

Evaluation of Cytotoxic effect of *Moringa peregrina* seeds on Oral Cancer, CAL 27 Cell Line and Red Blood Cells Hemolysis

Evaluación del efecto citotóxico de las semillas de *Moringa peregrina* en el cáncer oral, CAL 27, líneas celulares y hemólisis de glóbulos rojos

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Abstract

Background: *Moringa peregrina* Forssk is a well-known plant in ethnomedicine due to its widespread uses in various diseases like cough, wound healing, rhinitis, fever, and detoxification. The plant seeds contain compounds that are cytotoxic to many cancer cells. During the therapeutic use of plants via the oral route, some compounds present in the plants may be cytotoxic to normal cell lines and red blood cells.

Objective: This study was the first report of investigation of the cytotoxic profile on oral cancer, CAL 27, cell line, and hemolytic activities on human erythrocytes of *Moringa peregrina* seeds ethanolic extract (MPSE).

Methods: MPSE was screened for its cytotoxic effect against oral cancer, CAL 27, cell line using 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) assay. The toxicity of MPSE on human erythrocytes was determined by *in vitro* hemolytic assay.

Results: MPSE showed significant anti-proliferative activity against oral cancer, CAL 27 cell line at lower concentrations with half maximal inhibitory concentration (IC₅₀) value of 21.03 µg/mL. At 1,000 µg/ml of MPSE, the maximum hemolysis was found to be 14.3% which is within safer limit.

Conclusions: This study revealed a potential anti-oral cancer of MPSE and provided a baseline for its potential use in oral cancer treatment with minimum hemolytic effect on human RBCs.

Key words: CAL 27 cell line, *Moringa peregrina* seed extract, Erythrocyte hemolysis, Toxicity, Anticancer.

Resumen

Antecedentes: La *Moringa peregrina* Forssk es una planta muy conocida en etnomedicina debido a sus usos generalizados en diversas enfermedades como la tos, la cicatrización de heridas, la rinitis, la fiebre y la desintoxicación. Las semillas de la planta contienen compuestos citotóxicos para muchas células cancerosas. Durante el uso terapéutico de las plantas por vía oral, algunos compuestos presentes en ellas pueden ser citotóxicos para las líneas celulares normales y los glóbulos rojos.

Objetivo: Este estudio fue el primer informe de investigación del perfil citotóxico sobre el cáncer oral, CAL 27, línea celular, y las actividades hemolíticas en eritrocitos humanos del extracto etanólico de semillas de *Moringa peregrina* (MPSE).

Métodos: Se examinó el efecto citotóxico del MPSE contra la línea celular de cáncer oral CAL 27 mediante el ensayo con bromuro de 3-(4, 5-dimetiltiazol-2-il)-2, 5,-difeniltetrazolio (MTT). La toxicidad del MPSE sobre los eritrocitos humanos se determinó mediante un ensayo hemolítico in vitro.

Resultados: MPSE mostró una actividad antiproliferativa significativa contra el cáncer oral, línea celular CAL 27 a concentraciones más bajas con un valor de concentración inhibitoria media máxima (IC50) de 21,03 µg/mL. A 1.000 µg/ml de MPSE, la hemólisis máxima fue del 14,3%, lo que está dentro del límite de seguridad.

Conclusiones: Este estudio reveló un potencial anticancerígeno oral de MPSE y proporcionó una base para su uso potencial en el tratamiento del cáncer oral con un efecto hemolítico mínimo en los glóbulos rojos humanos.

Palabras clave: Línea celular CAL 27, Extracto de semilla de *Moringa peregrina*, Hemólisis de eritrocitos, Toxicidad, Anticáncer.

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Introduction

Cancer is one of the major diseases responsible for high mortality worldwide. Oral squamous cell carcinoma (OSCC) is the most common cancer in the oral cavity and ranked 6th and 10th commonest in males and females worldwide respectively (1, 2). In Iraq, studies showed a rapid increase in the incidence of OSCC (3). On clinical inspection by a dentist, it is comparatively easy to identify a tumor by visual examination or through some OSCC tumor markers, but due to ignorance by the patients at the early stage led to the progression of this tumor, and subsequently, OSCC would be diagnosed at the advanced stages which then lead to high mortality (4). Surgery and chemotherapy fail to cure or prevent the recurrence and metastasis of tumors and are often associated with side effects; thus, many researchers are focused on natural products to discover many new drugs for oral cancer treatment from some potential candidates (5, 6, 7). Natural products are well flourished for therapeutic actions preventing and treating different ailments (8-10). Many new anticancer therapies from natural products were reported in basics researches (11). In general preventive practice, cytotoxic and hemolytic properties of natural sources were screened to predict the possible toxic effects on mammalian cells before developing them into a therapeutic agent for future work (8, 9).

Moringa peregrina Forssk is a well-known, traditionally used plant grown in Africa and Asia (10). *M. peregrina* seeds contain highly pharmacologically active components such as oleic acid, linoleic acid, isothiocyanate, tocopherols, flavonoids, and phenolic compounds. These compounds are reported to have different biological activities like antidiabetic (11), anti-Herpes simplex virus (12), anti-inflammatory (13), antibacterial (14), and anticancer against various cell lines (CACO-2, MCF-7, HeLa, HepG2, and L929) (16).

Even though several pharmacological studies on *M. peregrina* seeds have been reported in the literature, the anti-proliferation effect of *M. peregrina* seeds against oral carcinoma cells and the evaluation of the hemolytic capacity towards erythrocytes have not been scientifically reported yet. Thus, this study was conducted to test cytotoxic activities on the CAL 27 cell line and the effect of MPSE on human erythrocytes.

Materials and methods

Preparation of plant

In 2013, *M. peregrina* seeds were collected and submitted at the Department of Botany, University of Khartoum, where Dr. Maha Kordofani, a resident botanist, authenticated these seeds. These *M. peregrina* seeds were air-dried for 5 days at 25-30°C, powdered, and macerated in 1:5 dried plant weights to ethanol volume ratio. The collected filtrate was dried under reduced pressure at 46 to 51°C using a rotary evaporator to obtain crude *M. peregrina* seed extract (MPSE) (9).

Cytotoxicity Analysis Using 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT) colorimetric assay

The MPSE was tested on the CAL 27 Cell lines to check the cytotoxic activity of this extract. CAL 27 Cell line was grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 1% penicillin/streptomycin, 10% FBS, and in a humidified 5% CO₂ chamber at 37°C; the cells were seeded at 5×10^3 cells/well into 96-well plates. The vehicle used to prepare the initial stock of MPSE was 0.1% dimethylsulfoxide (DMSO). After 24 h, these cells were treated with MPSE at different concentrations (12.5-100 µg/mL) and incubated for 48 h. After incubation, cell viability was measured by adding 20 µL of MTT dye (5 mg/mL) to all wells and further incubated for 4 h. The percentage of cytotoxicity of the extract was calculated by measuring the absorbance of each well at a wavelength of 540 nm using an ELISA plate reader (Tecan, California, USA) (15).

Hemolytic Activity

After getting approval from the Independent Ethics Committee of ICCBS, University of Karachi, with reference no ICCBS/IEC-047-HB-2019/Protocol/1.0, the hemolytic assay was carried out.

The hemolytic effect of MPSE was determined using the method described by Wiradharma et al., 2011⁽¹⁶⁾. Human blood (3 mL) withdrawn from a volunteer was collected in an EDTA-anticoagulated venoject tube and centrifuged at $700 \times g$, and the packed erythrocytes were separated from the plasma. The erythrocytes were washed thrice with 1X PBS and then diluted with PBS to obtain a 4% erythrocyte suspension. 100 μ L of erythrocyte suspension were placed in triplicates in the wells of the 96-well cell culture plates. The erythrocytes in the respective wells were treated with 250, 500, and 1000 μ g/mL of MPSE, and the plate was incubated at 37°C for 1 h to allow for hemolysis. The plate was then centrifuged at $800 \times g$ for 15 min. From each well, 100 μ L of supernatants were transferred to a new 96-well plate, and the absorbance was determined in a microplate reader (MultiSkanGo, Thermo Scientific) at 576 nm. Erythrocytes treated with Triton X-100 (0.5%) and PBS served as the positive and negative control, respectively. Erythrocytes in 2% DMSO were the solvent control.

Results

Effects of MPSE on CAL 27 Oral cancer cell line

MPSE inhibited *in vitro* proliferation of CAL 27 cell lines. At 100 μ g/mL, MPSE showed more than 80% inhibition (Figure 1). IC_{50} of MPSE was found to be 25 μ g/mL, significantly higher than doxorubicin $IC_{50} = 0.5 \mu$ g/mL⁽¹⁵⁾. The extract was active below 100 μ g/mL, demonstrating the presence of the potential compounds in the extract and showing anti-cancer activity.

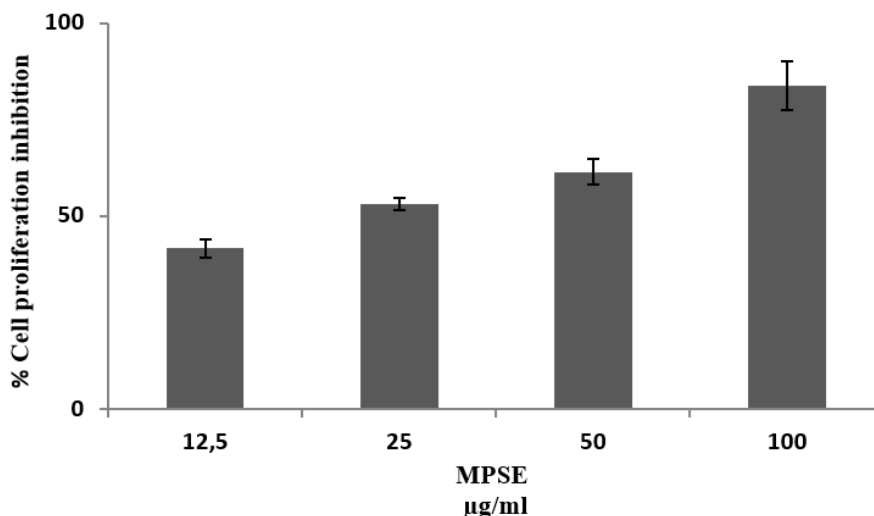


Figure 1. Cell proliferation inhibition (%) of the oral cancer cell line CAL 27 after treatment with *M. peregrina* seed extract (MPSE) for 48 h.

The hemolytic effect of MPSE on human erythrocytes

The hemolytic effect of MPSE augmented with increasing treatment concentrations. MPSE caused 14.3 % hemolysis at 1,000 $\mu\text{g/mL}$. This hemolysis was below 20%, demonstrating the non-hemolytic property of this extract. The positive (Triton X-100 (0.5%) control showed full hemolysis (100%), while the negative control (PBS-treated RBC cells) did not observe hemolysis (0%) (Figure 2).

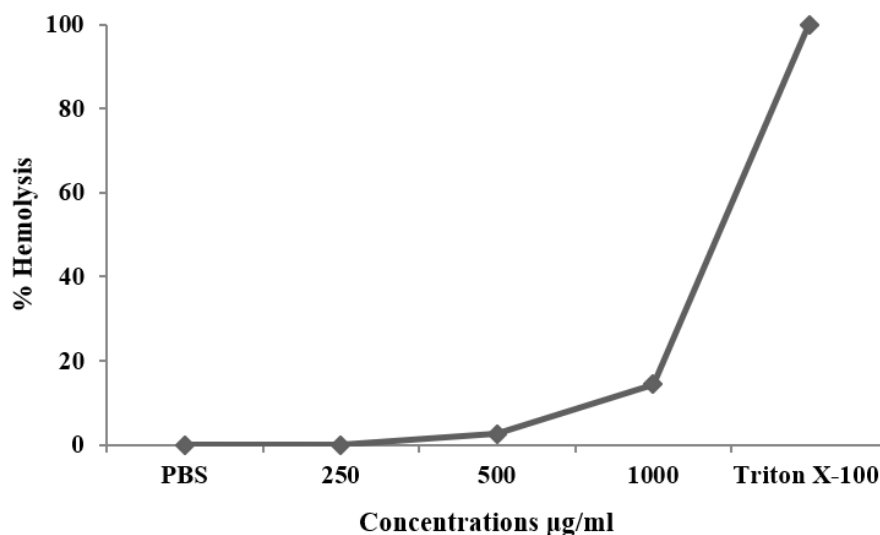


Figure 2. Hemolysis of human erythrocytes induced by *M. peregrina* seed extract (MPSE) after 1 h incubation.

Discussion

Since ancient times, people from distinct civilizations have used different plants from their surroundings to treat various disorders. Recently, the emergence of multidrug-resistant diseases, such as MDR pathogens, cancer, diabetes, and other inflammatory diseases, has motivated researchers to explore different natural sources, such as animals and plants, to find novel compounds to treat various conditions. To date, many antibacterial, anticancer compounds isolated from natural sources have been extensively used due to their effective therapeutic use ⁽¹⁷⁻²¹⁾. Recently, *M. peregrina* seed extracts have gained popularity due to their antioxidant ⁽¹¹⁾, anti-inflammatory ⁽¹³⁾, antibacterial ⁽¹⁴⁾, anticancer ⁽¹⁵⁾, antispasmodic ⁽¹⁴⁾ activities.

M. peregrina seed have been reported for its cytotoxic effect on CACO-2, HeLa, AU565, MCF-7, L929, HepG2, PC-3 cell lines ^(15, 19). Abou-Hashem *et al.* 2019 ⁽¹⁹⁾ reported the chloroform fraction of the ethanolic extract of *M. peregrina* seed extract as the most active antitumor fraction. HPLC analysis of this chloroform fraction reported many polyphenols like vanillin, syringic acid, naringenin, ferulic acid, quercetin, coumaric acid, daidzein, and cinnamic acid. Syringic acid is reported to have an antimutagenic effect on human colorectal

cancer cells (20). Coumaric acid (23) and vanillin (24) also inhibit colon cancer cells by inhibiting the cell cycle and inducing apoptosis. Naringenin and ferulic acid inhibit gastric cancer cells and osteosarcoma cells through apoptosis by downregulating the protein kinase (AKT) pathway (25, 26). Daidzein induced apoptosis and cell cycle arrest in human ovarian cancer cells (27). Cinnamic acid inhibits human melanoma cells through apoptosis by disrupting the cytoskeleton (28). This fraction also contains flavonoids like quercetin which interacts with DNA and activates the mitochondrial pathway of apoptosis in leukemia cancer cells (29). All these phenolic and flavonoid compounds were reported to have an anticancer effect on different cancer cell lines; therefore, these compounds may act synergistically on different targets of the complex cellular pathway and induce cytotoxicity toward cancer cells. These compounds induce apoptosis by activating various apoptotic pathways like upregulating caspase3 with the release of cytochrome c, downregulating anti-apoptotic genes (Bcl-2, Bcl-xL), cell cycle arrest, premature aging, activating mitochondrial apoptosis pathway, by enhancing the immune system to destroy cancer cells, reducing proliferation, angiogenesis, differentiation and metastasis of cancers (30-32).

Although many anticancer compounds had potent anticancer activity against various cancer cell lines, they failed to advance in clinical trials because they were cytotoxic to normal host cells and blood cells. Some were highly hemolytic, limiting them to topical use. Therefore, it is necessary to verify *in vitro* the hemolytic effect of *M. peregrina* on human erythrocytes since most anticancer agents damage blood cells, causing anemia and myelosuppression (33). Any compounds, formulations, or extracts with less than 10% hemolysis are non-hemolytic, and those with more than 25% are at risk of hemolysis (34).

Therefore, for the suitability of any compounds, formulation, or extracts to treat diseases, hemolysis must be less than 25%. In the present study, MPSE showed a maximum of 14.3% hemolysis when treated at 1,000 µg/mL, which is below 25%. Thus, this extract showed non-hemolytic activity and can be used for oral consumption for treating various diseases.

Conclusion

MPSE is toxic to oral cancer, CAL 27 cells, with a low hemolytic property; thus, this seed possesses hemocompatibility capacity to be used as a therapeutic agent.

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References

1. Abid AM, Merza MS. Immunohistochemical expression of Cyclin D1 and NF-KB p65 in oral lichen planus and oral squamous cell carcinoma (Comparative study). J Baghdad Coll Dent. 2014;26:80-7. <https://jbcd.uobaghdad.edu.iq/index.php/jbcd/article/view/301>.

2. Tsai LL, Yu CC, Chang YC, Yu CH, Chou MY. Markedly increased Oct4 and Nanog expression correlates with cisplatin resistance in oral squamous cell carcinoma. *J. Oral Pathol Med.* 201;40(8): 621-628. DOI: <https://doi.org/10.1111/j.1600-0714.2011.01015.x>
3. Fuoad SAA, Mohammad DN, Hamied MAS, Garib BT. Oro-facial malignancy in north of Iraq: a retrospective study of biopsied cases. *BMC Oral Health.* 2021;21(1):1-10. DOI: <https://doi.org/10.1186/s12903-021-01521-3>.
4. Usman S, Jamal A, Teh MT, Waseem A. Major Molecular Signaling Pathways in Oral Cancer Associated with Therapeutic Resistance. *Front Oral Health.* 2021;1. DOI: <https://doi.org/10.3389/froh.2020.603160>.
5. Ali M, Abbas A, Drea A. Green synthesis of nano binary oxide SiO₂/V₂O₅ NPs integrated ointment cream application on wound dressings and skin cancer cells. *Baghdad Sci J.* 2022;20(3):0734. DOI: <https://doi.org/10.21123/bsj.2022.7318>.
6. Nazhvani AD, Sarafraz N, Askari F, Heidari F, Razmkhah M.. Anti-cancer effects of traditional medicinal herbs on oral squamous cell carcinoma. *Asian Pac J Cancer Prev.* 2020;21(2):479. DOI: <https://doi.org/10.31557/APJCP.2020.21.2.479>.
7. Cardona-Mendoza A, Olivares-Niño G, Díaz-Báez D, Lafaurie GI, Perdomo SJ. Chemopreventive and Anti-tumor Potential of Natural Products in Oral Cancer. *Nutr Cancer.* 2022;74(3):779-795. DOI: <https://doi.org/10.1080/01635581.2021.1931698>.
8. Al-Bahrani RM, Radif HM, Albaayit SFA. Evaluation of potent silver nanoparticles production from *Agaricus bisporus* against *Helicobacter pylori*. *Pak J Agric Sci.* 2020;57(4):1197-1201. DOI: <https://doi.org/10.21162/PAKJAS/20.9893>.
9. Albaayit SFA, Rasedee A, Abdullah N. Zerumbone-loaded nanostructured lipid carrier gel facilitates wound healing in rats. *Rev. bras. farmacogn.* 2020;30(2):272-278. DOI: <https://doi.org/10.1007/s43450-020-00023-7>.
10. Albaayit SFA, Rasedee A, Abdullah N, Abba Y. Methanolic extract of *Clausena excavata* promotes wound healing via antiinflammatory and anti-apoptotic activities. *Asian PacJ Trop Biomed.* 2020;10(5):232-238. DOI: <https://doi.org/10.4103/2221-1691.281467>.
11. Albaayit SFA. *In vitro* evaluation of anticancer activity of Moringa peregrina seeds on breast cancer cells. *Eurasia J Math Sci Technol Educ.* 2020;11:163-166. <http://www.epstem.net/en/download/article-file/1439710>.
12. Albaayit SFA, Maharjan R, Khan M. Evaluation of hemolysis activity of Zerumbone on RBCs and brine shrimp toxicity. *Baghdad Sci J.* 2021;18(1):65-69. DOI: <https://doi.org/10.21123/bsj.2021.18.1.0065>.
13. Albaayit SFA, Ozaslan M. Cytotoxic and urease inhibition potential of *Moringa peregrina* seed ethanolic extract. *Int J Pharm.* 2019; 15(1): 151-155. DOI: <https://doi.org/10.3923/ijp.2019.151.155>.
14. Abd Rani NZ, Husain K, Kumolosasi E. Moringa genus: a review of phytochemistry and pharmacology. *Front Pharmacol.* 2018;9:108. DOI: <https://doi.org/10.3389/fphar.2018.00108>.
15. Soltan MM, Zaki AK. Antiviral screening of forty-two Egyptian medicinal plants. *J ethnopharmacol.* 2009;126(1):102-107. DOI: <https://doi.org/10.1016/j.jep.2009.08.001>
16. Albaayit SFA, Al-Khafaji ASK, Alnaimy HS. In vitro macrophage nitric oxide and interleukin-1 beta suppression by Moringa peregrina seed. *Turk J Pharm Sci.* 2019; 16(3): 362-365. DOI: <https://doi.org/10.4274/tjps.galenos.2018.52244>
17. Albaayit SFA, Khan M, Abdullah R. Zerumbone induces growth inhibition of Burkitt's lymphoma cell line via apoptosis. *Nat Volatiles Essent Oils.* 2021;8(3):56-63. <https://doi.org/10.37929/nveo.927770>

18. Albaayit SFA, Maharjan R. Immunomodulation of Zerumbone via Decreasing the Production of Reactive Oxygen Species from Immune Cells. *Pak J Biol Sci.* 2018;21(9):475-479. DOI: <https://doi.org/10.3923/pjbs.2018.475.479>.
19. Albaayit SFA. Enzyme inhibitory properties of zerumbone. *Pak J Agri Sci.* 2021;58(3):1207-1209. DOI: <https://doi.org/10.21162/PAKJAS/21.9759>.
20. Albaayit SFA. Evaluation of anti-methicillin resistant *Staphylococcus aureus* property of *Clausena excavata* leaves by using atomic force microscopy and flowcytometry techniques. *Pak J Agri Sci.* 2021;58(1):315-320. DOI: <https://doi.org/10.3389/froh.2020.603160>.
21. Albaayit SFA, Maharjan R, Abdullah R, Noor MHM. Anti-Enterococcus Faecalis, Cytotoxicity, Phytotoxicity, and Anticancer Studies on *Clausena excavata* Burum. f. (Rutaceae) Leaves. *BioMed Res Int.* 2021. DOI: <https://doi.org/10.1155/2021/3123476>.
22. Albaayit SFA, Mohammed MK. Antiangiogenic activity and ROS-mediated lung cancer cell line injury of zerumbone. *J Fac Pharm Ankara.* 2023;47(3). DOI: <https://doi.org/10.33483/jfpau.1112778>.
23. Wiradharma N, Khoe U, Hauser CA, Seow SV, Zhang S, Yang YY. Synthetic cationic amphiphilic α -helical peptides as antimicrobial agents. *Biomaterials.* 2011;32(8):2204-2212. DOI: <https://doi.org/10.1016/j.biomaterials.2010.11.054>.
24. Fang L, Gao L, Xie L, Xiao G. Eukaryotic translation initiation factor 5A-2 involves in doxorubicin-induced epithelial-mesenchymal transition in oral squamous cell carcinoma cells. *J Cancer.* 2018;9(19):3479-3488. DOI: <https://doi.org/10.7150/jca.26136>.
25. Sadraei H, Asghari G, Farahnaki F. Assessment of hydroalcoholic extract of seeds and leaves of *Moringa peregrina* on ileum spasm. *Res Pharm Sci.* 2015;10(3):252-258. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4621632/pdf/RPS-10-252.pdf>
26. Abou-Hashem MM, Abo-Elmatty DM, Mesbah NM, Abd EL-Mawgoud AM. Induction of sub-G0 arrest and apoptosis by seed extract of *Moringa peregrina* (Forssk.) Fiori in cervical and prostate cancer cell lines. *J Integr Med.* 2019;17(6):410-422. DOI: <https://doi.org/10.1016/j.joim.2019.09.004>.
27. Abaza MS, Al-Attiyah RA, Bhardwaj R, Abbadi G, Koyippally M, Afzal M. Syringic acid from *Tamarix aucheriana* possesses antimitogenic and chemo-sensitizing activities in human colorectal cancer cells. *Pharm. Biol.* 2013;51(9):1110-1124. DOI: <https://doi.org/10.3109/13880209.2013.781194>.
28. Jaganathan SK, Supriyanto E, Mandal M. Events associated with apoptotic effect of p-Coumaric acid in HCT-15 colon cancer cells. *World J Gastroenterol.* 2013;19(43):7726-7734. DOI: <https://doi.org/10.3748/wjg.v19.i43.7726>.
29. Maiyola F, Moodley R, Singh M. Phytochemistry, cytotoxicity and apoptosis studies of β -sitosterol-3- β -glucoside and β -amyrin from *Prunus africana*. *African Journal of Traditional. BMC Complement Altern Med.* 2016;13(4):105-112. DOI: <https://doi.org/10.21010/ajtcam.v13i4.15>.
30. Bao L, Liu F, Guo HB, Li Y, Tan BB, Zhang WX, Peng YH. Naringenin inhibits proliferation, migration, and invasion as well as induces apoptosis of gastric cancer SGC7901 cell line by downregulation of AKT pathway. *Tumour Biol.* 2016;37:11365-11374. DOI: <https://doi.org/10.1007/s13277-016-5013-2>.
31. Albaayit SFA, Khan M, Abdullah R, Noor MHM. Ethyl acetate extract of *Clausena excavata* induces growth inhibition of non-small-lung cancer, NCI-H460, cell line via apoptosis. *J Appl Biomed.* 2021;19(1):40-47. DOI: <https://doi.org/10.32725/jab.2021.007>

32. Wang T, Gong X, Jiang R, Li H, Du W, Kuang G. Ferulic acid inhibits proliferation and promotes apoptosis via blockage of PI3K/Akt pathway in osteosarcoma cell. *Am J Transl Res.* 2016;8(2):968-980. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4846940>.
33. Hua F, Li CH, Chen XG, Liu XP. Daidzein exerts anticancer activity towards SKOV3 human ovarian cancer cells by inducing apoptosis and cell cycle arrest, and inhibiting the Raf/MEK/ERK cascade. *Int J mol med.* 2018;41(6): 3485-3492. DOI: <https://doi.org/10.3892/ijmm.2018.3531>.
34. Niero EL, Machado-Santelli GM. Cinnamic acid induces apoptotic cell death and cytoskeleton disruption in human melanoma cells. *J Exp Clin. Cancer Res.* 2013;32(1):1-14. DOI: <https://doi.org/10.1186/1756-9966-32-31>.
35. Hashemzaei M, Delarami Far A, Yari A, Heravi RE, Tabrizian K, Taghdisi SM, Sadegh SE, Tsarouhas K, Kouretas D, Tzanakakis G, Nikitovic D. Anticancer and apoptosis-inducing effects of quercetin in vitro and in vivo. *Oncol Repo.* 2017;38(2): 819-828. DOI: <https://doi.org/10.3892/or.2017.5766>.