IN VITRO EFFECT OF TERPENOIDS - RICH EXTRACT OF *Momordica charantia* ON ALPHA GLUCOSIDASE ACTIVITY

EFECTO IN VITRO DEL EXTRACTO RICO EN TERPENOIDES DE Momordica charantia SOBRE LA ACTIVIDAD DE LA ALFA-GLUCOSIDASA

A. B. SALLAU¹, R. N. YAKUBU¹, S. M. ALIYU², A. SALIHU¹, B. Y. BONIFACE¹

Received: April 23 of 2018. Approved: November 29 of 2018.

ABSTRACT

Background: *Momordica charantia* is a vine, native to tropics and subtropics that has found use in herbal medicine. In Nigeria, the plant is locally used in the treatment of diabetes. **Objective:** To investigate the inhibitory effect of terpenoid-rich extract from *Momordica charantia* on α -glucosidase activity. **Method:** The inhibitory effect of the terpenoid-rich extract from *Momordica charantia* on the activity of alpha-glucosidase from *Saccharomyces cerevisiae* as well as the determination of the mode of inhibition were investigated using *in vitro* assay methods. **Results:** The results revealed that the terpenoid-rich extract had a high IC₅₀ of 1.60 mg/mL. The Lineweaver-Burk plot revealed that the extract exhibited partially mixed-type inhibition of α -glucosidase, further purification will be necessary on the extract so as to increase its potency as an inhibitor of α -glucosidase. The action of the extract on α -glucosidase provided an insight into the probable mechanism of action of *Momordica charantia* in reducing hyperglycemia.

Keywords: α-glucosidase, terpenoids, inhibitor, inhibition, hyperglycemia, extract.

RESUMEN

Antecedentes: *Momordica charantia* es una vid, nativa de las regiones tropicales y subtropicales que ha sido usada en la medicina herbal. En Nigeria, esta planta se ha usado localmente en el tratamiento de la diabetes. **Objetivo:** Investigar el efecto inhibitorio del extracto rico en terpenoides de *Momordica charantia* sobre la actividad de la α -glucosidasa. **Método:** El efecto inhibitorio del extracto rico en terpenoides de *Momordica charantia* sobre sobre la actividad de la α -glucosidasa y el modo de inhibición se evaluó con un ensayo *in vitro*. **Resultados:** Los resultados mostraron que el extracto rico en terpenoides tiene un alto IC₅₀ de 1,60 mg/mL. El gráfico Lineweaver-Burk reveló que el extracto presenta una inhibición combinada de la α -glucosidasa. **Conclusión:** De este estudio se infiere que aunque el extracto rico en terpenoides inhibió la α -glucosidasa, es necesario realizar una purificación con el fin de incrementar su potencia como inhibidor de la α -glucosidasa. La acción del extracto sobre la α -glucosidasa proporciona un entendimiento del mecanismo de acción probable de *Momordica charantia* en la reducción de la hiperglicemia.

Palabras clave: α-glucosidasa, terpenoides, inhibidor, inhibición, hiperglicemia, extracto.

¹ Department of Biochemistry, ABU Zaria.

² Department of Biochemistry, Umaru Musa Yar'adua University, Katsina

^{*} Autor de correspondencia: sallauabdullahi@gmail.com

INTRODUCTION

Type 2 diabetes is characterized by persistent hyperglycemia due to defects in insulin action, insulin secretion or both resulting in persistently elevated postprandial glucose (1). Postprandial hyperglycemia is a major risk factor for microand macrovascular complications associated with diabetes (2, 3) and so controlling postprandial plasma glucose level is critical in the early treatment of diabetes mellitus and in reducing chronic vascular complications (4). Oxidative stress in type 2 diabetics is triggered more by acute glucose fluctuations during the postprandial period, therefore therapy in type 2 diabetes should not only target mean glucose concentrations and hemoglobin A1c levels but should also target acute glucose swings (5, 6). It is therefore imperative to look into treatment strategies for type 2 diabetes that control to the barest minimum acute fluctuations in postprandial glucose and the best target for this is the inhibition of alpha-glucosidase.

Alpha-glucosidases anchored in the mucosal brush border of the small intestine catalyzes the last hydrolytic step of starch and sucrose (7). One of the strategies adopted in the treatment of diabetes mellitus involves inhibition of alpha-glucosidase in the gastrointestinal tract, which results in the retardation of intestinal glucose absorption and lowering of postprandial blood glucose levels (8).

Medicinal plants and their products continue to be important therapeutic aids for alleviating the ailments of humankind (9-12). Herbs for diabetes treatment are not new. Since ancient times, plants and plant extracts were used to combat diabetes. Many traditional medicines in use are derived from medicinal plants, minerals, and organic matter.

Momordica charantia (M. charantia), also known as Bitter melon ("Karela", Balsam pear, or Bitter gourd), is a popular plant used for the treating of diabetes and diabetic related conditions amongst the indigenous populations of Asia, South America, India, the Caribbean and East Africa (13, 14). *M. charantia* is one of the plants that have been investigated thoroughly for the treatment of diabetes (15). The traditional use is supported by modern scientific evidence of the beneficial function of *M. charantia* one of the most promising plants for diabetes today (13, 16). Investigation of the traditional uses of *M. charantia* in India revealed that it is widely used for lowering of blood glucose levels in patients with diabetes (17).

Various animal studies have repeatedly shown hypoglycemic effects of the seeds, fruit pulp, leaves and whole plant of *M. charantia* in normal animals (18-20). In particular, M. charantia improves glucose tolerance and suppresses postprandial hyperglycemia in rats (21-23). Amongst the main constituents of bitter melon which are responsible for the antidiabetic effects are the triterpenes, (24, 25). The structure of three cucurbitane triterpenoids was determined, as well as two other major compounds that were tested and shown to significantly lower blood glucose levels in diabetic mice (26) but the effect of terpenes from *M. charantia* on the activity of alpha-glucosidase has not been investigated. Working on this premise, it is therefore important to investigate the effect of terpenes from *M. charantia* on alpha-glucosidase. This will bring us a step closer to understanding how the compounds found in the plant work and what they act on to elicit their anti-diabetic activity, in this case, reduction of postprandial hyperglycemia.

MATERIALS AND METHODS

Alpha-glucosidase

Alpha-glucosidase was purchased from Sigma chemical company St Louis U.S.A.

Plant collection and preparation

The leaves of Mormodica charantia was collected from Zaria, Kaduna state and was authenticated at the Department of Botany, Faculty of Life Sciences, Ahmadu Bello University, Zaria and a voucher number (VN1139) was obtained. The collected plant sample was rinsed with clean water and air dried at room temperature. The dried samples were pulverized into powder using a laboratory mill, the powder obtained was used to prepare the extracts. Five hundred grams of the plant was weighed and extraction was carried out using the maceration technique. The solvent used for the extraction was methanol. The powdered plant was soaked in methanol for 48 hours at room temperature. The extract was then filtered with Whatman No. 1 filter papers and the filtrate was concentrated using a rotary evaporator. The obtained crude extract was stored in an air-tight bottle.

Determination of alpha-glucosidase inhibition

The effect of the plant extract on α -glucosidase activity was determined according to the method described by Kim et al. [27], with slight modifications. Alpha-glucosidase from Saccharomyces cerevisiae (g0660), REF: AA/Abiola/ vakubu/0105/07/13] was used for the assay. The substrate solution (5.0mM) para-nitrophenylα-D-glucopyranoside (pNPG) was prepared in 20mM phosphate buffer, pH 6.9. Using dimethyl sulphoxide (DMSO) to dissolve the extract, varying concentrations of the extract solution ranging from 0.2, 0.4, 0.6, 0.8 to 1.0 mg/mL was used for the assay. Then 50 μ L of α -glucosidase (1 U/ mL) was added to five tubes containing 20 μ L of the different concentrations (0.2, 0.4, 0.6, 0.8 and 1 mg/mL) of the extract. After which, 30 μ L of 5.0 mM (pNPG) was added to start the reaction. The reaction mixture was incubated at 37°C for 1 hour and stopped by adding 1 mL of 0.1 M sodium carbonate (Na₂CO₃). The α -glucosidase activity was determined by measuring the yellow colored para-nitrophenol released from pNPG at 400 nm using a spectrophotometer. The blank solution was prepared by adding the Na₂CO₃ to the reaction mixture before adding the enzyme. The results were expressed as a percentage of the negative control in which the extract was replaced with DMSO. All the reactions were conducted in triplicates.

Percentage inhibition was calculated as:

Concentrations of extracts resulting in 50% inhibition of enzyme activity (IC_{50}) was determined graphically using the concentration/inhibition plots [28].

Determination of the mode of inhibition of α -glucosidase

The mode of inhibition of the extract was determined according to the method described by Ali et al [29]. Briefly, 20 μ L of three different concentrations (0 mg/mL, 1.4 mg/mL, 2.9 mg/mL) of the extract was pre-incubated with 40 μ L of α -glucosidase solution for 10 mins at 25°C in three different sets of tubes. In another set of tubes, α -glucosidase was pre-incubated with 20 μ L of phosphate buffer (pH 6.9). Then 20 μ L of *p*NPG at increasing concentrations (0.625 - 5mM) was added to all the sets of reaction mixtures to start the reaction. The mixture was then incubated for 1hour at 37°C, and 0.8 mL 0.1 M of Na₂CO₃ was added to stop the reaction. The amount of reducing sugars

released was determined spectrophotometrically using a para-nitrophenol standard curve which was converted to reaction velocities. A double reciprocal plot (1/v versus 1/[S]) where V is reaction velocity and [S] is substrate concentration was plotted. The type (mode) of inhibition of the crude extract on α -glucosidase activity was determined by analysis of the double reciprocal (Lineweaver-Burk) plot using Michaelis-Menten kinetics.

RESULTS

Percentage inhibition of alpha-glucosidase by a terpenoid rich extract of *M. charantia*

From the Figure 1, it was observed that the inhibition of the enzyme by the extract (though weak) even at high concentrations of 1 mg/mL, was less than 50%. Thus, using the graph equation, the concentration of the extract that can inhibit 50% of the enzyme activity (IC₅₀) was calculated to be 1.45 mg/mL.



Figure 1. Changes in the percentage of inhibition of alpha-glucosidase activity using different concentrations of the terpenoid rich extract of *M. charantia*.

Lineweaver – Burk plot for the activity of alpha-glucosidase in the presence of varying concentrations of the terpenoid rich extract of M. charantia

The double reciprocal plot (Lineweaver-Burk plot) showing the activity of alpha-glucosidase in the presence of the extract was shown in Figure 2. This graph is necessary so as to be able to gain an insight into the kind of inhibition the extract exerts on the enzyme. Thus, the terpenoid rich extract of *M. charantia* is a partially mixed inhibitor of alpha-glucosidase since there is a change in both the K_m and the V_{max} of the enzyme in the presence of the extract (Figure 2).



Figure 2. The Lineweaver-Burk plot for the activity of alpha-glucosidase in the presence of the terpenoids-rich extract of *M. charantia*.

DISCUSSION

In order to effectively manage diabetes mellitus, there has to be a close monitoring of the levels of postprandial blood glucose and the most effective way to do this is by regulating the amount of glucose that enters the bloodstream after intake of a meal and an effective way to achieve this is through the inhibition of alpha-glucosidase. The available oral chemotherapeutic drugs available come with a range of uncomfortable side effects such as flatulence, abdominal pain, renal tumors, hepatic injury, acute hepatitis, abdominal fullness and diarrhea (30-31). As a result of this side effects and other constraints; patients now prefer to use locally available herbs to treat diabetes, especially those plant and plant parts that are already consumed as food or food condiments since these plants have been seen to have lower side effects and are quite affordable. Several plants have been reported to have alphaglucosidase inhibitory activity (32-35). Momordica charantia being one of such plants, also the fact that several types of research (13-17) have shown the potency of the plant in the management of diabetes, it, therefore, justifies the focus of this particular research.

One of the major characteristics of drugs used in the management of Type 2 diabetes is ability to inhibit carbohydrate digesting enzymes (36). The result of the *in vitro* investigation on the inhibition of alpha-glucosidase activity in the presence of the terpenoids-rich extract of *M. charantia* showed that a concentration of 1.45 mg/mL of the extract could inhibit 50% of the enzyme activity. This concentration seems to be high when compared to similar researches carried out on the same enzyme but using different plants (37-38). Now the implication of this is that there may be a need for further purification of the extract so as to eliminate all interfering substances and to give the active ingredient a better chance of inhibiting the enzyme. Moreover, this result is not suprising going by the various therapeutic attributes of *Momordica charantia* (39).

Though the result of the inhibition experiments showed that a high concentration of the extract was needed to inhibit the enzyme; investigations on the mode of inhibition indicated that the extract is a partial mixed-type inhibitor of the enzyme since there was a change in both the K_m and V_{max} of the enzyme in the presence of the extract. This result may be attributed to the presence of several interfering substances in the extract and a clearer picture of the mode of inhibition of this extract will also be seen upon further purification.

CONCLUSION

The results gotten from this study showed that the terpenoid rich extract of *M. charantia* is a potent inhibitor of alpha-glucosidase enzyme activity.

CONFLICTS OF INTEREST

The authors of this manuscript do not have any conflict of interest

AUTHORS CONTRIBUTION

A. B. Sallau – Responsible for designing the work, R. N. Yakubu – Responsible for part of the writing up, S. M Abdullahi – Responsible for part of the literature sourcing, A. Salihu – Responsible for proof reading the manuscript and B. Y. Boniface - Responsible for the laboratory experiments.

REFERENCES

- Parkin CG, Brooks N. ¿Is Postprandial Glucose Control Important?. ¿Is It Practical In Primary Care Settings?. Clin Diabetes. 2002; 20:71-76.
- Hanefeld M, Schmechel H, Julius U, Schwanebeck U. Determinants for coronary heart disease in non-insulindependent diabetes mellitus: lessons from the diabetes intervention study. Diabetes Res Clin Pract. 1996; 30(Suppl):67-70.
- Hanefeld M, Fischer S, Julius U, Schulze J, Schwanebeck U, Schmechel H, *et al.* Risk factors for myocardial infarction and death in newly detected NIDDM: the Diabetes Intervention Study, 11-year follow-up. Diabetología. 1996; 39:1577-1583.
- Ortiz-Andrade RR, García-Jiménez S, Castillo-España P, Ramírez-Ávila G, Villalobos-Molina R, Estrada-Soto S. Alpha-Glucosidase inhibitory activity of the methanolic extract from *Tournefortia hartwegiana*: an antihyperglycemic agent. J Ethnopharmacol. 2007; 109:48-53.
- Monnier L, Colette C. Contributions of fasting and postprandial glucose to hemoglobin A1c. Endocr Pract. 2006; 12(Suppl 1):42-46.
- 6. Sies H, Stahl W, Sevanian A. Nutritional, dietary and postprandial oxidative stress. J Nutr. 2005; 135:969-972.
- Nichols BL, Avery SE, Karnsakul W, Jahoor F, Sen P, Swallow DM, *et al.* Congenital maltase-glucoamylase deficiency associated with lactase and sucrase deficiencies. J Pediatr Gastroenterol Nutr. 2002; 35:573-579.
- Kwon YI, Apostolidis E, Shetty K. In vitro studies of eggplant (*Solanum melongena*) phenolics as inhibitors of key enzymes relevant for type 2 diabetes and hypertension. Bioresource Technol. 2008; 99: 2981-2988.
- Joseph B, Raj SJ. Phytopharmacological properties of *Ficus* racemosa Linn - An overview. Int J Pharm Sci Rev Res. 2010b; 3(2):134-138.
- Joseph B, Jini D, Ajisha SU. Phytochemical characterization of herbal drug formulation for arthritis. Res J Phytochem. 2012; 6(2):54-60.
- 11. Singh U, Singh S, Kochhar A. Therapeutic potential of antidiabetic neutraceuticals. Psychopharmacol. 2012; 2(1):144-169.
- Zohary D, Hopf M. Domestication of plants in the old world. Oxford: Oxford University Press: 2000. 122 p.
- Cefalu WT, Ye J, Wang ZQ. Efficacy of dietary supplementation with botanicals on carbohydrate metabolism in humans. Endocr Metab Immune Disord Drug Targets. 2008;8: 78-81. [PubMed]
- 14. Cousens G. There is a cure for diabetes: the tree of life 21-day program. California. North Atlantic Books: 2008. 191-192p.
- Hasan I, Khatoon S. Effect of *Momordica charantia* (bitter gourd) tablets in diabetes mellitus: Type 1 and Type 2. Prime Res Med (PROM). 2012; 2(2):72-74
- Bhushan MS, Rao CHV, Ojha SK, Vijayakumar M, Verma A. An analytical review of plants for anti-diabetic activity with their phytoconstituent and mechanism of action. IJPSR. 2010; 1(1):29-46.

- Paul A, Raychaudhuri SS. Medicinal uses and molecular identification of two *Momordica charantia* varieties - a review. E J Bio. 2010; 6(2):43-51
- Jeong J, Lee S, Hue J, Lee K, Nam SY, Yun YW, et al. Effect of bitter melon (*Momordica Charantia*) on antidiabetic activity in C57BL/6J Db/DB mice. Korean J Vet Res. 2008; 48(3):327-336.
- Ismail WFE, Abo-Ghanem II, Saleh RM. Some physiological effects of *Momordica charantia* and *Trigonella foenum*-graecum extract in diabetic rats as compared with Cidophage[®]. World Acad Sci Eng Technol. 2012; 64:1206-1214.
- Mohammady I, Elattar S, Mohammed S, Ewais M. An evaluation of anti-diabetic and anti-lipidemic properties of *Momordica charantia* (Bitter Melon) fruit extract in experimentally induced diabetes. Life Sci J. 2012; 9(2):363-374.
- Cummings E, Hundal HS, Wackerhage H, Hope M, Belle M, Adeghate E, *et al. Momordica charantia* fruit juice stimulates glucose and amino acid uptakes in L6 myotubes. Mol Cell Biochem. 2004; 261:99-104.
- Abdullah M, Zuki ABZ, Goh YM, Rezaeizadeh A, Noordin MM. The effects of *Momordica charantia* on the liver in streptozotocin-induced diabetes in neonatal rats. Afr J Biotechnol. 2010; 9(31): 5004-5012.
- 23. Uebanso T, Arai H, Taketani Y, Fukaya M, Yamamoto H, Mizuno A, *et al.* Extracts of *Momordica charantia* suppress postprandial hyperglycemia in rats. J Nutr Sci Vitaminol (Tokyo). 2007; 53(6):482-486.
- Saeed MK, Shahzadi I, Ahmad I, Ahmad R, Shahzad K, Ashraf M, et al. Nutritional analysis and antioxidant activity of bitter gourd (*Momordica charantia*) from Pakistan. Pharmacology online. 2010; 1:252-260.
- Budget P, Shotipruk A. Extraction of phenolic compounds from fruits of bitter melon (*Momordica charantia*) with subcritical water extraction and antioxidant activities of these extracts. Chiang Mai J Sci. 2008; 35(1): 123-130.
- Lee SY, Eom SH, Kim YK, Park NI, Park SU. Cucurbitanetype triterpenoids in *Momordica charantia* Linn. J Med Plants Res 2009; 3(13): 1264-1269.
- 27. Kim YM, Jeong YK, Wang MH, Lee WY, Rhee HI. Inhibitory effects of pine bark extract on alpha-glucosidase activity and postprandial hyperglycemia. Nutrition 2005; 21: 756-761
- Olaokun OO, McGaw LJ, Eloff JN, Naidoo V. Evaluation of the inhibition of carbohydrate-hydrolyzing enzymes, antioxidant activity and polyphenolic content of extracts of ten African *Ficus* species (Moraceae) used traditionally to treat diabetes. *BMC* Complement Altern Med. 2013; 13:94–103.
- Ali H, Houghton PJ, Soumyanath A. Alpha-amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to *Phyllanthus amarus*. J Ethnopharmacol. 2006; 107(3): 449-455.
- Meenakshi B, Smita SZ, Shobha YB, Ameeta R, Bimba NJ. Antidiabetic Indian Plants: A Good Source of Potent Amylase Inhibitor. Evid Based Complement Alternat Med. 2011; 2011: 1-6
- Suzuki Y, Sano M, Hayashida K, Ohsawa I, Ohta S, Fukuda K. Are the effects of alpha-glucosidase inhibitors on cardiovascular events related to elevated levels of hydrogen gas in the gastrointestinal tract? FEBS Letters. 2009; 583:2157-2159.
- 32. Aljohi A, Matou-Nasri S, Ahmed N. Antiglycation and antioxidant properties of *Momordica charantia*. PloS ONE. 2016;11:18e159985
- Flores-Bacanegra L, Gonzalez-Andrade M, Bye R, Linares E, Mata R. Alpha-glucosidase inhibitors from *Salvia circinata*. J. Nat Prod. 2017; 80(5): 1884-1593
- Verastegui-Omana B, Rebollar-Ramos D, Perez-Vasquez A, Martinez AL, Madariaga-Mazon A, Florez-Bocanegra L, et, al. Alpha-glucosidase inhibitors from *Malbranchae flovorosea*. J. Nat Prod. 2017; 80(1): 190-195

- Thao NP, Binh PT, Luyen NT, Hung M, Dang NH, Data NT. Alpha-amylase and alpha-glucosidase inhibitory activities of chemical constituents of *Wildelia chinesis* (Osbeck) Merr Leaves. J. Anal. Meth. Chem. 2018; 2018(5):1-8
- Chipiti T, Ibrahim MA, Singh M, Shahidullslam Md. In vitro alpha-amylase and alpha-glucosidase inhibitory effects and cytotoxic activity of *Albizia antunesiana* extracts. Pharmacogn Mag. 2015; 11(44): 231-236
- 37. Ali RB, Atangwho IJ, Kumar N, Ahmad M, Mahmud R, Asmawi MZ, *In vitro* and *in vivo* effects of standardized extract and fractions of *Phaleria macrocarpa* fruits pericarp on lead

carbohydrate digesting enzymes. BMC Complement Altern Med. 2013; 13:39.

- Mohamed EAH, Mohammad JAS, Lee FA, Sadikun A, Chan SH, et al. Potent α-glucosidase and α-amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from Orthosiphon stamineus Benth as an anti-diabetic mechanism. BMC Complement Altern Med. 2012; 12:176
- Saeed F, Afzaal M, Naiz B, Arshad MU, Tufail T, et., al. Bitter melon (Momordica charantia): a natural healthy vegetable. Int J Food Prop. 2018; 21(1):1270-1290.