleate isomerase activity occurs in lactic acid bacteria strains and is affected by pH and temperature. *Journal of Applied Microbiology*. 2011;111(3):593-606.
74. Lin TY, Lin CW, Wang YJ. Linoleic acid isomerase activity in enzyme extracts from *Lactobacillus acidophilus* and *Propionibacterium freudenreichii* ssp *shermanii*. *Journal of Food Science*. 2002;67(4):1502-5.
75. Ewaschuk JB, Walker JW, Diaz H, Madsen KL. Bioproduction of conjugated linoleic acid by probiotic bacteria occurs in vitro and in vivo in mice. *Journal of Nutrition*. 2006;136(6):1483-7.
76. Puniya AK, Chaitanya S, Tyagi AK, Singh SDK. Conjugated linoleic acid producing potential of lactobacilli isolated from the rumen of cattle. *Journal of Industrial Microbiology & Biotechnology*. 2008;35(11):1223-8.
77. Wei M, Ding X-L, Xue Z-L, Zhao S-G. Production of conjugated linoleic acid by permeabilized *Lactobacillus acidophilus* cells. *Journal of Molecular Catalysis B: Enzymatic*. 2014;108(0):59-63.
78. Lee SO, Hong GW, Oh DK. Bioconversion of linoleic acid into conjugated linoleic acid by immobilized *Lactobacillus reuteri*. *Biotechnology Progress*. 2003;19(3):1081-4.
79. Kishino S, Ogawa J, Omura Y, Matsumura K, Shimizu S. Conjugated linoleic acid production from linoleic acid by lactic acid bacteria. *Journal of the American Oil Chemists Society*. 2002;79(2):159-63.
80. Jiang J, Bjorck L, Fonden R. Production of conjugated linoleic acid by dairy starter cultures. *Journal of Applied Microbiology*. 1998;85(1):95-102.
81. Abd El-Salam MH, El-Shafei K, Sharaf OM, Effat BA, Asem FM, El-Aasar M. Screening of some potentially probiotic lactic acid bacteria for their ability to synthesis conjugated linoleic acid. *International Journal of Dairy Technology*. 2010;63(1):62-9.
82. Gutierrez L-F, Martínez JC, Barón-Núñez MR. Conjugated linoleic acid (CLA) content and fatty acid composition of some commercial yogurts from Colombia. *Rev. Fac. Nacional de Agronomía Medellín*. 2010;63(2):5685-92.
83. Osorio JA, Ramirez C, Novoa CF, Gutierrez LF. Conjugated linoleic acid, fatty acid profile and process properties in kumis - Fermented milk consumed in Colombia. *Rev.Vitae*. 2011;18(2):144-52.
84. Boylston TD, Beitz DC. Conjugated linoleic acid and fatty acid composition of yogurt produced from milk of cows fed soy oil and conjugated linoleic acid. *Journal of Food Science*. 2002;67(5):1973-8.
85. Akalin AS, Tokusoglu O, Gonc S, Aycan S. Occurrence of conjugated linoleic acid in probiotic yoghurts supplemented with fructooligosaccharide. *International Dairy Journal*. 2007;17(9):1089-95.
86. Dave RI, Ramaswamy N, Baer RJ. Changes in fatty acid composition during yogurt processing and their effects on yogurt and probiotic bacteria in milk procured from cows fed different diets. *Australian Journal of Dairy Technology*. 2002;57(3):197-202.
87. Serafeimidou A, Zlatanov S, Kritikos G, Tourianis A. Change of fatty acid profile, including conjugated linoleic acid (CLA) content, during refrigerated storage of yogurt made of cow and sheep milk. *Journal of Food Composition and Analysis*. 2013;31(1):24-30.
88. Florence ACR, da Silva RC, Santo APD, Gioielli LA, Tamime AY, de Oliveira MN. Increased CLA content in organic milk fermented by bifidobacteria or yoghurt cultures. *Dairy Science & Technology*. 2009;89(6):541-53.
89. Gorissen L, Raes K, De Smet S, De Vuyst L, Leroy F. Microbial production of conjugated linoleic and linolenic acids in fermented foods: Technological bottlenecks. *European Journal of Lipid Science and Technology*. 2012;114(4):486-91.
90. Santo APD, Cartolano NS, Silva TF, Soares F, Gioielli LA, Perego P, et al. Fibers from fruit by-products enhance probiotic viability and fatty acid profile and increase CLA content in yoghurts. *International Journal of Food Microbiology*. 2012;154(3):135-44.
91. Yadav H, Jain S, Sinha PR. Production of free fatty acids and conjugated linoleic acid in probiotic dahi containing *Lactobacillus acidophilus* and *Lactobacillus casei* during fermentation and storage. *International Dairy Journal*. 2007;17(8):1006-10.
92. Serafeimidou A, Zlatanov S, Laskaridis K, Sagredos A. Chemical characteristics, fatty acid composition and conjugated linoleic acid (CLA) content of traditional Greek yogurts. *Food Chemistry*. 2012;134(4):1839-46.
93. Florence ACR, Oliveira RPS, Silva RC, Soares F, Gioielli LA, Oliveira MN. Organic milk improves *Bifidobacterium lactis* counts and bioactive fatty acids contents in fermented milk. *Lwt-Food Science and Technology*. 2012;49(1):89-95.
94. Florence ACR, Beal C, Silva RC, Bogsan CSB, Pilleggi A, Gioielli LA, et al. Fatty acid profile, trans-octadecenoic, alpha-linolenic and conjugated linoleic acid contents differing in certified organic and conventional probiotic fermented milks. *Food Chemistry*. 2012;135(4):2207-14.
95. Kim YJ, Liu RH. Increase of conjugated linoleic acid content in milk by fermentation with lactic acid bacteria. *Journal of Food Science*. 2002;67(5):1731-7.
96. Lin TY. Influence of lactic cultures, linoleic acid and fructooligosaccharides on conjugated linoleic acid concentration in non-fat set yogurt. *Australian Journal of Dairy Technology*. 2003;58(1):11-4.
97. Chin SF, Liu W, Storkson JM, Ha YL, Pariza MW. Dietary sources of conjugated dienoic isomers of linoleic acid, a newly recognized class of anticarcinogens. *Journal of Food Composition and Analysis*. 1992;5(3):185-97.
98. Xu S, Boylston TD, Glatz BA. Conjugated linoleic acid content and organoleptic attributes of fermented milk products produced with probiotic bacteria. *Journal of Agricultural and Food Chemistry*. 2005;53(23):9064-72.
99. Espírito Santo APd, Silva RC, Soares FASM, Anjos D, Gioielli LA, Oliveira MN. Açai pulp addition improves fatty acid profile and probiotic viability in yoghurt. *International Dairy Journal*. 2010;20(6):415-22.
100. Ekinci FY, Okur OD, Ertekin B, Guzel-Seydim Z. Effects of probiotic bacteria and oils on fatty acid profiles of cultured cream. *European Journal of Lipid Science and Technology*. 2008;110(3):216-24.
101. Xu S, Boylston TD, Glatz BA. Effect of inoculation level of *Lactobacillus rhamnosus* and yogurt cultures on conjugated linoleic acid content and quality attributes of fermented milk products. *Journal of Food Science*. 2006;71(4):C275-C80.
102. Oliveira RPS, Florence ACR, Silva RC, Perego P, Converti A, Gioielli LA, et al. Effect of different prebiotics on the fermentation kinetics, probiotic survival and fatty acids profiles in nonfat symbiotic fermented milk. *International Journal of Food Microbiology*. 2009;128(3):467-72.
103. Lin TY. Conjugated linoleic acid concentration as affected by lactic cultures and additives. *Food Chemistry*. 2000;69(1):27-31.